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(54) Title: VITAMIN B ₁₂ RECEPTOR MODULATING AGENTS AND METHODS RELATED THERETO		
(57) Abstract Vitamin B ₁₂ receptor modulating agents capable of modulating cell surface receptors by affecting the cell surface receptor trafficking pathway are disclosed. The vitamin B ₁₂ receptor modulating agents are comprised of a covalently bound rerouting moiety and targeting moiety linked by a water-solubilizing linker.		

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VITAMIN B₁₂ RECEPTOR MODULATING AGENTS AND METHODS RELATED THERETO

Field of the Invention

5 The present invention is generally directed to vitamin B₁₂ receptor modulating agents that bind TcII cell surface receptors and affect the receptor trafficking pathway and methods related thereto.

Background of the Invention

10 Cell surface receptors constitute a class of proteins that are responsible for receptor-mediated endocytosis of specific ligands. Basically, the receptors serve as escorts for ligand delivery to intracellular destinations.

Ligand delivery is generally achieved through coated regions on the plasma membrane called "coated pits." These pits continually invaginate and pinch off, forming "coated vesicles" in the cytoplasm. Coated pits and vesicles provide a pathway for receptor mediated endocytosis of specific ligands. The ligands that bind
15 to specific cell surface receptors are internalized via coated pits, enabling cells to ingest large numbers of specific ligands without taking in correspondingly large volume of extracellular fluid. The internalized coated vesicles may or may not lose their coats and bind with other vesicles to form larger vesicles called "endosomes." In the endosome the ligand and the receptor are separated or "sorted." Endosomes that
20 sort ligands and receptors are known as "compartment of uncoupling of receptor and ligand" or "CURL."

Endosomes may fuse with primary lysosomes, where their contents are digested, or they may be delivered to other intracellular destinations. The receptor proteins are generally not digested, but are rather recycled to the cell membrane surface through a process called "exocytosis," or transferred to early or late endosomes via multivesicular bodies. The entire pathway is referred to as the "receptor trafficking pathway."

Some receptors deliver their ligand directly to the cytoplasm or other specific intracellular locations. Perhaps one of the most studied receptor trafficking pathways is that of iron transport. In this pathway, a serum carrier protein, transferrin, binds iron and transports it to transferrin receptors on the plasma membrane surface. After binding and internalization, via coated pits, the resulting vesicle combines first with early endosomes and then with late endosomes. This process results in the gradual drop in pH in the vesicle. The drop in pH causes the transferrin carrier protein to lose its affinity to iron. When this occurs, the iron translocates through the membrane of the vesicle and joins the intracellular pool of enzymes. The transferrin receptor may then recycle to the cell surface where it may repeat the process.

Other receptors may deliver their ligand directly to the lysosomes for digestion. For example, the epidermal growth factor ("EGF") receptor delivers its ligand directly to a lysosome for degradation (Prog. Histochem. Cytochem. 26:39-48, 1992). The EGF receptor may recycle to the cell surface depending on its state of phosphorylation (Cancer Treat. Rep. 61:139-160, 1992; J. Cell. Biol. 116:321-330, 1992).

A single receptor may utilize more than one receptor trafficking pathway within the same cell. For example in polarized cells, such as specialized transport epithelial cells, membrane trafficking is distinct between apical and basal sides of the cell (Sem. Cell. Biol. 2:387-396, 1991). Moreover, non-polarized epithelial cells may simultaneously follow two separate sorting pathways.

The control or regulation of cell surface receptors may be achieved by a variety of techniques. Regulation of cell surface receptors may be accomplished, at a very basic level, by the binding of naturally occurring ligands. As discussed above, receptor binding of a ligand will generally trigger the internalization of the ligand-receptor complex. Such internalization may desensitize the cell to further ligand binding. (J. Immunol. 150:3161-9, 1993; Mol. Endocrinol. 6:2090-102, 1992; J. Cell. Physiol. 154:281-8, 1993; Receptor 1:13-32, 1990-91; Biochem. J. 288:55-61, 1992; J. Immunol. 148:2709-11, 1992; J. Cell. Physiol. 148:24-34, 1991). This type of

regulation, however, is transient in nature and does not result in diminution of biologic response.

Regulation of cell surface receptors may also be accomplished by administration of receptor antagonists or agonists. Receptor antagonists are organic protein or peptide ligands generally derived through empirical structure-function studies, or through the use of detailed knowledge of ligand and receptor interaction. Essentially, an antagonist may constitute any molecule with similar binding activity to a natural ligand, but incapable of producing the biological response normally induced by the natural ligand. Thus, the antagonist competitively blocks receptor activity. With a competitive antagonist, the regulation of receptor activity is dependent upon both the antagonist's affinity for the receptor, as well as its extracellular concentration over time. Receptor agonists are protein or peptide ligands derived in a similar manner as antagonists. Essentially, an agonist may constitute any molecule that binds to the receptor in a manner superior to that of the natural ligand.

One receptor of particular interest is the vitamin B₁₂ receptor. As has been demonstrated in experimental *in vitro* data, pre-clinical animal models, and patient studies, vitamin B₁₂ is a co-enzyme necessary in cell division, as well as cellular metabolism, in proliferating normal and neoplastic cells. Insufficient vitamin B₁₂ causes cellular division to be held in abeyance and ultimately may result in apoptosis. The nutrient is generally derived from dietary intake and is transported throughout the body complexed to transport proteins. The complex of transport protein and vitamin B₁₂ is recognized by a cellular receptor that internalizes the complex and releases the vitamin intracellularly. The overall process has been reviewed in *GUT* 31:59, 1991. Vitamin B₁₂ is taken in through the diet. Binding proteins in the saliva (R-binder) and gut (intrinsic factor-(IF)) complex vitamin B₁₂ after release from endogenous binding proteins by action of enzymes and low pH in the stomach. Vitamin B₁₂ is transferred across the intestinal epithelium in a receptor specific fashion to transcobalamin II (TcII). The vitamin B₁₂/transcobalamin II complex is then transported throughout the body and recognized by receptors present on dividing cells, internalized and released within the cell where it is utilized by certain enzymes as a co-factor.

The high affinity receptor in dividing tissues or cells responsible for internalization of vitamin B₁₂ recognizes transcobalamin II complexed with vitamin B₁₂. The vitamin B₁₂/TcII receptor recognizes only the vitamin B₁₂/TcII complex and not the serum transport protein or the vitamin alone. The receptor is undetectable on non-dividing cells; the mechanism for supplying non-dividing cells with vitamin B₁₂ is

poorly understood. However, it is known that more vitamin B₁₂ is required during cell division than during metabolism, and that the vitamin B₁₂/TcII receptor is the only high affinity means for cellular uptake of vitamin B₁₂ during cell division. When stimulated to divide, cells demonstrate transient expression of this receptor leading to vitamin B₁₂ uptake that precedes actual DNA synthesis (J. Lab. Clin. Med. 103:70, 1984). Vitamin B₁₂ receptor levels may be measured by binding of ⁵⁷Co-vitamin B₁₂ complexed to transcobalamin II (present in serum) on replicate cultures grown in chemically defined medium without serum. No receptor mediated uptake occurs in the absence of carrier protein.

Dividing cells, induced to differentiate, lose receptor expression and no longer take up vitamin B₁₂. More importantly, leukemic cells, deprived of vitamin B₁₂, will stop dividing and die (Acta Haemat. 81:61, 1989). In a typical experiment, leukemic cell cultures were deprived of serum for 3 days, and then supplemented either with serum (a source of vitamin B₁₂) or a non-metabolizable analogue of vitamin B₁₂ and cultured up to five days. Cell cultures supplemented with vitamin B₁₂ continued to grow, whereas those deprived of the active nutrient stopped growing and die.

Based on these observations, it has been suggested that whole body deprivation of vitamin B₁₂ may be useful in the treatment of cancer or other disorders characterized by uncontrolled growth of cells. Moreover, because of the critical role played by vitamin B₁₂-containing enzymes in cell division, it is believed that vitamin B₁₂ deprivation may be used in combination with chemotherapeutic drugs that inhibit cellular replication. For example, when vitamin B₁₂ depletion was combined with methotrexate, the two modalities together were more efficient in depleting folate levels in leukemic cells than either alone (FASEB J. 4:1450, 1990; Arch. Biochem. Biophys. 270:729, 1989; Leukemia Research 15:165, 1991). Folate are precursors in the production of DNA and proteins. In typical experiments, cultures of leukemic cells were exposed to nitrous oxide for several hours to convert the active form of endogenous vitamin B₁₂ to an inactive form. Replicate cultures were then left without further treatment, or additionally treated with methotrexate. Cellular folate levels were measured three days later. Cells treated with the combination (*i.e.*, both methotrexate and inactive vitamin B₁₂) showed a more striking decrease in cellular folate levels than with either of the two approaches alone. This combination also results in a higher cell kill *in vitro*. When this approach was applied to the treatment of highly aggressive leukemia/lymphoma in animal models (Am. J. Haematol. 34:128, 1990; Anticancer Res. 6:737, 1986; Cancer Chemother. Pharmacol. 17:114,

1986; Br. J. Cancer 50:793, 1984), additive or synergy of anti-tumor action was observed, resulting in prolonged remissions and cures.

A key finding in the experiments described above was that short-term (hours to days), whole body depletion of vitamin B₁₂ can act synergistically with
5 chemotherapeutic drugs (such as methotrexate and 5-FU) to inhibit tumor growth and treat animals with leukemia/lymphoma. Despite synergistic anti-tumor activity, there was no toxicity attributable to the short-term vitamin B₁₂ depletion for proliferating normal cells. This combination therapy was demonstrated in multiple animal models. Observations in patients have indicated that long-term (months to years) vitamin B₁₂
10 depletion is required to produce significant normal tissue toxicity. Even in those cases, subsequent infusion of vitamin B₁₂ can readily reverse symptomology (Br. J. Cancer 5:810, 1989).

Because of the promise of this therapeutic approach, various methods have been sought to efficiently and controllably perform a temporary depletion of vitamin
15 B₁₂. Such methods, however, affect all of the body's stores of vitamin B₁₂. They include dietary restriction, high doses of vitamin B₁₂ analogues (non-metabolizable-competitive antagonists that act as enzyme inhibitors), and nitrous oxide (transformation of vitamin B₁₂ to inactivate form). These different methods have been used in culture systems and in animals to deplete vitamin B₁₂. The most efficient and
20 the most utilized method has been the inhalation of nitrous oxide (laughing gas). Animals are maintained typically under an atmosphere of 50% to 70% of nitrous oxide for periods from a few hours to a few days, causing the conversion of endogenous vitamin B₁₂ into an inactive form. This methodology has been utilized in combination with drugs for therapy of leukemia/lymphoma. A further method for
25 vitamin B₁₂ depletion involves infusion of a non-metabolizable analogue of vitamin B₁₂ that essentially dilutes out the active form. This form of therapy is not specific for dividing cells but affects liver dependent metabolic processes. Another approach includes restricting the dietary intake of vitamin B₁₂. This method, however, requires very long periods of dietary restriction and is offset by hepatic storage of vitamin B₁₂.
30 All of these methods suffer from problems of specificity, since they affect both vitamin B₁₂-dependent growth as well as basal metabolism, and therefore are not particularly suited to the development of anti-proliferative pharmaceutical products.

In view of the biological importance of cell surface receptors, receptor-controlling agents have emerged as a class of pharmaceutical drugs. Moreover, with
35 the advent of genetic engineering for the isolation and amplification of genes for cell

surface receptors, as well as computer programs to model the interactions between ligands and receptors (*i.e.*, "rational" drug design), the production of receptor-controlling drugs has been significantly enhanced.

To date, many months or even years of scientific research, as well as
5 significant financial resources, are required to produce new receptor antagonists or agonists. To speed up this process, new screening technologies have been developed that utilize peptide or antibody recombinant libraries (*see, e.g.*, Gene 73:305, 1988; Proc. Nat. Acad. Sci. (USA) 87:6378, 1990; Biochromatography 5:22, 1990; Protein Engineering 3:641, 1989). While library screening does not require the same degree
10 of knowledge of a specific receptor/ligand system, it does involve an intensive screening effort utilizing functional receptor-specific assays. Moreover, the initial compounds identified by such screening programs are generally only precursors to the development of therapeutic products through more typical structure-functional assessments.

15 While antagonists and agonists are generally capable of regulating a biological response, the surface receptors that bind such ligands are continually being re-expressed on the cell surface. Thus, effective regulation by antagonists or agonists must rely on a relatively high and sustained serum concentration in order to bind the new surface receptors continually being expressed on the cell surface.

20 Accordingly, there is a need in the art for agents that bind cell surface receptors and thus regulate biological responses associated therewith, and that further effect normal cellular trafficking of the bound receptor. There is also a need in the art for agents that, when bound by a cell surface receptor and internalized, promote retention of the receptor within the cell. Moreover, there exists a need for methods
25 relating to the administration of such agents to regulate a biological response. The present invention fulfills these needs and provides further related advantages.

Summary of the Invention

Briefly stated, the present invention provides a vitamin B₁₂ receptor modulating agent, comprising a vitamin B₁₂ molecule coupled to a rerouting moiety
30 by a linker. The invention further provides vitamin B₁₂ receptor modulating agents wherein the linker is selected from a water-solubilizing linker or a non-water solubilizing linker and embodiments wherein the linker is covalently coupled to a vitamin B₁₂ coupling site selected from b-, d-, and e- coupling sites or the ribose 5'-OH coupling site.

These and other aspects of the present invention will become evident upon reference to the following detailed description and attached drawings. In addition, various references set forth below, that describe certain procedures or compositions in more detail, are incorporated by reference in their entirety.

5 Brief Description of the Drawings

The foregoing aspects and many of the attendant advantages of this invention will become more readily appreciated as the same becomes better understood by reference to the following detailed description, when taken in conjunction with the accompanying drawings, wherein:

FIGURE 1 is a schematic illustrating a mechanism of action of a receptor modulating agent of the present invention. A healthy receptor will internalize when bound by the appropriate ligand, release the ligand within the cell and then recycle to the cell surface. Receptor modulating agents of the present invention impede the receptor trafficking pathway by inhibiting the recycling of receptors to the cell surface. Essentially, the targeting moiety on receptor modulating agents binds the receptor and the rerouting moiety redirects the receptor/receptor modulating agent complex to other points within the cell, where it may be retained or degraded. (Not shown in this schematic are receptors synthesized *de novo*);

FIGURE 1 illustrates a formula representing a vitamin B₁₂ (cyanocobalamin) molecule and identifies a preferred coupling site suitable for use in the present invention for derivatization and conjugation;

FIGURE 2 is a schematic depicting a representative reaction scheme for the synthesis of a vitamin B₁₂-GABA adduct;

FIGURE 3A is a schematic depicting a representative reaction scheme for the
25 synthesis of a vitamin B₁₂ derivative comprising a vitamin B₁₂ molecule with a
diaminododecane linker arm coupled to any one of coupling sites *d*-, *e*-, or *b*-;

FIGURE 3B is a schematic depicting a representative reaction scheme for coupling a succinic anhydride to a vitamin B₁₂ diaminododecane adduct in preparation for coupling the adduct to a rerouting moiety, or other molecule, with an amino reaction site:

FIGURE 4 is a schematic depicting a representative reaction scheme for the synthesis of a vitamin B₁₂ derivative comprising a vitamin B₁₂ molecule and a diaminododecane linker arm coupled to a ribose coupling site;

FIGURE 5 is a schematic depicting a representative reaction scheme for
35 coupling vitamin B₁₂ or a vitamin B₁₂-GABA adduct to amikacin;

FIGURE 6 is a schematic depicting a representative reaction scheme for coupling vitamin B₁₂ or a vitamin B₁₂-GABA adduct to streptomycin;

FIGURE 7 is a schematic depicting a representative reaction scheme for coupling a vitamin B₁₂ carboxylate derivative or a vitamin B₁₂-GABA adduct to acridine;

FIGURE 8 is a schematic depicting a representative reaction scheme for the synthesis of a bivalent receptor modulating agent, a vitamin B₁₂ dimer, using a trifunctional linker. The trifunctional linker allows for coupling with additional compounds (*e.g.*, R-NH₂) such as, by way of example, aminoglucosides, aminoacridines, glycosylation inhibitors and biotin;

FIGURE 9 is a schematic depicting a representative reaction scheme for the synthesis of a vitamin B₁₂ dimer using a homobifunctional or homotrifunctional cross-linking reagent;

FIGURE 10 is a schematic depicting a representative reaction scheme for the synthesis of a vitamin B₁₂ dimer using a heterobifunctional cross-linker;

FIGURES 11 are schematics depicting representative reaction schemes for the synthesis of various receptor modulating agents generally comprised of a rerouting moiety, designated by the reactive group and R, and a vitamin B₁₂ molecule or derivative thereof as a targeting moiety;

FIGURE 12 is a graph illustrating the binding curve of Transcobalamin II to the cyanocobalamin monocarboxylic acids produced in Example 1. AD = cyanocobalamin (1); AL = cyanocobalamin *b*-monocarboxylic acid (2); AM = cyanocobalamin *e*-monocarboxylic acid (3); and AN= cyanocobalamin *d*-monocarboxylic acid (4);

FIGURE 13 is a graph illustrating the binding curve of Transcobalamin II to the cyanocobalamin diaminododecane adducts produced in Example 3 and 4. AH = cyanocobalamin *b*-monocarboxylic acid conjugate diaminododecane (7); AI = cyanocobalamin *e*-monocarboxylic acid conjugate diaminododecane (8); AJ = cyanocobalamin *d*-monocarboxylic acid conjugate diaminododecane (9); AK = cobalamin *e*-monocarboxylic acid conjugate diaminododecane, and AE = cyanocobalamin ribose-succinate (11);

FIGURE 14 is a graph illustrating the binding curve of transcobalamin II to a series of vitamin B₁₂ dimers. Dimer X = *b*-acid dimer with isophthaloyl dichloride (36); dimer Y = *e*-acid dimer with isophthaloyl dichloride (37); dimer Z = *d*-acid dimer with isophthaloyl dichloride (38); dimer A= *b*-acid dimer with *p*-iodo benzoyl

isophthaloyl dichloride (58); dimer B = *e*-acid dimer with *p*-iodo benzoyl isophthaloyl dichloride (59); and dimer C = *d*-acid dimer with *p*-iodo benzoyl isophthaloyl dichloride (60). These dimers were prepared as set forth in the Examples below (see Examples 13 and 16); and

- 5 FIGURE 15 is a graph illustrating the binding curve of transcobalamin II to a series of biotinylated vitamin B₁₂ molecules. AA = cyanocobalamin *b*-monocarboxylic acid conjugate diaminododecane and biotin (17); AB = cyanocobalamin *e*-monocarboxylic acid conjugate diaminododecane and biotin (18); AC = cyanocobalamin *d*-monocarboxylic acid conjugate diaminododecane and biotin (19);
- 10 AF = cyanocobalamin ribose-succinate conjugate diaminododecane (13); and AG = cyanocobalamin ribose-succinate conjugate diaminododecane and biotin (20). These biotinylated molecules were prepared as set forth in Examples below. (see Example 8.)

Detailed Description

- 15 The present invention is generally directed to a vitamin B₁₂ receptor modulating agent that is capable of binding to a vitamin B₁₂ cell surface receptor to form a receptor modulating agent/receptor complex ("agent/receptor complex"). The binding of a suitable receptor modulating agent to a cell surface receptor generally results in invagination of the agent/receptor complex into the cell into the vesicular
- 20 system in the same manner as the natural ligand. However, once internalized, or as part of the internalization process, a receptor modulating agent of the present invention affects the receptor trafficking pathway by effectively impeding, preventing, or delaying the receptor from recycling to the surface, thus depriving the cell of receptors able to engage in binding the cell's natural ligand and triggering related
- 25 biological responses.

- Within the context of the present invention, "affecting the receptor trafficking pathway" refers to impeding the receptor trafficking pathway in such a manner as to affect biological response. This would include trapping, delaying, retaining, redirecting, or degrading the cell surface receptor. A "receptor modulating agent" is
- 30 comprised of at least one targeting moiety covalently attached to at least one rerouting moiety. A "targeting moiety," as described in detail below, is a moiety capable of specifically binding to a vitamin B₁₂ cell surface receptor to yield an agent/receptor complex and, in a preferred embodiment, has an affinity for the cell surface receptor of within a hundredfold, and more preferably, within tenfold, of the
- 35 affinity of the natural ligand for the receptor. A preferred targeting moiety is a

vitamin B₁₂ molecule. In contrast, a "rerouting moiety" is a moiety that redirects an agent/receptor complex, resulting in prolonged retention, degradation, and/or modulation of the receptor within the interior of a cell or on the cell surface, including, by way of example, retaining the receptor in the cell membrane or directing
5 the receptor to a lysosome within the cell. Suitable rerouting moieties are described in detail below.

A targeting moiety is coupled to a rerouting moiety to yield the receptor modulating agent by any suitable means known in the art, including direct covalent linkage of an appropriate chemical linker or through a very tight association in non-
10 covalent attachment. By way of example for the latter, in one embodiment, coupling is accomplished through the combination of an avidin or streptavidin conjugate with a vitamin B₁₂/biotin conjugate. Coupling of the targeting moiety and the rerouting moiety should be of a nature that resists cleavage by the enzymatic and low pH conditions normally encountered within the internal portion of the cell, including
15 endosomes and lysosomes. Suitable linkers are noted below. The ability to resist cleavage may be detected by any means known in the art, including exposing the receptor modulating agent to enzymes at low pH and measuring release of the targeting or rerouting moiety using techniques known in the art.

Coupling of a targeting moiety and a rerouting moiety should not significantly
20 hinder the ability of the targeting moiety to specifically bind the cell surface receptor. The receptor modulating agent may also include additional moieties, so long as they do not interfere with either the targeting or the rerouting moieties. For example, such moieties may be coupled to the receptor modulating agent through the use of a trifunctional linker or they may be coupled to a rerouting or targeting moiety.
25 Optimal attachment of the two moieties may be determined by comparing the affinity of binding of the receptor modulating agent with free targeting moiety in assays of inhibition of binding.

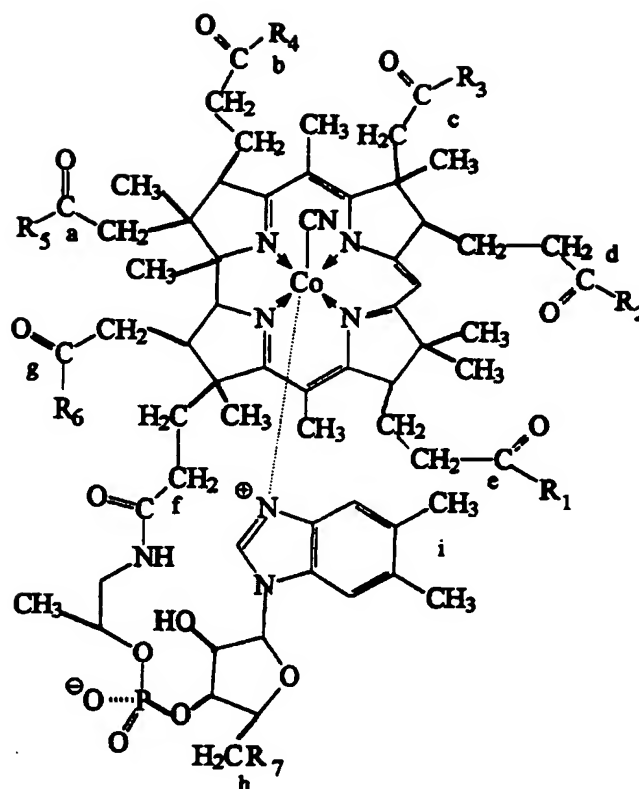
These, and other suitable techniques, are described in detail in Sambrook et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor, 1989.

Coupling of a targeting moiety and a rerouting moiety should also not
30 significantly affect the ability of the rerouting moiety to retain or delay the agent/receptor complex within the cell. This may be empirically determined by any one of several methods known in the art, including using labeling techniques to compare intracellular retention of the targeting moiety versus that of the receptor
35 modulating agent as exemplified below.

As noted above, targeting moieties of a receptor modulating agent include any moiety that specifically binds to a vitamin B₁₂ cell surface receptor. Suitable targeting moieties include a vitamin B₁₂ molecule. Vitamin B₁₂ is an essential nutrient for dividing cells. By inhibiting its uptake, the growth of dividing cells can be halted.

- 5 The cell surface receptor for vitamin B₁₂ is the transcobalamin II/vitamin B₁₂ ("TcII/B₁₂") receptor, that is characterized by a high affinity for the carrier protein, transcobalamin II (TcII), when complexed with vitamin B₁₂ ("TcII/B₁₂ complex"). The TcII/B₁₂ receptor does not recognize vitamin B₁₂ alone, but does recognize the carrier protein TcII with reduced affinity when not complexed with vitamin B₁₂. In
10 many respects, this receptor system is similar to that for transferrin/iron in that the goal of the receptor system is to deliver vitamin B₁₂ into cells such that it can be utilized by enzymes involved in DNA synthesis. Within the context of the present invention, the term "vitamin B₁₂" refers to the class of compounds known as cobalamins and derivatives thereof, including, by way of example, cyanocobalamin.
15 The term "vitamin B₁₂" is used interchangeably with the term cyanocobalamin.

- Suitable vitamin B₁₂ molecules include any vitamin B₁₂ capable of coupling to another molecule while maintaining its ability to form a TcII/B₁₂ complex. A preferred vitamin B₁₂ targeting moiety is generally comprised of a vitamin B₁₂ molecule, such as a cyanocobalamin, and a linker, described in detail below. The
20 linker may be coupled to any one of several sites on a vitamin B₁₂ molecule, including potential carboxyl coupling sites *a*- through *g*-, an alcohol (ribose) coupling site ("coupling site *h*"), or a benzimidazole coupling site ("coupling site *i*"). (See structure I below.) Preferably, a linker is coupled to coupling sites *b*-, *d*- or *e*- on a vitamin B₁₂ molecule. Even more preferably, a linker is coupled to coupling site *d*- or *e*-. This
25 embodiment of the present invention includes compounds represented by the following formula:



STRUCTURE I

wherein at least one of R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , and R_7 is a linker. One of ordinary skill in the art will appreciate that a number of other coupling sites on the vitamin B_{12} molecule may be chemically altered without affecting coupling of the molecule with a linker or TcII. Coupling sites that are not occupied by a linker may have a variety of chemical moieties attached thereto, including an amino, secondary amino, tertiary amino, hydroxy, lower alkyl, lower alkoxy, alkoxyalkyl, alkoxyalkoxy, cycloalkylalkoxy, and thioalkyl groups.

In a preferred embodiment, R_1 , R_2 , or R_4 is a linker and the remaining R groups are $-NH_2$, with the exception of R_7 , that is preferably $-OH$. In an especially preferred embodiment, R_2 is a linker, R_1 , R_3 - R_6 are $-NH_2$, and R_7 is $-OH$.

In another preferred embodiment, R_7 is a linker and R_1 - R_6 are $-NH_2$.

Suitable linkers include any one of several linkers, preferably containing at least two coupling or reactive groups, allowing the linker to bind to both vitamin B_{12} and a rerouting moiety. In the context of the present invention, the terms "coupling group" and "reactive group" are used interchangeably. By way of example, a linker

may be homobifunctional, heterobifunctional, homotrifunctional, or heterotrifunctional. Homobifunctional agents may facilitate cross-linking, or dimerization of vitamin B₁₂ molecules in a single step, hence a coupling reaction using these agents should be performed with an excess of homobifunctional agents, unless
 5 dimerization is the desired result, as in the synthesis of dimers described in detail below.

Suitable homobifunctional agents include, but are not limited to: disuccinimidyl suberate (DSS)*; bis(sulfosuccinimidyl) suberate (BS³)*; disuccinimidyl suberate (DSS)*; bis(sulfosuccinimidyl) suberate (BS³)*;
 10 disuccinimidyl tartarate (DST)*; disulfosuccinimidyl tartarate (Sulfo-DST)*; bis[2-(succinimidooxycarbonyloxy)ethyl] sulfone (BSOCOES)*; bis[2-(sulfosuccinimidooxycarbonyloxy) ethyl]sulfone (Sulfo-BSOCOES)*; bismaleimidohexane (BMH)*; 1,5-Difluoro-2,4-dinitrobenzene (DFDNB)*; dimethyl adipimidate-2 HCl (DMA)*; dimethyl pimelimidate-2 HCl (DMP)*; dimethyl
 15 subevimidate-2 HCl (DMS)*; isophthaloyl dichloride** (*Pierce Chemical, Co., Rockford, Illinois, **Aldrich Chemical Co., Milwaukee, Wisconsin)

Heterobifunctional agents facilitate cross-linking in a stepwise method, allowing more than one linker to be incorporated and a variety of targeting agents, such as vitamin B₁₂ molecules, to be linked. Suitable heterobifunctional agents
 20 include, but are not limited to:

N-succinimidyl-3-(2-pyridyldithio)propionate (SPDP)*; succinimidyl 6[3(2-pyridyldithio) propionamido] hexanoate (LC-SPDP)*; sulfosuccinimidyl 6-[3-(2-pyridyldithio) propionamido] hexanoate (Sulfo-LC-SPDP)*; succinimidyl 4-(N-maleimidomethyl)cyclohexane-1-carboxylate (SMCC)*; sulfosuccinimidyl 4-(N-maleimidomethyl)cyclohexane-1-carboxylate (Sulfo-SMCC)*; m-maleimidobenzoyl-
 25 N-hydroxysuccinimide ester (MBS)*; m-maleimidobenzoyl-N-hydroxysulfosuccinimide ester (Sulfo-MBS)*; N-succinimidyl(4-iodoacetyl)aminobenzoate (SIAB)*; sulfosuccinimidyl(4-iodoacetyl)aminobenzoate (Sulfo-SIAB)*; succinimidyl-4-(p-maleimidophenyl)butyrate (SMPB)*;
 30 sulfosuccinimidyl-4-(p-maleimidophenyl)butyrate (Sulfo-SMPB)* (*Pierce Chemical, Co., Rockford, Illinois)

Homo- and heterotrifunctional linkers are coupled to a rerouting moiety and a vitamin B₁₂ molecule as described above, with the additional advantage of a third coupling site on the linker. One of ordinary skill in the art will appreciate that this
 35 allows for any number of different molecules to couple with the rerouting moiety,

including, by way of example, markers, such as radiolabeled and fluorescent molecules; proteins and peptides, such as antibodies; and conjugating molecules, such as biotin. Suitable trifunctional linkers include, but are not limited to:

5 5-N-Boc amino isophthoyl ditetrafluorophenyl ester; 3,5-diamino methyl benzoate; 5-(*p*-iodobenzoyl)amino-1,3-isophthaloyl ditetrafluorophenyl ester; 5-(*p*-tri-N-butylisomylbenzoyl)-amino-1,3-isophthaloyl ditetrafluorophenyl ester; 4-(4-iodo(3-amidoethyl)-4'-2-(1,3-bistosyl) propylphthalate; 4-(4-tri-n-butylstannyl(3-amidoethyl)-4'-2-(1,3-bistosyl) propyl phthalate. (*Aldrich Chemical Co., Milwaukee, Wisconsin)

10 Homobifunctional, heterobifunctional, homotrifunctional, and heterotrifunctional linkers are commercially available.

The preferred length of a linker is dependent upon a number of empirical factors based upon the nature of the receptor modulating agent, including its component targeting and rerouting moieties. In general, a linker should have a length
15 sufficient such that the targeting moiety and the rerouting moiety of the receptor modulating agent may perform their designed functions free from steric inhibition. There are three primary areas of function that must be taken into consideration: (1) binding of the rerouting moiety to the targeting moiety, (2) binding of other molecules, such as TcII on the targeted receptor, to the receptor modulating agent, and (3) ability to interfere with the receptor trafficking. By way of example, a linker
20 should have a length sufficient to facilitate the specific binding of a targeting moiety to a cell surface receptor to yield an agent/receptor complex. Additionally, a linker should also have a length sufficient to permit a rerouting moiety to redirect an agent/receptor complex so as to interfere with the receptor trafficking pathway. Thus, empirical factors such as the size (*e.g.*, molecular weight and molecular conformation) and the nature (*e.g.*, charge and constituency) of receptor modulating agents, linkers, targeting moieties, cell surface receptors, rerouting moieties, and the receptor trafficking pathway will all affect the length of the linker.

By way of specific examples, linkers for vitamin B₁₂ receptor modulating
30 agents should have a length sufficient to allow for binding of a vitamin B₁₂ derivative to transcobalamin II to form a TcII/B₁₂ complex and, subsequently, to permit the binding of a TcII/B₁₂ complex to a TcII/B₁₂ cell surface receptor. Linkers for receptor modulating agents including a biotin moiety should be of a length sufficient to facilitate binding of the receptor modulating agent to avidin (or streptavidin).

Suitable linkers are generally relatively linear molecules greater than 4 atoms in length, typically between 6 and 50 atoms in length, and preferably are 8 to 35 atoms in length. In one preferred embodiment, the linker is a linear molecule of 12-15 atoms in length. In the context of the present invention, the term "atom" refers to a chemical
5 element such as, by way of example, C, N, O, or S. The ranges provided above are based on the relatively linear accounting of the linker. One of ordinary skill in the art will appreciate that a linker may be linear, branched, and even contain cyclical elements.

In another aspect of the present invention, the linker is a water-solubilizing
10 linker. The term "water-solubilizing linker" refers to any linker that, when covalently coupled to a rerouting and/or targeting moiety, increases the water solubility of either the components or the receptor modulating agent. The term "water solubility" refers to solubility in water or any other aqueous medium. In general, the solubility of a compound may be determined as described in "Handbook of Solubility Parameters
15 and Other Cohesion Parameters" by A.F.M. Benton, CRC Press, 1983. The water-solubilizing linkers may also enhance the water solubility of the receptor modulating agents.

The water-solubilizing linkers are composed of hydrophilic moieties (e.g., polar functional groups) including electronically neutral and charged (i.e., ionic)
20 moieties. Suitable hydrophilic moieties include electronically neutral moieties containing polar functional groups (i.e., groups that contain atoms of differing electronegativities such as organic compounds containing nitrogen, oxygen, and sulfur) that increase their hydrophilicity. Typically, these neutral hydrophilic moieties contain functional groups that hydrogen bond with water. Such hydrogen bonding
25 groups include ether (-O-), hydroxy (-OH), amino (-NR₂, -NHR, -NH₂), and to a lesser extent thioether (-S-), and thiol (-SH) groups.

Other polar functional groups that may serve as hydrophilic moieties include ethers and carbonyl containing groups such as acids, esters, amides, ketones, and aldehydes. Moieties that contain multiple polar functional groups are more
30 hydrophilic than those moieties that contain a single polar functional group. Suitable moieties that contain multiple polar groups include, by way of example, polyhydroxy, polyamino, polyether, polyphosphoric acid, polyalcohol and polyamine moieties. Polyhydroxy moieties include, by way of example, glycols, glycerols, and polysaccharides including glucose, fructose, galactose, idose, inositol, mannose,
35 tagatose, and N-methylglucamine. Polyalcohol moieties include, by way of example,

N-methylglucamine and glucose derivatives. Polyether moieties include, by way of example, polyethylene glycol, ethoxy ethanol, and ethoxy ethoxy ethanol. Polyamine moieties include, by way of example, spermine or spermidine.

Suitable charged hydrophilic moieties include those moieties that become
5 either formally negatively or positively charged in water. Suitable negatively charged moieties include acid anions resulting from the dissociation of acids in water. For example, carboxylic acids ($-\text{CO}_2\text{H}$) dissociate to form negatively charged carboxylate ions ($-\text{CO}_2^-$) at pH greater than about 5. Other stronger acids such as phosphoric ($-\text{PO}_3\text{H}_2$) and sulfonic ($-\text{SO}_3\text{H}$) acids ionize to form phosphonate ($-\text{PO}_3^{2-}$) and
10 sulfonate ($-\text{SO}_3^-$) anions, respectively, at pH greater than about 2. Other more weakly acidic moieties, such as phenols and thiols, may also dissociate to form their corresponding anionic derivatives that are also water solubilizing. Depending upon the pH of the aqueous solution, basic moieties may become formally positively charged moieties in water. These moieties become highly water soluble through
15 protonation in aqueous solution. For example, at pH about 5, amines ($-\text{NR}_2$, $-\text{NHR}$, $-\text{NH}_2$) become ammonium ions ($-\text{NHR}_2^+$, $-\text{NH}_2\text{R}^+$, $-\text{NH}_3^+$), all of that are highly water solubilizing moieties. Quaternary ammonium moieties ($-\text{NR}_3^+$) are extremely water-solubilizing at all pHs. Suitable charged solubilizing moieties also include polylysine groups.

20 The water solubility of a vitamin B_{12} derivative may be evaluated by any one of several means, including, by way of example, simply combining the derivative with an aqueous medium and observing the solubility at various temperatures. Alternatively, solubility may be ascertained by dissolving the derivative in water, stirring the solution, and allowing the solution to stand at room temperature for about
25 24 hours. The solution is then centrifuged and the resultant aqueous layer analyzed using high-pressure liquid chromatography ("HPLC"). The HPLC analysis was conducted isocratically using acetonitrile as the solvent on a LiChrospher 100, C-18 column ($5\text{ }\mu\text{M}$, $125 \times 4\text{ mm}$) using a flow rate of 2 mL/min .

The quantitation of a vitamin B_{12} containing solution may be accomplished by
30 HPLC using UV detection. In the quantitation, an aqueous solution of a vitamin B_{12} derivative is prepared and analyzed by HPLC as described above. A series of vitamin B_{12} aqueous solutions of known concentration are prepared and analyzed by HPLC. The results of these HPLC analyses are then used to construct a standard curve where the concentration of the vitamin B_{12} standard is plotted against the HPLC signal for
35 the standard. Once such a standard curve has been constructed, aqueous solutions of

various vitamin B₁₂ derivatives may be similarly analyzed and the concentration of the derivative in the solution determined.

Alternatively, the water solubility of a vitamin B₁₂ derivative (*i.e.*, the concentration of a vitamin B₁₂ derivative) may be determined directly by absorbance spectroscopy. Briefly, a known amount of vitamin B₁₂ is dissolved in a known amount of water to provide an aqueous solution of known concentration (*e.g.*, 10 mg derivative/10 mL water). The absorbance of this solution (or dilutions of the solution) is then measured by a UV absorbance spectrophotometer. The absorbance of the solution of known concentration provides the vitamin B₁₂ derivative's absorptivity. Once the vitamin B₁₂ derivative's absorptivity has been determined in this manner, the concentration (or the amount of the vitamin B₁₂ derivative in the solution) of subsequent aqueous solutions of the vitamin B₁₂ derivative may be determined by measuring the absorbance of the solution.

In a preferred embodiment, the water-solubilizing linker is a polyether or a polyhydroxy linker. In a particularly preferred embodiment, the water-solubilizing linker is the polyether linker such as a 4,7,10-trioxa-1,13-tridecanediamine linker or a 3,6-dioxa-1,8-octanediamine linker. The synthesis of a representative vitamin B₁₂ derivative having water-solubilizing linker is described in Example 23. The synthesis of a representative vitamin B₁₂-biotin conjugate having a water-solubilizing linker is described in Example 24. The syntheses of representative vitamin B₁₂ dimers having water-solubilizing linkers are described in Example 25. Coupling or reactive groups include any functional group capable of coupling a linker to a vitamin B₁₂ molecule. Suitable coupling groups include, nucleophilic and electrophilic functional groups. Suitable nucleophilic groups include hydroxy groups, amino groups, and thio groups. Suitable electrophilic groups include carboxylic acid groups and carboxylic acid derivatives including acid halides, acid anhydrides, and active esters, such as NHS esters.

Suitable homobifunctional linkers include, by way of example, diaminoalkanes, such as those represented by the formula $\text{NH}_2(\text{CH}_2)_x\text{NH}_2$, wherein $x = 2-20$. A preferred linker is a diaminododecane. Suitable heterobifunctional linkers include those represented by the formula $\text{NH}_2(\text{CH}_2)_y\text{COOH}$, wherein $y = 3-12$. Those of ordinary skill in the art will appreciate that a protecting group may be necessary when utilizing a heterobifunctional group.

A linker may be coupled to the preferred *b*-, *d*-, or *e*- coupling sites (see Structure I above) by any one of several suitable means, including, by way of

example, activating a vitamin B₁₂ molecule by hydrolyzing its propionamide groups to produce monocarboxylates, purifying the resulting monocarboxylates, and coupling a linker to a selected coupling site. Hydrolysis of the coupling sites may be accomplished by exposing vitamin B₁₂ to aqueous acid for a period of time and under
5 suitable conditions to hydrolyze the desired propionamide groups. Preferably, hydrolysis is performed by exposure of the amide to dilute aqueous acid for a period of about 6 to 12 days, typically about 9 to 11 days, and most preferably about 10 days at room temperature. Suitable aqueous acids include, by way of example, 0.1N hydrochloric acid, 0.5N phosphoric acid or 0.5N sulfuric acid.

10 Purification of *b*-, *d*-, and *e*- monocarboxylates can be accomplished by any one of several means, including column chromatography, such as gel-permeation chromatography, adsorption chromatography, partition chromatography, ion-exchange chromatography, and reverse-phase chromatography. Preferably, column chromatography is preparative reverse-phase liquid chromatography. These
15 techniques are described in detail in Lim, HPLC of Small Molecules, IRL Press, Washington, D.C., 1986. Purification of monocarboxylates by preparative liquid chromatography (LC) should be accomplished at a very slow flow rate. For example, LC purification may be conducted at a flow rate of 0.15 mL/min. on a 5 µm, 4.6 X 250 mm propylamine column (RAININ microsorb-MV amino column) eluting with
20 58 µM pyridine acetate, pH 4.4 in H₂O:THF (96:4) solution. Even more preferably, the coupling reaction is monitored using analytical high pressure liquid chromatography (HPLC). Reverse-phase HPLC chromatography is preferably carried out using an analytical version of above-noted propylamine column using a gradient solvent system at a flow rate of 1 mL/min. Within the context of the present
25 invention, the *d*- isomer is identified as the longest retained peak (third), the *e*- isomer is identified as the second retained peak, and the *b*- isomer is identified as the shortest retained peak (first) eluant from the LC column. The *e*-isomer may also be identified as that vitamin B₁₂ derivative demonstrating the greatest biological activity, as noted below.

30 A ribose coupling site (coupling site *h*, see structure I) may be activated by any one of several suitable means including activating a hydroxyl group at coupling site *h* by reaction with a suitable reagent (e.g., succinic anhydride) to yield a ribose derivative that bears a reactive group (e.g., a carboxylate group). The term "ribose coupling site" and "coupling site *h*" are used interchangeably. This technique is
35 described in detail in Toraya, Bioinorg. Chem. 4:245-255, 1975. Separation and

purification of the activated molecule may be accomplished on a C18 column as noted below. Once coupling site *h* has been activated, a linker may be coupled to this site in the same manner as described below.

Preferably, a 5'-OH ribose coupling site is activated using any one of several
5 suitable reagents including esterifying agents and ether forming reagents. These suitable reagents provide a cyanocobalamin derivative having a reactive group for further coupling reactions.

Esterification of the ribose 5'-OH may be accomplished with esterifying agents including, by way of example, carboxylic acid derivatives such as anhydrides, acid
10 halides, and reactive esters including TFP and NHS esters. For example, treatment with succinic anhydride provides a 5'-O-ribose ester derivative having a carboxylic acid group as a reactive group for subsequent coupling reactions. Similarly, treatment with a suitable N-protected reactive ester of 4-aminobutyric acid (GABA) yields a 5'-O-ribose ester derivative having, after N-deprotection, an amino group as a reactive
15 group for subsequent coupling reactions.

Suitable 5'-O-ribose ether derivatives may be prepared by any one of several methods including by activation of the 5'-OH followed by nucleophilic displacement. In one representative method, the 5'-OH group may be first converted to a good leaving group (*e.g.*, a *p*-toluenesulfonic acid group) followed by displacement with a
20 suitable nucleophile. Suitable nucleophiles include alcohol, amine, and sulfhydryl derivatives that produce 5'-O-ribose ether, amine, and thioether derivatives, respectively. Suitable alcohol, amine, and sulfhydryl derivatives include those derivatives having a reactive group, such as a carboxylic acid or amine group, protected as necessary, for subsequent coupling reactions. By way of example,
25 suitable ether forming reagents include N-protected alcohols, monoprotected diamines, and N-protected thioamines. Accordingly, depending upon the selection of the ether forming reagent, ether linkages including alkyl ether and benzyl ether linkages may be formed.

Suitable 5'-O-ribose ether derivatives may also be prepared by 5'-OH
30 alkylation with suitable alkylating agents. Suitable alkylating agents include alkylating agents having a reactive group, such as a carboxylic acid or amine group, protected as necessary, for subsequent coupling reactions. Preferred alkylating agents include active halide compounds such as haloacetates, benzyl halides, and silyl halides. Alkylation with a haloacetate such as methyl bromoacetate or trimethylsilyl
35 bromoacetate, or a benzyl halide such as methyl 4-(bromomethyl)benzoate provide

5' -O-ribose ethers (i.e., alkyl and ether linkages) having, after ester hydrolysis, a carboxylic acid group for subsequent coupling reactions. Alkylation with a silyl halide such as methyl 11-(chlorodimethylsilyl)undecanoate provides a 5'-O-ribose silyl ether (i.e., a silyl ether linkage) having, after hydrolysis, a carboxylic acid group for subsequent coupling reactions. The synthesis of a representative 5'-O-ribose ether derivative by alkylation with methyl bromoacetate is described in Example 26.

After activating the vitamin B₁₂ molecule at a selected coupling site, linkers may be coupled to a vitamin B₁₂ molecule to form a vitamin B₁₂ linker adduct using any one of several means, including, by way of example, an amide forming reaction, employing an amine group on the linker and a carboxylate coupling site on a vitamin B₁₂ molecule. Alternatively, a linker may be coupled to a vitamin B₁₂ molecule through an amide forming reaction, employing a carboxylate group on the linker and an amino group on a B₁₂ molecule. The amide forming reaction may include the use of a coupling agent. Suitable coupling agents include carbodiimide coupling agents, such as, by way of example, 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDC), 1-benzyl-3-(3-dimethylaminopropyl) carbodiimide (BDC), 1-cyclohexyl-3-(2-morpholinyl-4-ethyl)carbodiimide (CMC), and 1,3-dicyclohexylcarbodiimide (DCC). Preferably, the coupling agent is water soluble. Even more preferably, the coupling agent is EDC.

Alternatively, the amide forming reaction coupling the linker to a B₁₂ molecule may employ a reactive carboxylic acid group and an amine. Suitable reactive carboxylic acid groups include carboxylic acid derivatives that yield an amide upon reaction with an amine. Such reactive groups include, by way of example, any reactive carboxylic acid derivative, including, by way of example, carboxylic acid halides, such as acid chlorides and bromides; carboxylic acid anhydrides, such as acetic anhydrides and trifluoroacetic anhydrides; esters, such as p-nitrophenyl esters and N-hydroxysuccinimide esters. Such techniques are described in detail in Bodanszky, Principles of Peptide Synthesis, Springer Verlag, Berlin, 1984.

Although coupling of a linker through a cyano coupling site is possible, it is not preferred, due to the instability of linkers coupled to this site. Dolphin, D., [205] Methods Enzymol. 18C:34-52, 1971. Additionally, a linker may be coupled to a benzimidazole (coupling site *i*, see Structure I) using techniques described in detail in Jacobsen, Anal. Biochem. 113:164-171, 1981.

Vitamin B₁₂ linker adducts may be separated and purified using any suitable means, including column chromatography, such as gel-permeation chromatography,

adsorption chromatography, partition chromatography, ion-exchange chromatography, and reverse-phase chromatography. Preferably, column chromatography is preparative LC. These techniques are described in detail in Lim, HPLC of Small Molecules, IRL Press, Washington, D.C., 1986.

5 As noted above, the vitamin B₁₂ receptor modulating agents of the present invention must be capable of binding transcobalamin II. The ability of a receptor modulating agent to bind TcII may be ascertained using any one of several means known in the art, including competitive binding assays with the receptor modulating agent competing with native vitamin B₁₂.

10 Rerouting moieties of the present invention include any moiety that is capable of affecting the receptor trafficking pathway. This characteristic can be assessed by employing a receptor modulating agent having a radiolabeled targeting moiety and following its path through the cell. This is accomplished using techniques known in the art, including using radiolabeled, biotinylated, or FITC labeled targeting moiety
15 followed by binding assays, ELISA, or flow cytometry. A preferred receptor modulating agent is one that results in the removal of the highest percent of receptor for the longest period of time.

Suitable rerouting moieties of this invention do not significantly detract from the selectivity of the targeting moiety. Whether a rerouting moiety detracts from the
20 selectivity of a targeting moiety may be determined by any one of several methods known in the art, including comparing binding of the receptor modulating agent on receptor positive and receptor negative cells, as assessed by ELISA, flow cytometry, or other binding assays.

Rerouting moieties cause the retention/degradation of an agent/receptor
25 complex within at least one cell type, but not necessarily in all cells. In like fashion, a rerouting moiety causes retention of an agent/receptor complex in some cells, but not necessarily other agent/receptor complexes in other cells. Different rerouting moieties may also distinguish between receptor species, for example, as in polarized epithelium where the same receptor may independently traffic on the apical, basal, or basolateral
30 sides of the cell. To determine if a particular rerouting moiety is suitable, a rerouting moiety is covalently attached to the targeting moiety, and the resulting receptor modulating agent is compared for receptor modulation on different receptor-bearing cells using binding or functional assays known in the art.

Suitable rerouting moieties of this invention may be categorized into five
35 different functional classes: (1) lysosomotropic moieties; (2) intracellular

polymerizing moieties; (3) protein sorting signals or sequences; (4) conditional membrane binding peptides; and (5) bi- or multivalent receptor cross-linking moieties. While such rerouting moieties may have different functional mechanisms of action, all promote retention of the agent/receptor complex within the intracellular vesicular system. All of these classes of rerouting moieties will impart the ability to affect the receptor trafficking pathway.

In one aspect of the present invention, a first functional class of rerouting moieties, lysosomotropic moieties, are disclosed. Within the context of the present invention, the term "lysosomotropic moieties" refers to moieties that route the agent/receptor complex to the lysosomes. Numerous suitable lysosomotropic moieties are known, and are reviewed in Biochem. Pharmacol. 23:2495-2531, 1974.

A preferred lysosomotropic moiety includes an aminoglycoside antibiotic marked by the characteristic ability to accumulate in lysosomes after intracellular protonation. Intracellular protonation occurs in the increasingly acidic conditions that occur during the transfer from early to late endosomes and, finally, to the lysosome. Strong positive charges prohibit the lysosomotropic moiety from leaving the membrane-enclosed vesicles, thus trapping the agent/receptor complex in the vessel.

Aminoglycoside antibiotics are similar in structure, but are divided into structurally related families of compounds based upon the sugar units. These families include gentamycin, kanamycin, neomycin, and streptomycin. The gentamycin family includes gentamycin C₁, gentamycin C₂, gentamycin C_{1a}, sisomicin, and netilmicin; the kanamycin family includes kanamycin A, tobramycin, and amikacin; the neomycin family includes neomycin B, paromomycin, ribostamycin, and bytirosin B; and the streptomycin family includes streptomycin A and streptomycin B.

In a particularly preferred embodiment of the present invention, the rerouting moiety is gentamycin, that accumulates in lysosomes in concentration as much as three-hundredfold that of the extracellular concentration (J. Pharmacol. Exp. Ther. 255:867-74, 1990; Ren. Fail. 14:351-7, 1992).

Suitable aminoglycosides have reactive amine groups capable of being coupled through peptide or other chemical linkers. Thus, a targeting moiety may be readily attached via covalent linkage to these rerouting moieties using any one of several techniques known in the art to form covalent bonds, for example, using thioether, disulfide, ether, ester, and peptide bonds. Since many of the aminoglycoside antibiotics have several amines that could be derivatized in a conjugation procedure, a primary amine contained in these compounds can be selectively reacted to favor

covalently attachment to the targeting moiety through the 5' amine. With regard to streptomycin, covalent attachment to the targeting moiety may be accomplished by converting the aldehyde moiety to an amine, and attaching to the targeting moiety using carbodiimide or other suitable activated carboxylic acid. Aminoglycosides are
5 water soluble and do not readily bind to other proteins, and thus do not impart nonspecific binding to a receptor modulating agent.

Particularly preferred aminoglycosides include those that allow for preferential derivation of a selected amine. Specifically, preferred aminoglycosides include those compounds to that protective groups can be added to various nitrogen atoms thereof
10 and, subsequently, selectively deprotected to yield a single free amine. The free amine can be further derivatized, for example, by addition of a peptide linker or covalently attached directly to the targeting moiety. These rerouting moieties include ribostamycin, kanamycin, amikacin, and streptomycin. Ribostamycin is particularly preferred, due to its relative low toxicity and its derivatization chemistry, allowing an
15 acyl migration reaction to be effected on a hydroxyl protected ribostamycin to yield a single amine adduct. Kanamycin may also be used in a selective protection/acylation reaction; amikacin is commercially available in a form that allows attachment without deprotecting its amines or alcohol groups; and streptomycin can also be readily derivatized by protonating guanidinium groups under physiologic conditions to
20 provide the polycations necessary for cellular or lysosomal retention.

In another aspect of the present invention, nonaminoglycoside lysosomotropic compounds that may accumulate after intracellular protonation are also suitable rerouting moieties. Suitable nonaminoglycoside compounds exhibiting this characteristic are known in the art, a series of aminoacridine and amino quinoline
25 dyes, typified by cholquinine and quinacrine; a group of amino naphthalenes, typified by dansyl cadaverine; and derivatives thereof. Such dyes are characterized by cellular retention and low toxicity. All of these compounds have characteristic sites for covalent attachment to a targeting moiety via the amine and may be attached thereto as described above.

30 Another aspect of the present invention utilizes a lysosomotropic peptide subject to charge modification under intracellular conditions as a rerouting moiety. Once charge-modified, the rerouting peptide acts to retain an agent/receptor complex in the intracellular vesicular system until membrane flow delivers it to the lysosome for degradation. Preferably, these peptides are capable of being phosphorylated by

intracellular protein kinases. When phosphorylated by the intracellular enzymes, such peptides would be highly anionic.

Charge-based retention can be an inherent property of the rerouting peptide or can be imparted by intracellular modification. Intracellular modification may be accomplished by any of several means known in the art, including phosphorylation of certain residues of some receptors (*e.g.*, the EGF receptor) may cause intracellular rerouting (Cancer Treat. Res. 61:139-160, 1992; J. Cell. Biol. 116:321-30, 1992).

The rerouting peptides may be covalently attached to a targeting moiety by any means, including, for example, covalently linking the peptide directly to the targeting moiety, or by use of an appropriate linker moiety, such as G-G-G, that may be derivatized and covalently attached to the targeting moiety.

Preferred rerouting peptides include protein kinase-substrate peptides that incorporate serine. These peptides are particularly preferred for enhancement of receptor rerouting in tumor target cells, that have increased levels of protein kinase activity for serines or tyrosines. Increased levels of kinase activity within tumor cells may be attributed to the presence of oncogene products, such as H-ras, on the cytoplasmic side of tumor cell plasma membranes (C.I.B.A. Found. Symp. 164:208-18, 1992).

Suitable rerouting peptides also include protein kinase substrates and peptides that possess a single positive charge. The latter type of rerouting peptide may form an ion pair with a "glutamate-like" residue of an attached or closely associated residue(s) of the receptor. Particularly preferred rerouting peptides may be derived, using technologies known in the art, from the proteins and the amino acid sequences identified in Table 1.

TABLE 1 REROUTING PEPTIDES	
PEPTIDE SOURCE	AMINO ACID SEQUENCE
EGF receptor	DVVDAD EYLIPQ
EGF fragment	CMHIESLDSYTC
Phosphorylase kinase	RTKRSGSVYEPLKI
Protein kinase C pseudosubstrate	RFARK-GALRQKNV
Myelin basic protein	S/T-XAA-K/R (where XAA is an uncharged residue)
Kemptide	RGYALG or RGYSLG
Glycogen synthetase	PLSRTL SVAA
Transferrin receptor	FSLAR
III histone	ASGSFKL
Casein kinase II substrate	AAAAAASEEE or AAAAAASDDD
Insulin receptor auto-phosphorylation substrate	DIYETDYR
calmodulin-dependent protein kinase II	<u>Waxman and Arenowski Biochem. 32(11):2923-30, 1993</u>
Neurogranin	<u>Chen et al., Biochem. 32(4):1032-9, 1993</u>
MARCKS	<u>Heemskerk et al., Biochem. Biophys. Res. Commun. 190(1):236-41, 1993</u>
Glycogen synthase	<u>Marais et al., FEBS Letters 277:151-5, 1990</u>
Ribosomal protein S6	<u>Munro et al., Biochem. Biophys. Acta 1054:225-30, 1990</u>
Co-polymers that serve as substrates for protein kinase A, C, P	<u>Abdel-Ghony et al., Proc. Nat'l. Acad. Sci. 86:1761-5, 1989; Abdel-Ghony et al., Proc. Nat'l. Acad. Sci. 85:1408-11, 1988</u>
Serine-threonine kinases	<u>Abdel-Ghony et al., Proc. Nat'l. Acad. Sci. 86:1761-5, 1989; Abdel-Ghony et al., Proc. Nat'l. Acad. Sci. 85:1408-11, 1988</u>

In another aspect of the present invention, the rerouting moiety is a lysosomotropic amino acid ester that, in high concentration, can cause the lysis of granule-containing cells, such as NK cells, cytolytic T cells and monocytes. The concentration must generally be maintained below 100 mM to avoid lysis. Suitable lysosomotropic amino acid esters and their sources are presented in Table 2.

TABLE 2 LYSOSOMOTROPIC AMINO ACID ESTERS	
Leu-O-Me	<u>Res. Immunol.</u> 143:893-901, 1992 <u>Eur. J. Immunol.</u> 23:562-5, 1993 <u>Intl. Arch. Aller. & Immunol.</u> 100:56-59, 1993 <u>Cell. Immunol.</u> 139:281-91, 1992 <u>Exp. Pathol.</u> 42:121-7, 1991
Iso-leu-O-Me	<u>Res. Immunol.</u> 143:893-901, 1992
L-Val-O-Me	<u>J. Immunol.</u> 134:786-93, 1985
Phe-O-Me	<u>J. Immunol.</u> 148:3950-7, 1992 <u>Blood</u> 79:964-71, 1992
Phe-, Ala-, Met-, Trp-, Cys-, Try-, Asp-, & Glu-O-Me	<u>Int. J. Immunopharmacol.</u> 13:401-9, 1991

The lysosomotropic amino acid esters identified in Table 2 can be used to retain the agent/receptor complex in lysosomes after intracellular cleavage of the ester. In one embodiment, such amino acid esters may be utilized as the C-terminal portion of a larger peptide containing a linker sequence and/or a phosphorylation substrate sequence, and with suitable residues, such as cysteine, for covalent attachment to a targeting moiety, such as a sequence encoding a peptide or protein ligand for a given cell surface receptor.

In another embodiment of the present invention, a second functional class of rerouting moieties is disclosed. This class includes peptides that undergo polymerization within endosomes or lysosomes, inhibiting their passage through intracellular membranes.

Intracellular polymerizing compounds can be incorporated into a larger peptide containing the targeting moiety and a linker. Suitable peptides include the dipeptide ester referenced in Table 2 (*i.e.*, L-Leucyl-L-Leucine-O-Me). When transported into cells, these dipeptide esters preferentially accumulate in lysosomes and secondary granules of cytotoxic cells. These dipeptides also undergo self-association and polymerization, that result in trapping at low concentrations, and membrane rupture at higher concentrations.

TABLE 3 POLYMERIZING DI-PEPTIDE ESTER: L-LEUCYL-L-LEUCINE-O-ME	
<u>J. Invest. Dermat.</u> 99:805-825, 1992	
<u>J. Clin. Invest.</u> 84:1947-56, 1989	
<u>Transpl.</u> 53:1334-40, 1992	
<u>J. Immunol.</u> 138:51-7, 1987	
<u>J. Immunol.</u> 148:3950-7, 1992	
<u>J. Immunol.</u> 136:1038-48, 1986	
<u>Cryobiology</u> 29:165-74, 1992	
<u>Acta. Biochem Biophys. Hung</u> 24:299-311, 1989	
<u>Blood</u> 79:964-71, 1992	
<u>Blood</u> 78:2131-8, 1991	
<u>J. Immunol.</u> 139:2137-42, 1987	
<u>J. Exp. Med.</u> 172:183-194, 1990	
<u>J. Clin. Invest.</u> 78:1415-20, 1986	
<u>PNAS</u> 87:83-7, 1990	
<u>J. Immunol.</u> 137:1399-406, 1986	
<u>PNAS</u> 82:2468-72, 1985	

- Suitable intracellular polymerizing compounds also include peptides that can self-associate into alpha-helical structures termed "leucine zippers". In the context of this invention, such structures may be used to form intracellular polymers that are incapable of exiting intracellular vesicles. Such sequences can be selected by
- 5 observing self-association of the compounds in solution, and the formation of polymers capable of binding to DNA. Suitable peptide sequences that can self-associate into alpha-helical structures are presented in Table 4.

TABLE 4 LEUCINE ZIPPERS	
Boc(t-butoxycarbonyl)-Aib(alpha-aminoisobutyryl) Glu(OB _n l)-(benzoyl ester)-Leu-Aib-Ala-Leu-Aib-Ala-	
Boc-Aib-Leu-Aib-Aib-Leu-Leu-Aib-Leu-Aib-O-Me <u>Proteins</u> 12:324-30, 1992 Lys(Z)(benzyloxy-carbonyl)-Aib-O-Me <u>PNAS</u> 87:7921-5, 1990	
GELEELLKHLKELLKGER <u>Biochem.</u> 31:1579-84, 1992	

In another embodiment of the present invention, a third functional class of rerouting moieties is disclosed. This class includes moieties that can be recognized by intracellular receptors. Such sequences are identified by their ability to stop movement of endogenously synthesized proteins to the cell surface. Suitable peptides include certain peptide sequences (such as sorting or signal sequences) associated with the trafficking of endogenously synthesized proteins (Cur. Opin. Cell. Biol. 3:634-41, 1991). Such peptide sequences, when covalently attached to the C-terminus of an exogenously added targeting moiety, result in the retention of the agent/receptor complexes in the endoplasmic reticulum ("ER"), Golgi apparatus, or lysosomes.

Such peptide sequences are recognized by intracellular receptors, examples of that include both mammalian and bacterial versions of ER receptors described in detail in J. Cell. Biol. 120:325-8, 1993; Embo. J. 11:4187-95, 1992; Nature 348:162-3, 1990. Further exemplary peptide sequences and variants thereof (shown in parentheses) that can be recognized by intracellular receptors are set forth in Table 5, Sections A and B.

Certain signal sequences may be preferred for retention by one type of organism versus another type. For example, REDLK is a preferred sequence recognized by prokaryotic cells and to a lesser degree by eukaryotic cells (see Table 5, section C). Thus, employing this sequence as the rerouting moiety, receptor modulating agents can be constructed to selectively inhibit a receptor-mediated process in bacteria, while having little effect on mammalian cells.

TABLE 5
PEPTIDE SEQUENCES THAT BIND INTRACELLULAR RECEPTORS

A. Endoplasmic Reticulum or Golgi Retention Peptides

1. KDEL (DKEL, RDEL, KNEL, SDEL, KEEL, QDEL, KEDL, KDEL)	<u>J. Biol. Chem.</u> 265:5952-5, 1990 <u>Biochem. Biophys. Res. Commun.</u> 172:1384-91, 1990 <u>J. Virol.</u> 65:3938-42, 1991 <u>Exp. Cell Res.</u> 197:119-24, 1991 <u>Growth Factors</u> 5:243-53, 1991 <u>J. Biol. Chem.</u> 267(10):7022-6, 1992 <u>J. Biol. Chem.</u> 267:10631-7, 1992 <u>J. Cell Biol.</u> 118:795-811, 1992 <u>J. Cell Biol.</u> 119:85-97, 1992 <u>Exp. Cell Res.</u> 203:1-4, 1992 <u>P.N.A.S.</u> 90:2695-9, 1993 <u>Mol. Biochem Parasitol</u> 48:47-58, 1991 <u>Embo J.</u> 4:2345-55, 1992 <u>J. Biol. Chem.</u> 266:14277-82, 1991 <u>Mol. Cell Biol.</u> 11:4036-44, 1991
2. HDEL (HVEL, HNEL, HTEL, TEHT, DDEL, HIEL)	<u>J. Biol. Chem.</u> 268:7728-32, 1993 <u>Mol. Biochem Parasitol</u> 57:193-202, 1993 <u>J. Cell Sci</u> 102:261-71, 1992 <u>Eur J. Biochem.</u> 206:801-6, 1992 <u>J. Biol. Chem.</u> 266:20498-503, 1991
3. ADEL	<u>Embo J.</u> 11:1583-91, 1992
4. REDLK	<u>J. Biol. Chem.</u> 266:17376-81, 1991
5. SEKDEL	<u>Growth Factors</u> 5:243-53, 1991

6.	KTEL	<u>J. Virol.</u> <u>66</u> :4951-6, 1992
B. LYSOSOMAL RETENTION PEPTIDES		
1.	KFERQ	<u>Trends Biochem Sci</u> <u>15</u> :305-9, 1990
2.	Tyrosine-containing polypeptides	<u>J. Cell Biol.</u> <u>111</u> :955-66, 1990
C. ORGANISM-SPECIFIC RETENTION PEPTIDES		
1.	REDLK	<u>J. Biol. Chem.</u> <u>266</u> :17376-17381, 1991
D. CLATHRIN-BINDING PEPTIDES (INTERNALIZATION SIGNALS)		
1.	LLAV	<u>J. Cell. Biol.</u> <u>199</u> :249-57, 1992
2.	YKYSKV	<u>J. Cell. Biol.</u> <u>199</u> :249-57, 1992 <u>Embo. J.</u> <u>7</u> :3331-6, 1988
3.	PPGYE	<u>Cell</u> <u>67</u> :1203-9, 1991 <u>Curr. Opin. Cell Biol.</u> <u>3</u> :1062, 1991

A further class of peptide sequences of this invention, termed "internalization signals," function by binding to clathrin, both in the coated pits, as well as those intracellular vesicles that maintain a clathrin coat. Representative examples of such clathrin-binding peptides (CBP) are disclosed in Table 5, section D. The CBP binds clathrin in the coated pits initially located on the cell surface causing retention of the targeting moiety to that it is conjugated.

A further class of moieties capable of recognizing intracellular receptors includes carbohydrates. Suitable carbohydrates include any carbohydrate that is

capable of binding to intracellular carbohydrate (CHO) receptors but not to cell surface CHO receptors. Such carbohydrates include: mannose-6-phosphate and glucose-6-phosphate. Suitable carbohydrate moieties include those that bind to the insulin-like growth factor II/mannose-6-phosphate (IGF II/M6P) receptor, including
5 analogs of mannose-6-phosphate, as well as other phosphorylated saccharides (Carbohydrate Res. 213:37-46, 1991; FEBS Lett. 262:142-4, 1990).

The affinity of the rerouting moiety can be varied by changes in the chemical nature of the phosphorylated saccharides (J. Biol. Chem. 264:7970-5, 1989; J. Biol. Chem. 264:7962-9, 1989) (monosaccharides bind with the lowest affinity, while di- or
10 tri-saccharides bind with increasingly higher affinity). Clustering of phosphorylated saccharides on protein carriers can dramatically increase affinity to the intracellular receptor.

Syntheses of various oligosaccharides are reviewed in Sem. Cell. Biol. 2:319-326, 1991. Although, mannose-6-phosphate receptor expression is primarily
15 intracellular, expression also occurs on cell surfaces. Thus, in the context of the present invention, covalent attachment of a targeting moiety with a carbohydrate that binds the mannose-6-phosphate receptor should be constructed so as to give at least a hundredfold difference in binding affinity between the targeting moiety and the rerouting moiety. For example, a vitamin B₁₂/transcobalamin II receptor targeting
20 moiety, in this case vitamin B₁₂, would have a binding affinity for the carrier protein, transcobalamin II (TcII), of $\geq 10^{-10}$ M and an affinity for the IGF II/M-6-P receptor of 10^{-8} M or less. This will maintain the specificity of the vitamin B₁₂ binding (via TcII), while allowing transfer of the receptor modulating agent from serum M-6-P soluble receptor to cell surface receptor.

25 In addition to IGF II/M-6-P receptor moieties, other carbohydrate-based rerouting moieties also promote retention of the modulating agent/receptor complex in the ER or Golgi complex. Such moieties are based on the recognition by various glycosyl transferases of carbohydrate moieties, either as a natural substrate or as an inhibitor. Such moieties are reviewed in Sem. Cell. Biol. 2:289-308, 1991. For
30 example, saccharide recognition moieties include penultimate sugars, such as glucose and N-acetyl glucosamine (that are natural substrates). More preferred, however, are glycosylation inhibitors that are recognized by glycosyl transferases, but cannot serve to append further carbohydrate residues on growing chains (Sem. Cell. Biol. 2:309-318, 1991).

In yet another embodiment of the present invention, a fourth functional class of rerouting moieties is disclosed. This class is generally comprised of rerouting moieties that anchor the receptor to the cell membrane. By way of example, this class includes membrane-binding peptides that exhibit conditional pH-dependent membrane binding. Such peptides exhibit α -helical character in acid but not neutral pH solutions. When a conditional membrane-binding peptide assumes a helical conformation at an acidic pH, it acquires the property of amphiphilicity, (*e.g.*, it has both hydrophobic and hydrophilic interfaces). More specifically, within a pH range of approximately 5.0-5.5, such a peptide forms an alpha-helical, amphiphilic structure that facilitates insertion of the peptide into a target membrane. An alpha-helix-induced acidic pH environment may be found, for example, in the low pH environment present within cellular endosomes or lysosomes. In aqueous solution at physiological pH, a conditional, membrane-binding peptide is unfolded (due to strong charge repulsion among charged amino acid side chains) and is unable to interact with membranes.

Suitable conditional membrane-binding peptide sequences include the charged amino acids glutamate, aspartate, and histidine. A preferred conditional membrane-binding peptide includes those with a high percentage of helix-forming residues, such as glutamate, methionine, alanine, and leucine. Further, conditional membrane-binding peptide sequences include ionizable residues having pKas within the range of pH 5-7, so that a sufficiently uncharged membrane-binding domain will be present within the peptide at pH 5 to allow insertion into the target cell membrane. Conditional membrane-binding peptides can be incorporated through covalent bonds to a chemical or peptide targeting moiety or synthesized as an entire peptide sequence including a linker and peptide targeting moiety.

A particularly preferred conditional membrane-binding peptide is aa1-aa2-aa3-EAALA(EALA)₄-EALEALAA-amide, that represents a modification of a published peptide sequence (Biochemistry 26:2964, 1987). Within this peptide sequence, the first amino acid residue (aa1) is preferably a unique residue such as cysteine or lysine, that facilitates chemical conjugation of the conditional membrane-binding peptide to a targeting protein. The peptide can also be incorporated into a fusion protein with a protein or peptide targeting moiety (see Example 7). Amino acid residues 2-3 (*i.e.*, aa2-aa3) may be selected to modulate the affinity of the translocating peptide for different membranes. For instance, if both residues 2 and 3 are lysine or arginine, the peptide will have the capacity to bind to membranes or patches of lipids having a

negative surface charge. If residues 2-3 are neutral amino acids, the peptide will insert into neutral membranes.

Yet another preferred conditional membrane-binding peptide can be derived from sequences of apo-lipoprotein A-1 and B; peptide toxins such as melittin, bombolittin, delta hemolysin and the pardaxins; antibiotic peptides, such as alamethicin; peptide hormones, such as calcitonin, corticotrophin releasing factor, beta endorphin, glucagon, parathyroid hormone, and pancreatic polypeptide. Such peptides normally bind membranes at physiologic pH but through attachment of substituents the peptides can be enhanced in their ability to form alpha-helices at acidic pH and reduced in their membrane-binding at physiologic pH. An example of such a modified peptide having pH-dependent membrane binding at acidic pH is fully succinylated melittin. In this example, a peptide (melittin) that normally binds to membranes at physiological pH is converted to a pH-dependent peptide through succinylation of lysines. Upon succinylation, the peptide displays an amphipathic character only at acidic pHs.

Insertion of a conditional membrane-binding peptide into a target cell membrane is enhanced through stabilization of the amphiphilic alpha-helix. Helix stabilization may be achieved: (1) by adding repeating "EALA" units to form a longer peptide; (2) by placing an amide at the C-terminus of the peptide, in order to counteract the helical dipole; (3) by polymerizing the peptide; (4) by substituting a natural helix-former for one or more of the stacked glutamates; or (5) by attaching the peptide to a targeting moiety through use of a longer linker, in order to provide sufficient distance between the membrane binding peptide and the targeting moiety for the peptide to contact and interact with the target cell intracellular membranes.

In yet another embodiment of the present invention, a fifth functional class of rerouting moieties is disclosed. In this context, the rerouting moiety merely functions as a modulating agent in that the moiety disables the receptors by crosslinking the same. This class includes bi- or multivalent receptor crosslinking moieties formed from monovalent binding targeting moieties. Cross-linking of receptors in some receptor systems is sufficient to cause a rerouting of cell surface receptors to lysosomes for degradation, rather than their normal pathway of receptor recycling. The synthesis of a bivalent receptor modulating agent is exemplified in greater detail in the examples below.

A preferred cross-linking receptor modulating agent is a vitamin B₁₂ dimer. In this embodiment, each vitamin B₁₂ molecule acts as a targeting agent and a rerouting

agent; cross-linking the B₁₂ dimer will cross-link the vitamin B₁₂ receptors, thus impeding the receptor trafficking pathway. A preferred vitamin B₁₂ dimer is generally comprised of two vitamin B₁₂ molecules, such as cyanocobalamin, coupled by one or more linkers through coupling sites independently selected from *a-g*, *h* (ribose), and *i* (benzimidazole). Preferably, cross-linking occurs between *b*- or *e*- coupling sites on both molecules. The dimer must be capable of forming a B₁₂/TcII complex. As noted above, this characteristic may be assayed using any one of several techniques known in the art, including competitive binding assays.

A vitamin B₁₂ may be coupled to a second vitamin B₁₂ molecule in the same manner as described in detail for conjugation of rerouting moieties to vitamin B₁₂ targeting moieties. As noted above, dimers may be synthesized using one or more linkers of various lengths, solubilities, and any combination of homobifunctional, heterobifunctional, homotrifunctional, or heterotrifunctional linkers. As noted above, the use of a trifunctional linker allows for coupling with any number of additional moieties.

In selecting a linker for dimer synthesis, it should be noted that the total number of atoms comprising the linker between the vitamin B₁₂ molecules should generally be greater than 10 atoms, typically be in the range of 30 to 55 atoms and, preferably be about 45. As noted above, one of ordinary skill in the art will appreciate that, although the number of atoms is calculated relative to a linear chain of atoms, linear chain, branched chain, and cyclical chain linkers or combinations thereof would be suitable. Hence, the structure of the atom chain in a linker would include, by way of example, alkyl, heteroalkyl, alkylaryl, and heteroalkyl aryl.

By way of example, a dimer may be synthesized by combining two different vitamin B₁₂ linker adducts in the presence of a coupling agent. The linkers couple and dimers may then be separated and purified using the same methods outlined above. The synthesis of a representative vitamin B₁₂ linker adduct is described in Example 23.

Alternatively, activated vitamin B₁₂ may simply be combined with a homobifunctional or homotrifunctional linker. Preferably, in this embodiment, the ratio of vitamin B₁₂ to linker should be in the range of 2:1. Preferably, a 1:1 ratio is used in preparation of mixed dimers (e.g., *b*- and *e*-acid derivatives) or mixed ligands (e.g., B₁₂ and hormone). Dimers may be separated and purified as noted above.

In still another alternative, vitamin B₁₂ linker adducts, synthesized as described above, may be coupled by a third linker. The third linker, a "cross-linker," serves to bridge the linkers on the vitamin B₁₂ linker adducts. Suitable cross-linkers include

those noted above. The synthesis of representative vitamin B₁₂ dimers having bridging cross-linkers is described in Example 25.

In still another alternative, vitamin B₁₂ dimers may also be prepared utilizing a cross-linker that functions through high-affinity specific binding interactions. The high specific binding affinity of biotin for avidin ($K_a = 10^{15}$) provides such a high-affinity binding. Accordingly, the receptor modulating agents of the present invention include vitamin B₁₂ dimers and vitamin B₁₂ tetramers bound through the avidin-biotin interaction. An vitamin B₁₂/avidin conjugate may be readily prepared by the addition of a suitable vitamin B₁₂/biotin conjugate to avidin (or streptavidin).

Avidin and streptavidin are tetrameric proteins having four biotin binding sites situated in pairs on either side of the protein. Thus, a range of vitamin B₁₂/avidin conjugates may be prepared in that the number of vitamin B₁₂ molecules or derivatives per avidin may be varied from one to four. In one preferred embodiment, the vitamin B₁₂/avidin conjugate has four vitamin B₁₂ molecules or derivatives per conjugate. As noted above, these tetrameric vitamin B₁₂ conjugates may be prepared by binding a suitable vitamin B₁₂/biotin conjugate (*i.e.*, a vitamin B₁₂/biotin conjugate composed of one biotin and one vitamin B₁₂ molecule or derivative) to avidin under conditions in that all four avidin binding sites are utilized. Suitable vitamin B₁₂/biotin conjugates include those conjugates in that the biotin is covalently linked to a vitamin B₁₂ coupling site with a linker of a length sufficient to permit binding of the vitamin B₁₂ conjugate to avidin. Subject to the considerations noted above, suitable linkers are in the range of about 12 to 50 atoms in length, and preferably about 25 to 45.

In another preferred embodiment, the vitamin B₁₂/avidin conjugate is a vitamin B₁₂ dimer having two vitamin B₁₂ molecules or derivatives per conjugate. These vitamin B₁₂/avidin conjugates may be prepared using a suitable vitamin B₁₂/biotin conjugate having two biotins linked to a single vitamin B₁₂ molecule or derivative. In this embodiment, suitable vitamin B₁₂/biotin conjugates include conjugates in that the biotins of the conjugate may both bind to each of one pair of avidin binding sites. The length of the linker between the vitamin B₁₂ and the biotins in the vitamin B₁₂/biotin conjugate may be of a length sufficient to permit binding to a pair of avidin binding sites, but not so lengthy as to be capable of binding to one binding site on one side of the avidin, and another binding site on the opposite side of the avidin. In a preferred embodiment, the vitamin B₁₂/avidin conjugate prepared from avidin and two vitamin B₁₂/biotin conjugates (having two biotins per vitamin B₁₂) has each pair of biotin binding sites occupied by a vitamin B₁₂/biotin conjugate.

Polymerization of peptides for use in the present invention may be accomplished by placing a cysteine residue at each end of a peptide, followed by oxidation using dissolved oxygen or other mild oxidizing agent, such as oxidized glutathione. The average length of a polymerized peptide may be controlled by
5 varying the polymerization reaction conditions.

The amino acid sequence of any of the peptides of this invention may be selected to include all L-amino acids or all D-amino acids having a side chain pK_a from 5.0 to 9.0. D-amino acids may be advantageously used to form nonproteolyzable peptides, since the D-amino acids are not metabolized within the
10 cell. Further, the peptides of the present invention may include a combination of L- and D-amino acids, wherein D-amino acids are substituted for L-amino acids on either side of a proteolytic cleavage site. Yet another preferred noncleavable peptide incorporates peptide bond analogs that are not susceptible to proteolytic cleavage by cellular enzymes.

As discussed above, the receptor modulating agents of this invention comprise a targeting moiety coupled to the rerouting moiety. The rerouting moieties identified
15 above may be covalently attached to the targeting moiety by any one of several techniques known in the art, including (a) by chemical modifications such as a disulfide formation, thioether formation, amide formation or a reduced or nonreduced Schiff's base, (b) by direct peptide bond formation as in a fusion protein, or (c) by use
20 of a chemical and peptide linker. Suitable peptide linkers in this regard correspond to two or more amino acid residues that allow the rerouting peptide to assume its active conformation independent of its interaction with the targeting moiety, and that allows sufficient distance for rerouting moiety access to, for example, intracellular
25 membranes from the peptide attachment site on the targeting moiety.

In one embodiment, a rerouting moiety may be conjugated to a vitamin B_{12} targeting moiety by any one of several means, including, by way of example, coupling a rerouting moiety to a reactive group on a vitamin B_{12} linker adduct; coupling a
30 vitamin B_{12} to a reactive group on a rerouting moiety linker adduct or an appropriate side chain thereof; coupling a vitamin B_{12} linker adduct to a rerouting moiety linker adduct or an appropriate side chain thereof; coupling a rerouting moiety/biotin binding protein conjugate to a vitamin B_{12} /biotin conjugate; or coupling a rerouting moiety biotin conjugate to a vitamin B_{12} /biotin binding protein conjugate.

Coupling of a rerouting moiety to a vitamin B_{12} linker adduct, or a vitamin
35 B_{12} to a rerouting moiety linker adduct, may be accomplished using the same

techniques noted above for coupling a vitamin B₁₂ molecule with a linker. The only critical consideration of this aspect of the invention is that the total linker length must be sufficient to avoid steric hindrance. Preferably, the total linker length is at least 6 atoms.

5 Coupling of a rerouting moiety/biotin binding protein conjugate to a vitamin B₁₂/biotin conjugate may be accomplished using any one of several means described in detail in Avidin-Biotin Chemistry: A Handbook, ed. D. Savage, Pierce Chemical Co., 1992. Briefly, a biotin binding protein conjugate is prepared using a rerouting moiety or, as in a second embodiment, a vitamin B₁₂ molecule. Suitable biotin binding
10 proteins include avidin or streptavidin. In some circumstances, a linker may be utilized to distance the molecules. For example, when coupling a vitamin B₁₂ to an avidin, a linker of at least 6 atoms is preferred.

 A biotin conjugate is prepared using a vitamin B₁₂ molecule or, as in a second embodiment, a rerouting moiety. By way of example, a vitamin B₁₂ molecule is
15 combined with an NHS ester of biotin. Preferably, the vitamin B₁₂ molecule is a vitamin B₁₂ linker adduct as described above. Even more preferably, the vitamin B₁₂ molecule is a vitamin B₁₂ linker adduct characterized by a 14-atom linear linker coupled to the *b*- or *e*- coupling site. The synthesis of a representative vitamin B₁₂/biotin conjugate is described in Example 24.

20 Once formulated, coupling between the biotin conjugates and biotin binding protein conjugates is easily accomplished by combining the complementing conjugates, *i.e.*, a vitamin B₁₂/biotin conjugate with a rerouting moiety/avidin conjugate.

 In another aspect of the present invention, a B₁₂/biotin conjugate is utilized to
25 couple a vitamin B₁₂ to any number of compounds through biotin binding protein conjugates. Using a vitamin B₁₂/biotin conjugate, any compound that is capable of coupling a biotin binding protein may be coupled to a vitamin B₁₂ and thereby internalized into cells expressing the vitamin B₁₂ receptor. Such compounds include, in addition to the rerouting moieties described in detail below, hormones, enzymes,
30 antibodies or fragments thereof, markers, or therapeutics. Coupling any of these compounds to a biotin binding protein, such as avidin or streptavidin, may be accomplished using techniques described in detail in Avidin-Biotin Chemistry: A Handbook, ed. D. Savage, Pierce Chemical Co., 1992.

 In one aspect of this embodiment, a vitamin B₁₂/biotin conjugate is coupled to
35 a therapeutic/avidin conjugate directed at neoplastic disorders. Neoplastic disorder

therapeutics that may be coupled to a vitamin B₁₂/biotin conjugate through avidin include doxorubicin, daunorubicin, etoposide, teniposide, vinblastine, vincristin, cyclophosphamide, cisplatin, and nucleoside antimetabolites such as arabinosylcytosine, arabinosyladenine, and fludarabine.

5 In another aspect of this embodiment, a vitamin B₁₂/biotin conjugate is coupled to a marker conjugated with a biotin binding protein. Suitable markers include, by way of example, fluorescent molecules or radiolabeled molecules. This combination may be utilized as a detection system incorporated into a screening device to identify patients with low receptor bearing cells or in the evaluation of
10 receptor up-regulation, for example, following treatment of patients for any one of a wide variety of receptor modulation disorders.

In another aspect of this embodiment, a vitamin B₁₂/biotin conjugate is coupled to a radioisotope conjugated to a biotin binding protein. Suitable radioisotopes include, any high energy emitting radioisotopes capable of conjugating a
15 biotin binding protein. This combination may be utilized as a targeted radiodiagnostic or radiotherapeutic.

In yet another aspect of this embodiment, a vitamin B₁₂/biotin conjugate is used to immobilize vitamin B₁₂ to a solid matrix or avidin-coated substrate. By way of example, this would enable one to isolate TcII, TcII receptors, and evaluate
20 coupling sites on the Vitamin B₁₂.

The receptor modulating agents of this invention regulate receptor-dependent biological responses through alterations in the receptor trafficking pathway. As illustrated in FIGURE 1, with specific reference to the receptor for vitamin B₁₂, cell surface receptors are often associated with clathrin-coated pits. When bound by the
25 receptor modulating agent of the present invention, the coated pits invaginate to form vesicles. The vesicles are then directed by the rerouting agent to lysosomes for receptor degradation or delivered to endosomes where the rerouting agent securely binds or delays the agent/receptor complex. Thus, the receptor modulating agents can incapacitate the receptors normally undergoing recycling.

30 Newly synthesized receptors will eventually replace the internalized receptor on the cell surface. However, this process is far more time consuming than recycling-many cells require hours or days to achieve maximal receptor reexpression. Continued exposure of the cell to the receptor modulating agents will exhaust the intracellular receptor pools. Thus, by modulating a plasma membrane receptor,

reexpression of the receptor can be substantially delayed, thereby regulating a biological response associated with that receptor for a prolonged period of time.

Biological activity of receptor modulating agents of the present invention may be ascertained in vitro by any one of several means known in the art, including competition binding assays or cell proliferation studies. These techniques are described in detail in Laboratory Techniques in Biochemistry and Molecular Biology: An Introduction to Radioimmunoassay and Related Techniques, 3rd Edition, ed. Burdon and van Knippenberg, Elsevier, 1987. By way of example, a receptor modulating agent may be cultured with a suitable cell line, such as K562 cells (ATCC CCL 243), under conditions representing in vivo conditions. Such conditions would include the provision of a human source of TcII (such as human serum), vitamin B₁₂, and, preferably, by careful removal by chromatography, of all TcII from other medium supplements such that proliferation is solely dependent on a known amount of exogenous TcII. Cell cultures deprived of vitamin B₁₂ gradually lose their proliferative capacity, eventually resulting in cell death. Biological activity may be evaluated in vivo using techniques described in detail in Shieh et al., J. Immunol. 152(2):859-866, 1994 in that human tumor cell lines are injected into nude mice, followed by therapy with receptor modulating agents. Next, tumor cells are removed, single-cell suspensions prepared and TcII cell surface receptor density may be evaluated by flow cytometry and biotinylated vitamin B₁₂ and avidin FITC.

The receptor modulating agent of the present invention may be administered in a therapeutically effective amount to treat a variety of disorders characterized in that control of the disease process or symptoms can be achieved by modulation of one or more receptor systems and the associated biological responses. Such disorders include neoplastic disorders, autoimmune diseases, rheumatic arthritis, cardiovascular disease, and neurodegenerative diseases.

Common to many non-neoplastic disease processes is a stage in that the disease process itself, or its symptoms, can be halted or ameliorated by the use of an antiproliferative agent such as vitamin B₁₂/TcII receptor modulating agents. These commonly recognized stages include a sensitization or elicitation phase in that immune cells responsible for the disease become turned on by antigen specific or nonspecific means, followed by a proliferative phase in that the immune cells expand in number, and finally a symptomatic phase in that the expanded immune cells create tissue damage directly or indirectly. Neoplastic disorders include, by way of example, leukemia, sarcoma, myeloma, carcinoma, neuroma, melanoma, cancers of the breast,

lung, liver, brain, colon, cervix, prostate, Hodgkin's disease, and non-Hodgkin's lymphoma. Because of this, antiproliferative chemotherapeutic drugs are commonly utilized in the treatment of many diseases other than cancer, but are limited in use to life-threatening situations due to their associated toxicity. Antiproliferative agents, such as the ones of the present invention (with little of the direct toxicity of chemotherapeutic drugs), may be used more widely. More specifically, the vitamin B₁₂ receptor modulating agents of the present invention are not destructive to plasma membrane processes (*e.g.*, ion transport). In addition, the antiproliferative activity is reversible by administration of vitamin B₁₂. Furthermore, the agents of this invention may not be mutagenic, teratogenic, or carcinogenic since they act at the level of the plasma membrane, and not at the level of the nucleus, and DNA by intercalation or cross-linking (as many chemotherapeutic drugs act).

An understanding of the pharmaceutical applications for B₁₂/TcII receptor modulating agents requires a knowledge of the cell types targeted by such therapy. To this end, various pharmaceutical applications are disclosed in Table 6 below.

TABLE 6
TARGET CELLS FOR VITAMIN B₁₂ RECEPTOR MODULATING AGENTS

	TARGET CELL	OTHER PROLIFERATION ASSOCIATED MARKERS	POTENTIAL PHARMACEUTICAL APPLICATIONS
5	Activated T-Cell	IL-2 receptor Transferrin Receptor Insulin Receptor Class II Histocompatibility Antigens	Graft versus Host Disease Organ Transplants Auto-Immune Diseases Asthma Crohn's Disease
10	Tumor Cells	Tumor Assoc. Ags. Ki67 Transferrin Receptor	Tumor Therapy (alone and in combination with chemotherapeutic drugs)
15	Bone Marrow Stem Cells	CD-34 Transferrin Receptor Class II Histocompatibility Antigens IL-1, IL-3 Receptors	Allogeneic Bone Marrow Transplants Reduction in Toxicity of Chemotherapy
20	Proliferating Fibroblasts	Thy 1.1 Transferrin Receptor Insulin & Insulin-like Growth-Factor Receptors Fibroblast Growth-Factor Receptor	Inhibition of Adhesions, Scarring Scleroderma
25			
30	Proliferating Epithelium or Epidermal (Keratinocytes)	EGF Receptor Proto-Oncogenes	Psoriasis

Proliferating and activated T-cells can cause a wide variety of diseases ranging from the chronic inflammation of Crohn's disease to more acute organ graft rejection. In all of these diseases, the T-cell may serve a central pathogenic role or a more accessory role. Antiproliferative chemotherapeutic drugs serve to reduce symptomatology and in some cases lead to long-term remission. Similarly, proliferating fibroblasts and epithelial cells may give rise to diseases characterized by cell overgrowth. Vitamin B₁₂ receptor modulating agents may be used to replace or used in combination with existing chemotherapeutic regimens in these diseases. An important aspect of the use of antiproliferative vitamin B₁₂ receptor modulating agents in these diseases is not to apply it so aggressively or with improper timing such that normal healing (adhesions, scarring) or cell renewal (psoriasis) processes are also inhibited. As such, low doses of receptor modulating agents may be used during healing and higher doses used once healing is completed. Alternatively, receptor modulating agents may not be administered at all until after healing is completed.

As previously mentioned, B₁₂/TcII receptor modulating agents can be used to deprive neoplastic cells of vitamin B₁₂. It has already been shown that sufficient deprivation leads to the death of rapidly proliferating lymphoid neoplasms such as leukemia and lymphoma. Moreover, short-term treatment to reduce cellular availability of this nutrient, combined with existing chemotherapeutic agents, markedly improves therapeutic efficacy.

For solid tumors, vitamin B₁₂ depletion may induce cytostasis and differentiation as well as cell death. Thus, B₁₂/TcII receptor modulating agents may be used to induce differentiation in hormonally responsive solid tumors. An increase in the number of cells expressing a differentiated phenotype should translate into an increase in expression of hormone receptors. The hormone receptor status of tumors, such as breast and prostate cancer, are directly correlated with their response to hormonal therapy. Accordingly, B₁₂/TcII receptor modulating agents can be used to increase the number of receptor positive tumor cells or increase receptor density in order to enhance efficacy of subsequent hormonal therapy.

Vitamin B₁₂ receptor modulating agents may affect both replicating neoplastic and normal cells. However, bone marrow progenitors demonstrate differential sensitivity or response. Thus, B₁₂ receptor modulating agents can be used to modulate sensitivity of bone marrow progenitors so as to enhance their resistance to the toxic effects of chemotherapeutic agents. Such chemotherapeutic drugs act primarily on replicating cells, with nonreplicating cells being much less sensitive.

Decreasing the sensitivity of progenitors to toxic drugs would increase the bone marrow reserves and enhance subsequent response to colony stimulating factors, and enable higher doses of chemotherapy or reduce the interval to reconstitution. It should also be recognized that such positive effects on bone marrow progenitors, as a natural consequence of B₁₂ receptor therapy for cancer, are an additional mechanism by that the therapeutic index of chemotherapeutic drugs other than 5-FU and methotrexate can be improved.

In a variety of autoimmune diseases, graft versus host disease, ectopic allergy, and organ transplantation, an initial "induction" phase, in that the patient becomes sensitized to self- or allo-antigens, is followed by a "proliferative" phase in that forbidden or unregulated clones of B- or T-cells are expanded. It has long been known that treatment with antiproliferative, chemotherapeutic drugs following induction can inhibit expansion of forbidden clones, inhibit progression of disease, and restore a stable state of tolerance.

Inflammation is an application for that antibodies are already being utilized in clinical trials. The primary emphasis has been on inhibiting the early manifestations of inflammation by inhibiting recruitment or binding of inflammatory cells to the vascular endothelium of injured tissue. It also well recognized that proliferation of cells at the site of inflammation contributes to the pathology and tissue destruction of both acute as well as chronic inflammation. To this end, antiproliferative, chemotherapeutic drugs have been widely used to inhibit sequelae of inflammation.

Methotrexate is one such drug commonly used to treat symptoms associated with rheumatoid arthritis. The drug acts to reduce both localized (*e.g.*, synovium) and generalized inflammation associated with disease progression. Methotrexate acts synergistically with vitamin B₁₂ depletion in therapy of leukemia. B₁₂ receptor modulating agents can therefore be combined with methotrexate to enhance efficacy in rheumatoid arthritis. Other methotrexate applications include treating the destructive inflammation associated with chronic heart disease and colitis.

Surgery, radiation, or chemotherapy to the abdomen is often complicated by the development of tissue adhesions. These represent a considerable clinical problem because they lead to bowel blockage and require surgical intervention. Peritoneal adhesions arise as a result of proliferation of the cells of the peritoneal membrane lining the abdomen. A nontoxic means of interfering with such proliferation could lead to restoration of these normal cells to homeostatic control mechanisms and thereby inhibition of adhesion formation. A similar process of benign proliferation

and subsequent scarring is a complication of retinal surgery. Direct installation of a small molecule analog of an antibody receptor antagonist could prevent such disabling complications.

5 The term "treatment" as used within the context of the present invention, refers to reducing or alleviating symptoms in a subject, preventing symptoms from worsening or progressing, inhibition or elimination of the causative agent, or prevention of the infection or disorder in a subject who is free therefrom. Thus, for example, treatment of infection includes destruction of the infecting agent, inhibition of or interference with its growth or maturation, neutralization of its pathological effects and the like. A disorder is "treated" by partially or wholly remedying the deficiency that causes the deficiency or that makes it more severe.

The receptor modulating agents of the present invention are administered in a therapeutically effective dose. A therapeutically effective dose may be determined by in vitro experiment followed by in vivo studies.

15 Pharmaceutical compositions containing the receptor modulating agents in an admixture with a pharmaceutical carrier or diluent can be prepared according to conventional pharmaceutical compounding techniques. The carrier may take a wide variety of forms depending on the form of preparation desired for administration (*e.g.*, intravenous, oral, topical, aerosol, suppository, parenteral or spinal injection). Preferably, administration is via stereotactical injection.

The following examples are offered by way of illustration, not limitation.

EXAMPLES

In summary, the examples that follow disclose the synthesis of several receptor modulating agents of this invention utilizing different functional classes of rerouting moieties. More specifically, a series of examples are presented that employ vitamin B₁₂ as a targeting moiety in a receptor modulating agent.

25 All chemicals purchased from commercial sources were analytical grade or better and were used without further purification unless noted. Isophthaloyl dichloride was purchased from Lancaster Synthesis Inc. (Windham, NH). All other reagents were obtained from Aldrich Chemical Co. (Milwaukee, WI). Solvents for HPLC analysis were obtained as HPLC grade and were filtered (0.2 μ m) prior to use. Ion exchange chromatography was conducted with 200-400 mesh strongly basic anion 2% cross-linking Dowex-1-chloride (Aldrich Chemical Co). Amberlite XAD-2 nonionic polymeric adsorbent and octadecyl functionalized silica gel for column chromatography were obtained from Aldrich Chemical Co.

¹H NMR were obtained on a Bruker AC-500 (500 MHz) instrument. The chemical shifts are expressed as ppm (δ) using tetramethylsilane as internal reference. IR data were obtained on a Perkin-Elmer 1420 infrared spectrophotometer. UV data were obtained on a Perkin-Elmer Lambda 2 UV/V spectrophotometer. Mass spectral data were obtained on a VG 7070H mass spectrometer using fast atom bombardment (FAB).

HPLC separations of compounds were obtained on Hewlett-Packard quaternary 1050 gradient pumping system with a UV detector. Analysis of the HPLC data was obtained using Hewlett-Packard HPLC Chemstation software.

HPLC for Monomers: HPLC separations were conducted at a flow rate of 1 mL/min. on a 5 mm, 250 mm NH₂ column (RAININ microsorb-MV amino column) eluting with 58 mM pyridine acetate, pH 4.4 in H₂O:THF (96:4) solution. Retention times were: 1 = 4.3 min; 2 = 6.5 min; 3 = 8.0 min; 4 = 8.8 min; 5 = 10.9 min; 6 = 2.3 min; 7 = 2.3 min; 8 = 3.0 min; 9 = 2.9 min; 10 = 2.9 min; 13 = 3.4 min. Reverse-phase HPLC chromatography was carried out using a Hewlett-Packard LiChrospher 100 RP-18 (5 mm, 125 X 4 mm) C-18 column using a gradient solvent system at a flow rate of 1 mL/min. Solvent A in the gradient was methanol. Solvent B was H₂O. Starting from a 40% A, the gradient was increased to 100% A over 10 min. The gradient was then brought back to 40% A over a 5 min period. Retention times under these conditions for biotin conjugates were: 17 = 7.1 min; 18 = 7.2 min; 19 = 6.9 min; 20 = 6.4 min.

Preparative LC was conducted to separate the mixture of monocarboxylic acids using RAININ Rabbit-plus peristaltic pumping system with a DYNAMAX (model UV-1) UV-visible absorbance detector at a flow rate of 0.15 mL/min. An ID column (Alltech, 150 psi), (1000 mm X 25 mm) packed with aminopropyl silica (40-63 μ m) was used.

HPLC for Dimers: For dimers 36, 37, and 38 solvent A in the gradient was methanol. Solvent B was H₂O. The gradient was held at the starting mixture of 70% A for 2 min, then the percentage of A was linearly increased to 100% over the next 10 min. The gradient was held at 100% A for 20 min. Retention times under these conditions for dimers were: 36 = 8.7 min; 37 = 9.0 min; 38 = 8.9 min. For dimers 58-60 and 64-66 solvent A in the gradient was methanol. Solvent B was aqueous 1% acetic acid. The gradient was begun at 40% A and was held at that composition for 2 min, then the percentage of A was linearly increased to 100% over the next 10 min.

Retention times for the compounds examined under these conditions were: 58 = 14.0 min; 59 = 14.1 min; 60 = 13.9 min; 64 = 8.7 min; 65 = 8.6 min; 66 = 9.0 min.

Example 1

PREPARATION AND PURIFICATION OF CYANOCOBALAMIN MONOCARBOXYLATES: MODIFICATION ON THE RING

5 This example serves to demonstrate the hydrolysis of *b*-, *d*- and *e*-propionamide sites on a vitamin B₁₂ molecule using dilute acid in preparation for coupling of a linker to the sites. Importantly, the hydrolysis of the *b*-, *d*- and *e*-propionamides is selective over the hydrolysis of *α*-, *c*- and *g*-acetamides, or the *f*-amide in the heterocyclic chain connecting the benzimidazole. An optimal yield of
10 monocarboxylate to di- and tri-carboxylate derivatives was obtained at room temperature in 0.1 N HCl over a 10-day period. The nonhydrolyzed vitamin B₁₂ and the di- and tri-carboxylates produced were readily isolated from the desired
15 monocarboxylates by preparative liquid chromatography.

Specifically, cyanocobalamin (1) (3.7 mmol, 5 g) was dissolved in 500 mL of 0.1 N HCl and stirred at room temperature for 10 days under argon atmosphere. The solution was then neutralized with 6 N NaOH and the cobamides were desalted by extraction into phenol and applied to a 200 g (60 x 4 cm, 200-400 mesh) Dowex
20 Cl⁻ x 2 column (acetate form; prepared by washing with saturated sodium acetate until it was free from Cl⁻, then washing with 200 mL water). The column was eluted with water to remove unreacted cyanocobalamin and then eluted with 0.04 M sodium acetate (pH 4.67).

The first fraction of the elution contained three monocarboxylic acids. These
25 were desalted by extraction into 100 mL of 90% (w/w) phenol, twice with 25 mL and once with 10 mL of phenol. Three volumes of ethyl ether (3 x 160 mL) and one volume of acetone (160 mL) were added to the combined phenol extracts. Monocarboxylic acids were removed from the organic phase by extraction with water (2 x 100 mL). The combined aqueous phases were extracted twice with 20 mL of
30 ether to remove residual phenol. The aqueous solution of monocarboxylic acids was evaporated to dryness. Yield: 2.5 g (50%).

The mixture of three acids (0.350 g) was then applied to a 200 g (1000 mm x 25 mm) column of aminopropyl-coated silica (40-63 mm) and was eluted with 58 mM pyridine acetate pH 4.4 in H₂O:THF(96:4); the eluant was collected with an
35 automatic fraction collector. The first eluted acid was found to be

b-monocarboxylic acid (2), the second eluted acid was *e*-monocarboxylic acid (3), and the third eluted acid was *d*-monocarboxylic acid (4). The acid fractions were desalted by phenol extraction. The solids obtained were crystallized from aqueous acetone.

5 *b*-acid (2): yield 0.122 g (35%), mp 267-270°C with decomposition, ¹H NMR (MeOH-d₄, δ) 0.43 (s, 3H, C-20 CH₃); 1.00 (m, 2H); 1.18 (s, 3H, C-46 CH₃); 1.24 (d, 3H, Pr₃ CH₃); 1.36 (br s, 9H, C-47 CH₃, C-54 CH₃); 1.4 (s, 3H, C-25 CH₃); 1.9 (d, 7H, C-36 CH₃, C-30 CH₂, C-48 CH₂); 2.26 (d, 6H, B10 & B11, CH₃); 2.36 (d, 2H, C-26 CH₂); 2.57 (s, 10H, C-35 CH₃, C-31 CH₂, C-37 CH₂, C-53 CH₃); 2.8 (m, 2H, C-60 CH₂); 3.3 (m, 3H, C-8H, C-13H); 3.6 (m, 2H, Pr₁ CH₂); 3.7 (d, 1H, R₅); 3.9 (d, 1H, R₅); 4.0 (m, 1H, R₄); 4.12 (d, 1H, C-19); 4.17 (s, 1H, C-3); 4.3 (m, 1H, R₂); 4.5 (m, 1H); 4.7 (m, 1H, R₃); 6.0 (s, 1H, C-10); 6.2 (s, 1H, R₁); 6.5 (s, 1H, B4); 7.1 (s, 1H, B2); 7.2 (s, 1H, B7). MS (FAB⁺): m/e 1357 (M⁺ +1). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹. UV (MeOH): λ₃₆₀ (ε₂₃₄₄₁).

15 *e*-acid (3): yield 0.168 g (48%), mp 245-250° C with decomposition, ¹H NMR (MeOH-d₄, δ) 0.43 (s, 3H, C-20 CH₃); 1.01 (m, 2H); 1.15 (s, 3H, C-46 CH₃); 1.23 (d, 3H, Pr₃ CH₃); 1.36 (br s, 9H, C-47 CH₃, C-54 CH₃); 1.4 (s, 3H, C-25 CH₃); 1.83 (s, 4H, C-55 CH₂); 1.93 (m, 6H, C-36 CH₃, C-30 CH₂, C-48 CH₂); 2.22 (d, 6H, B10 & B11 CH₃); 2.35 (s, 3H, C-26 CH₂); 2.5 (d, 13H, C-35 CH₃, C-31 CH₂, C-37 CH₂, C-53 CH₃); 2.9 (m, 1H, C-60 H); 3.2 (m, 1H, C-13H); 3.4 (m, 1H, C-8 H); 3.6 (d, 1H, Pr₁ CH); 3.7 (d, 1H); 3.9 (d, 1H); 4.0 (m, 2H); 4.1 (d, 1H); 4.2 (m, 2H); 4.6 (m, 1H); 6.0 (s, 1H, C-10); 6.3 (d, 1H, R₁); 6.5 (s, 1H, B4); 7.0 (s, 1H, B2); 7.2 (s, 1H, B7). MS (FAB⁺): m/e 1357 (M⁺ +1). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹. UV (MeOH): λ₃₆₀ (ε₂₁₈₄₂).

25 *d*-acid (4): yield 0.060 g (17%), mp > 300° C, ¹H NMR (MeOH-d₄, δ) 0.43 (s, 3H, C-20 CH₃); 1.04 (m, 2H); 1.15 (s, 3H, C-46 CH₃); 1.25 (d, 3H, Pr₃ CH₃); 1.36 (br s, 9H, C-47 CH₃, C-54 CH₃); 1.4 (s, 3H, C-25 CH₃); 1.85 (s, 4H); 2.01 (s, 6H); 2.23 (d, 8H, B10 & B11 CH₃); 2.38 (d, 3H, C-26 CH₂); 2.53 (d, 13H, C-36 CH₃, C-30 CH₂, C-48 CH₂); 2.6 (m, 5H); 2.9 (m, 1H, C-60 H); 3.3 (d, 1H, C-13H); 3.4 (m, 1H, C-8 H); 3.6 (d, 1H, Pr₁ CH); 3.7 (d, 1H); 3.9 (d, 1H); 4.0 (m, 2H); 4.1 (d, 1H); 4.3 (m, 2H); 6.0 (s, 1H, C-10); 6.3 (d, 1H, R₁); 6.5 (s, 1H, B4); 7.1 (s, 1H, B2); 7.2 (s, 1H, B7); UV (MeOH): λ₃₆₀ (ε₂₂₁₂₇). MS (FAB⁺): m/e 1357 (M⁺ +1). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹.

Example 2

CYANOCOBALAMIN MODIFIED ON RIBOSE: SUCCINATE CONJUGATE (5)

This example serves to demonstrate the activation of the ribose coupling site coupling site *h* (see structure I) with succinic anhydride. Cyanocobalamin (1) (0.15 mmol, 200 mg) was dissolved in 40 mL of dimethylsulfoxide (DMSO) containing 8 g (80 mmol) of succinic anhydride and 6.4 mL of pyridine. After 14-16 h at room temperature, the excess of succinic anhydride was destroyed by adding 500 mL of water and keeping the pH of the reaction mixture at 6 with 10% KOH. KCN was then added at a final concentration of 0.01 M and the pH of the solution was readjusted to 6 with 3 N HCl. After 1 h the cyanocobalamin components were desalted by phenol extraction and applied to a 100 g of Dowex Cl⁻ (60 x 2.5 cm) column (acetate form, 200-400 mesh). The cyanocobalamin was eluted with water. Succinate conjugate (5) was eluted with NaOAc (0.04 M, pH 4.67) that yielded 180 mg (85 %) after isolation. The O2',O5'-disuccinyl derivative remained absorbed on the column under these conditions. mp 208-210° C with decomposition.

¹H NMR (D₂O-d₄, δ): 0.43 (s, 3H, C-20 CH₃); 0.95 (m, 2H); 1.15 (s, 3H); 1.2 (d, 3H); 1.35 (d, 7H); 1.4 (s, 3H); 1.8 (s, 3H); 1.9 (s, 12H); 2.2 (d, 6H); 2.36 (d, 2H); 2.5 (d, 10H); 2.6-2.7 (m, 7H); 3.0 (m, 1H); 3.3 (d, 1H); 3.37 (m, 1H); 3.5 (d, 1H); 4.0 (d, 1H); 4.18 (m, 2H); 4.25 (m, 3H); 4.54 (d, 1H); 6.0 (s, 1H); 6.3 (d, 1H); 6.4 (s, 1H); 7.0 (s, 1H); 7.2 (s, 1H). MS (FAB⁺): m/e 1455 (M⁺ + 1). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹; UV (MeOH): λ₃₆₀ (ε₂₆₀₄₁).

Example 3

COUPLING OF CYANOCOBALAMIN MONOCARBOXYLIC ACIDS WITH 1,12-DIAMINODODECANE: REACTION WITHOUT SODIUM CYANIDE

This example serves to demonstrate the coupling of a linker to a cyanocobalamin monocarboxylate. Coupling of the monocarboxylates (2, 3, 4) with diaminododecane was first attempted using N-ethyl-N'-dimethylamino-propylcarbodiimide hydrochloride (EDC) in H₂O according to Yamada and Hogenkamp, *J. Biol. Chem.* **247**, 6266-6270, 1972. However, the products obtained did not have a reactive amino group. Alteration of the reaction conditions by changing the reaction mixture to DMF/H₂O and adding NaCN/N-hydroxysuccinimide (see Example 4) to the reaction mixture gave the desired diaminododecane adducts.

A mixture of cyanocobalamin monocarboxylic acid (0.370 mmol, 500 mg) and 1,12-diaminododecane (3.6 g) in 100 mL H₂O was adjusted to pH 6 with 1 N HCl.

The solution was then treated with N-ethyl-N'-dimethylamino-propyl-carbodiimide-hydrochloride (EDC) (726 mg) and stirred at room temperature for 22 h. In five intervals of 6 to 14 h, 650 mg of EDC was added to the reaction mixture. After a total reaction time of 4 days (HPLC monitoring) the solution was evaporated to dryness, the residue was digested with 100 mL of acetone, and the solvent was decanted. The solid residue was dissolved in 50 mL of water and applied to an 175 g Amberlite XAD-2 (60 x 4 cm) column. Contaminants were washed from the column with 1L water, then the crude product was eluted with 500 mL of methanol. The solution was evaporated to dryness, the residue was dissolved in 25 mL of water and was applied to a 100g Dowex Cl⁻ (60 x 2.5 cm) column (acetate form, 200-400 mesh). The final product was eluted using 250 mL of water, thereby leaving nonconverted acid bound to the column, that was later eluted with 0.04 mol/L sodium acetate buffer pH 4.67. The fraction containing the final product was evaporated to dryness.

The mass spectral value obtained indicated that HCN was lost from the desired product. Further, ¹H NMR data suggested that some protons were being affected by the cobalt. Thus, this reaction was conducted with NaCN (Example 4) to drive the equilibrium towards retention of Co-CN. N-hydroxy succinimide was also added to facilitate the coupling reaction.

e-acid adduct (6): Yield: 222 mg (40%). mp 172-174° C with decomposition. ¹H NMR (MeOH-d₄, δ): 0.43 (m, 3H, C-20 CH₃); 1.06 (t, 4H, C-46 CH₃); 1.16 (m, 5H); 1.2 (m, 5H); 1.33 (m, 7H); 1.43 (s, 3H); 1.68 (m, 4H); 1.86 (m, 5H); 2.2 (m, 8H); 2.3 (m, 6H); 2.4 (m, 10H); 2.55 (m, 10H); 2.8 (m, 4H); 3.1 (m, 6H); 3.3 (m, 5H); 3.6 (m, 2H); 3.7 (m, 2H); 3.8 (m, 1H); 4.0 (m, 1H); 4.1 (m, 1H); 4.16 (m, 1H); 4.3 (m, 1H); 4.48 (m, 1H); 4.6 (m, 1H); 6.0 (d 1H, C-10); 6.2 (m, 1H, R1); 6.5 (m, 1H, B4); 7.1 (m, 1H, B2); 7.2 (m, 1H, B7). MS (FAB⁺): m/e 1512. IR (KBr): 3400, 3200, 2950, 1660, 1570, 1490, 1060 cm⁻¹. UV (MeOH): λ₃₆₀ (ε₂₁ 877).

d-acid adduct (7): yield: 225 mg (45%), mp 195-198° C with decomposition. ¹H NMR (MeOH-d₄, δ): 0.43 (m, 3H, C-20 CH₃); 1.09 (m, 7H); 1.14 (m, 6H); 1.2 (m, 10H); 1.27 (m, 10H); 1.33 (m, 6H); 1.5 (m, 3H); 1.77 (s, 3H); 2.2 (m, 8H); 2.26 (s, 2H); 2.5 (m, 10H); 2.7 (m, 5H); 3.0 (m, 2H); 3.1 (m, 2H); 3.2 (m, 3H); 3.5 (m, 2H); 3.6 (m, 1H); 3.8 (m, 1H); 3.9 (m, 1H); 4.0 (m, 1H); 4.1 (m, 1H); 4.2 (m, 1H); 4.4 (m, 1H); 4.6 (m, 1H); 6.0 (d 1H, C-10); 6.1 (m, 1H, R₁); 6.4 (m, 1H,

B4); 7.0 (m, 1H, B2); 7.1 (m, 1H, B7); MS (FAB⁺): m/e 1512, IR (KBr): 3400, 3200, 2950, 1660, 1570, 1490, 1060 cm⁻¹; UV (MeOH): λ 360 (ϵ 22 680).

Example 4

COUPLING OF CYANOCOBALAMIN MONOCARBOXYLIC ACIDS WITH

5 1,12-DIAMINODODECANE: REACTION CONTAINING SODIUM CYANIDE

Cyanocobalamin monocarboxylic acid (2, 3, 4) (0.370 mmol, 500 mg) and N-hydroxysuccinimide (1.48 mmol, 170 mg) were dissolved in a mixture of DMF:H₂O(1:1) (18.4 mL) and 363 mg of NaCN was added. 1,12-Diaminododecane was dissolved in a mixture of DMF : H₂O (1:1) (18.4 mL) and the pH was adjusted to
10 6 with 1 N HCl. The diaminododecane solution was then added in one portion to the cyanocobalamin solution. EDC (285 mg) was added and the pH of the solution was readjusted to 5.5. The reaction mixture was then stirred overnight in the dark at room temperature. In five intervals of 6-14 h, 170 mg of N-hydroxysuccinimide and 285 mg of EDC were added to the solution, readjusting the pH value 5.5 each time. After
15 a total reaction time of 4 days (reaction followed by HPLC), the solution was evaporated to dryness. The residue was digested with 100 mL of acetone and the solvent was decanted. The solid residue was dissolved in 50 mL of H₂O and applied to an 200 g Amberlite XAD-2 (60 x 4 cm) column. The column was eluted with 1 L water to remove undesired materials, then the desired product was eluted with
20 500 mL methanol. The solution was evaporated to dryness, the residue was dissolved in 25 mL of water and was applied to a 100 g Dowex Cl⁻ (60 x 2.5 cm) column (acetate form, 200-400 mesh). The desired product was eluted from the column with 250 mL water, leaving any nonreacted acid bound to the column. This was followed by elution with 0.04 mol/L sodium acetate buffer pH 4.7. The fractions
25 containing the final product were evaporated to dryness.

b-isomer (8): yield 410 mg (82%), mp 172-174° C with decomposition. ¹H NMR (MeOH-d₄, δ) 0.43 (s, 3H, C-20 CH₃); 1.18 (s, 4H); 1.3 (m, 13H); 1.39 (m, 13H); 1.45 (s, 5H); 1.6 (m, 4H); 1.72 (m, 2H); 1.9 (s, 6H); 2.25 (d, 6H, B10 & B11 CH₃); 2.35 (m, 5H); 2.56 (m, 5H); 2.8-3.0 (m, 8H); 3.15 (m, 4H); 3.3
30 (m, 2H); 3.4 (m, 2H); 3.6 (m, 1H); 3.68 (m, 1H); 3.75 (m, 1H); 3.9 (d, 1H); 4.07 (m, 1H); 4.12 (d, 1H); 4.2 (br s, 1H); 4.3 (m, 1H); 4.47 (m, 1H); 4.7 (m, 1H); 6.0 (s, 1H, C-10); 6.2 (d, 1H, R₁); 6.5 (s, 1H, B4); 7.1 (s, 1H, B2); 7.2 (s, 1H, B7); MS (FAB⁺): m/e 1539 (M⁺ +1). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹. UV (MeOH): λ 360 (ϵ 15409).

e-isomer (9): yield: 430 mg (86%), mp 175-180°C with decomposition, ¹H NMR (MeOH-d₄, δ) 0.43 (s, 3H, C-20 CH₃); 1.17 (s, 4H, C-46 CH₃); 1.22 (d, 4H, Pr₃ CH₃); 1.29 (s, 24H); 1.36 (br s, 6H); 1.4 (s, 6H); 1.6 (m, 3H); 1.87 (s, 8H); 2.05 (m, 2H); 2.25 (s, 6H, B10 & B11 CH₃); 2.36 (m, 3H); 2.55 (d, 10H); 2.8 (s, 4H); 3.06 (t, 2H); 3.1 (m, 3H); 3.3 (s, 1H); 3.34 (m, 1H); 3.4 (m, 1H); 3.58 (m, 1H); 3.65 (m, 1H); 3.75 (d, 1H); 3.9 (d, 1H); 4.0 (m, 1H); 4.1 (d, 1H); 4.16 (m, 1H); 4.3 (m, 2H); 4.48 (m, 2H); 4.6 (m, 1H); 6.0 (s, 1H, C-10); 6.3 (d, 1H, R₁); 6.5 (s, 1H, B₄); 7.0 (s, 1H, B₂); 7.2 (s, 1H, B₇); MS (FAB⁺): m/e 1539 (M⁺ +1). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹. UV (MeOH): λ₃₆₀ (ε16 720).

d-isomer (10): yield: 400 mg (80%), mp 174-178° C with decomposition, ¹H NMR (MeOH-d₄, δ) 0.43 (s, 3H, C-20 CH₃); 1.07 (m, 3H, C-46 CH₃); 1.2 (d, 4H, Pr₃ CH₃); 1.27 (m, 15H); 1.35 (br s, 9H); 1.42 (s, 3H); 1.53 (m, 2H); 1.6 (m, 4H); 1.86 (s, 4H); 2.25 (d, 6H, B10 & B11 CH₃); 2.5 (d, 10H); 2.8 (s, 3H); 2.9 (m, 6H); 3.15 (m, 3H); 3.2 (m, 4H); 3.4 (m, 3H); 3.6 (d, 1H); 3.75 (d, 1H); 3.96 (d, 1H); 4.08 (m, 2H); 4.19 (m, 1H); 4.3 (m, 2H); 4.65 (m, 1H); 6.0 (s, 1H, C-10); 6.3 (d, 1H, R₁); 6.5 (s, 1H, B₄); 7.1 (s, 1H, B₂); 7.2 (s, 1H, B₇); UV (MeOH): λ₃₆₀ (ε17 665). MS (FAB⁺): m/e 1539 (M⁺ +1). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹.

20

Example 5

COUPLING OF CYANOCOBALAMIN MONOCARBOXYLIC ACIDS WITH GAMMA-AMINOBUTYRIC ACID (GABA)

This example serves to demonstrate the coupling of a gamma-aminobutyric acid (GABA) linker to a vitamin B₁₂ molecule. This reaction scheme is represented in
25 FIGURE 3.

Gamma-aminobutyric acid (GABA) *tert*-butyl ester (11) (1 mmol) and cyanocobalamin monocarboxylates (2, 3, 4) (0.1 mmol.) are mixed in 20 mL H₂O and sufficient 0.1 N HCl is added to adjust to pH to 6.0. N-ethyl-N¹-dimethylaminopropylcarbodiimide hydrochloride (EDC) (0.5 mmol) is added to the
30 solution. The reaction mixture is stirred at room temperature for 24 hours and then the mixture is dried under vacuum. This reaction mixture is treated with TFA to remove the *tert*-butyl ester. A cyanocobalamin-GABA adduct (12) was purified. Reverse-phase HPLC chromatography is carried out as described above. A cyanocobalamin-GABA adduct (12) can be further activated with a carbodiimide and
35 coupled to a moiety as described below.

Example 6**CYANOCOBALAMIN MODIFIED ON RIBOSE:****SUCCINATE-DIAMINODODECANE CONJUGATE (13)**

Cyanocobalamin-ribose-succinate (5) (0.370 mmol, 538 mg) and N-hydroxysuccinimide (1.48 mmol, 170 mg) were dissolved in a mixture of DMF:H₂O (1:1) (18.4 mL) and 363 mg of NaCN was added. This reaction scheme is represented in FIGURE 11. 1,12-Diaminododecane was taken in a mixture of DMF:H₂O (1:1) (18.4 mL), pH was adjusted to 6 with 1N HCl. The diaminododecane solution was then added in a portion to the cyanocobalamin solution. EDC (285 mg) was added, the pH of the solution was readjusted to 5.5 and the reaction mix. was stirred overnight in the dark at room temperature. In five intervals of 6 to 14 h 170 mg of N-hydroxysuccinimide and 285 mg of EDC was added to the solution, readjusting the pH 5.5 each time. After a total reaction time of 4 days (HPLC monitored) the solution was evaporated to dryness, the residue was digested with 100 mL of acetone, and the solvent was decanted. The solid residue was dissolved in 50 mL of H₂O and applied to a 200 g Amberlite XAD-2 (60 x 4 cm) column. Contaminants were washed from the column with 1 L water and then the crude product was eluanted with 500 mL methanol. The solution was evaporated to dryness, the residue was dissolved in 25 mL of water and was applied to a 100 g Dowex Cl⁻ (60 x 2.5 cm) column (acetate form, 200-400 mesh). The final product was eluanted using 250 mL water, thereby leaving nonconverted acid bound to the column, that was later eluanted with 0.04 mol/L sodium acetate buffer pH 4.7. The fraction containing the final product (13) was evaporated to dryness. Yield: 425 mg (70%), mp 185-187°C with decomposition.

¹H NMR (MeOH-d₄, δ): 0.43 (s, 3H, C-20 CH₃); 1.15 (s, 3H); 1.2 (d, 3H); 1.3 (s, 27H); 1.4 (m, 3H); 1.55 (m, 6H); 1.85 (m, 12H); 2.2 (d, 6H); 2.3 (d, 6H); 2.5 (d, 10H); 2.8 (m, 10H); 3.0 (t, 3H); 3.1 (t, 3H); 3.2 (s, 6H); 3.3 (m, 4H); 3.58 (m, 2H); 3.6 (d, 1H); 4.1 (d, 1H); 4.2 (m, 2H); 4.3 (m, 1H); 4.4 (d, 1H); 6.0 (s, 1H); 6.2 (d, 1H); 6.5 (s, 1H); 7.1 (s, 1H); 7.2 (s, 1H). MS (FAB⁺): m/e 1638 (M⁺). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹; UV (MeOH): λ360.

Example 7

MODIFICATION OF CYANOCOBALAMIN MONOCARBOXYLIC ACIDS CONJUGATED WITH 1,12-DIAMINODODECANE: REACTION WITH SUCCINIC ANHYDRIDE

This example serves to demonstrate modification of an amino terminus linking moiety to a carboxylate terminus. Such a modification may be necessary for conjugating amino containing rerouting agents (*e.g.*, aminosugars) to cyanocobalamin derivatives containing a linker.

Cyanocobalamin carboxylic acid diaminododecane conjugate (8, 9, 10) (0.138 mmol, 200 mg) was dissolved in 40 mL of dimethylsulfoxide (DMSO) containing 8 g (80 mmol) of succinic anhydride and 6.4 mL of pyridine. After 14-16 h at room temperature, the excess of succinic anhydride was destroyed by adding 500 mL of water and keeping the pH of the reaction mixture at 6 with 10% KOH. KCN was then added at a final concentration of 0.01 M and the pH of the solution was readjusted to 6 with 3 N HCl. After 1 h the cyanocobalamin components were desalted by phenol extraction. The residue was digested with 100 mL of acetone and the solvent was decanted. It was dissolved in 40 mL of H₂O. 1N NaOH (2 mL) was added to it and the reaction was stirred at room temperature for 15-20 min. It was then neutralized with 1N HCl and the cyanocobalamin components (14, 15, 16) were desalted by phenol extraction. Yield: 80 mg (40%); mp 190-198°C with decomposition.

¹H NMR (MeOH-d₄, δ): 0.43 (s, 3H, C-20 CH₃); 1.17 (s, 4H, C-46 CH₃); 1.23 (d, 4H, Pr₃ CH₃); 1.29 (s, 24H); 1.36 (br s, 6H); 1.4 (s, 6H); 1.87 (s, 4H); 2.05 (m, 2H); 2.25 (s, 6H, B10 & B11 CH₃); 2.35 (m, 3H); 2.4 (m, 5H); 2.55 (d, 10H); 2.7 (s, 5H); 2.8 (m, 2H); 3.1 (m, 6H); 3.3 (s, 6H); 3.4 (m, 1H); 3.65 (m, 2H); 3.75 (d, 1H); 3.9 (d, 1H); 4.0 (m, 1H); 4.1 (d, 1H); 4.16 (m, 1H); 4.3 (m, 1H); 4.48 (m, 1H); 4.6 (m, 2H); 6.0 (s, 1H, C-10); 6.3 (d, 1H, R₁); 6.5 (s, 1H, B4); 7.1 (s, 1H, B2); 7.2 (s, 1H, B7). MS (FAB⁺): m/e 1639 (M⁺). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹. UV (MeOH): λ₃₆₀ (ε 22 564).

Example 8

CYANOCOBALAMIN MODIFIED ON MONOCARBOXYLIC ACID: DIAMINODODECANE-BIOTIN CONJUGATES

This example serves to demonstrate coupling a vitamin B₁₂ derivative and biotin. Biotin conjugates (17, 18, 19) were obtained by reaction of activated

cyanocobalamin monocarboxylic acid diaminododecane (14), (15), and (16) with the NHS ester of biotin (Sigma Chemical Co.).

To a solution of cyanocobalamin monocarboxylic acid diaminododecane conjugate (14, 15, 16) (300 mg, 0.195 mmol) in DMF (35 mL) was added triethylamine (0.027 mL, 0.195 mmol). N-Hydroxysuccinimidobiotin (100 mg, 0.295 mmol) was then added over a period of 10-15 min and evaporated to dryness. The solid residue was dissolved in 20 mL of water and applied to an 75 g of Dowex Cl⁻ (40 x 2 cm) (acetate form, 200-400 mesh) column. The product was eluated using 250 mL of water. It was then evaporated to dryness, the residue was dissolved in a 10 mL of methanol - water (7:3 v/v) and the solution was applied to a reverse-phase C-18 column (500 mm x 25 mm, Alltech, 150 psi) that was developed with the same solvent. RAININ Rabbit-plus peristaltic pumping system was used with a DYNAMAX (model UV-1) UV visible absorbance detector. The eluant was collected with an automatic fraction collector. The fractions containing the final product (HPLC monitored) were evaporated to dryness.

b-isomer (17): yield 159 mg (53%), mp 210-212°C with decomposition, ¹H NMR (MeOH-d₄, δ): 0.43 (s, 3H, C-20 CH₃); 1.18 (s, 4H); 1.3 (m, 13H); 1.39 (m, 13H); 1.45 (s, 5H); 1.6 (m, 4H); 1.72 (m, 2H); 1.9 (s, 6H); 2.2 (d, 8H, B10 & B11 CH₃); 2.6 (d, 12H); 2.7 (m, 3H); 2.8-3.0 (m, 8H); 3.1 (m, 3H); 3.2 (m, 2H); 3.4 (s, 1H); 3.6 (m, 2H); 3.68 (d, 1H); 3.75 (m, 1H); 3.9 (d, 1H); 4.07 (m, 1H); 4.12 (d, 1H); 4.2 (s, 1H); 4.3 (m, 1H); 4.47 (m, 1H); 4.7 (m, 1H); 6.0 (s, 1H, C-10); 6.2 (d, 1H, R1); 6.5 (s, 1H, B4); 7.1 (s, 1H, B2); 7.2 (s, 1H, B7); MS (FAB⁺): m/e 1764 (M⁺). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹. UV (MeOH): λ₃₆₀ (ε₂₃ 746).

Anal. Calcd. for C₈₅H₁₂₇N₁₇O₁₆CoPS·11H₂O: C, 51.98; H, 7.59; N, 12.13. Found: C, 51.91; H, 7.81; N, 12.31.

e-isomer (18): yield 174 mg (58%), mp 222-224°C with decomposition, ¹H NMR (MeOH-d₄, δ): 0.43 (s, 3H, C-20 CH₃); 1.17 (s, 4H, C-46 CH₃); 1.22 (d, 4H, Pr₃ CH₃); 1.29 (s, 24H); 1.36 (br s, 6H); 1.4 (s, 6H); 1.6 (m, 4H); 1.72 (m, 2H); 1.87 (s, 4H); 2.17 (m, 3H); 2.25 (s, 6H, B10 & B11 CH₃); 2.36 (m, 3H); 2.55 (d, 10H); 2.64 (m, 2H); 2.8 (s, 4H); 2.97 (s, 4H); 3.1 (m, 3H); 3.3 (m, 1H); 3.4 (m, 1H); 3.58 (m, 1H); 3.65 (m, 1H); 3.75 (d, 1H); 3.9 (d, 1H); 4.0 (m, 1H); 4.1 (d, 1H); 4.16 (m, 1H); 4.3 (m, 2H); 4.48 (m, 2H); 4.6 (m, 1H); 6.0 (s, 1H, C-10); 6.3 (d, 1H, R1); 6.5 (s, 1H, B4); 7.0 (s, 1H, B2); 7.2 (s, 1H, B7); MS (FAB⁺):

m/e 1764 (M^+). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm^{-1} . UV (MeOH): λ_{360} ($\epsilon_{24\ 441}$).

Anal. Calcd. for $\text{C}_{85}\text{H}_{127}\text{N}_{17}\text{O}_{16}\text{CoPS}\cdot 9\text{H}_2\text{O}$ (13): C, 52.96; H, 7.53; N, 12.35. Found: C, 52.85; H, 7.55; N, 12.30.

5 *d-isomer* (19): yield 165 mg (55%), mp 216-218°C with decomposition, ^1H NMR (MeOH- d_4 , δ): 0.43 (s, 3H, C-20 CH_3); 1.16 (s, 3H, C-46 CH_3); 1.2 (d, 4H, Pr_3CH_3); 1.28 (s, 15H); 1.35 (br s, 9H); 1.42 (s, 3H); 1.53 (m, 2H); 1.6 (m, 4H); 1.72 (m, 2H); 1.86 (s, 6H); 2.16 (m, 3H); 2.02 (m, 4H); 2.25 (d, 6H, B10 & B11 CH_3); 2.5 (d, 10H); 2.7 (d, 1H); 2.8 (m, 5H); 3.1 (m, 6H); 3.2 (m, 3H); 3.4 (m, 1H); 3.57 (m, 1H); 3.6 (d, 1H); 3.7 (d, 1H); 3.9 (d, 1H); 4.0 (m, 1H); 4.11 (d, 1H); 4.17 (m, 1H); 4.3 (m, 2H); 4.4 (m, 2H); 4.6 (m, 1H); 6.0 (s, 1H, C-10); 6.3 (d, 1H, R1); 6.5 (s, 1H, B4); 7.1 (s, 1H, B2); 7.2 (s, 1H, B7); MS (FAB $^+$): m/e 1764 (M^+); IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm^{-1} ; UV (MeOH): λ_{360} ($\epsilon_{29\ 824}$).

15 Anal. Calcd for $\text{C}_{85}\text{H}_{127}\text{N}_{17}\text{O}_{16}\text{CoPS}\cdot 10\text{H}_2\text{O}$: C, 52.46; H, 7.56; N, 12.24. Found: C, 52.27; H, 7.56; N, 12.34.

Example 9

CYANOCOBALAMIN MODIFIED ON RIBOSE:

SUCCINATE-DIAMINODODECANE-BIOTIN CONJUGATE (20)

20 This example serves to demonstrate the conjugation of the ribose-linked diaminododecane adduct (13) with biotin to produce a cyanocobalamin biotin conjugate (20).

To a solution of (11) (300 mg, 0.183 mmol) in DMF (35 mL), triethylamine (0.025 mL, 0.183 mmol) was added. N-hydroxysuccinimidobiotin (100 mg, 0.295 mmol) was added over a period of 10-15 min. and then evaporated to dryness. The solid residue was dissolved in 20 mL of water and adjusted to pH 10 with 1N NaOH and applied to a 75 g Dowex Cl^- (40 x 2 cm) (200-400 mesh) column. The water fraction was discarded. The product was then eluanted with 0.1N NH_4OAc and was desalted by phenol extraction. The residue was dissolved in a 10 mL of methanol - water (7:3 v/v) and the solution was applied to a reverse-phase column (octadecyl), that was developed with the same solvent. The fractions containing the final product (20) (HPLC monitored) were evaporated to dryness. Yield 135 mg (45 %), mp 198-205°C with decomposition.

35 ^1H NMR (MeOH- d_4 , δ): 0.43 (s, 3H, C-20 CH_3); 1.15 (s, 3H); 1.2 (d, 3H); 1.3 (s, 27H); 1.36 (m, 6H); 1.4 (m, 3H); 1.6 (m, 4H); 1.7 (m, 2H); 1.85 (m, 12H);

2.0 (d, 3H); 2.17 (m, 3H); 2.2 (d, 6H); 2.3 (d, 6H); 2.5 (d, 10H); 2.64 (m, 2H); 2.8 (m, 10H); 3.1 (m, 6H); 3.25 (m, 6H); 3.58 (m, 2H); 4.0 (m, 1H); 4.1 (m, 1H); 4.16 (m, 1H); 4.4 (m, 1H); 4.6 (s, 2H); 4.7 (m, 1H); 6.0 (s, 1H); 6.2 (d, 1H); 6.5 (s, 1H); 7.1 (s, 1H); 7.2 (s, 1H). MS (FAB⁺): m/e 1866 (M⁺). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹. UV (MeOH): λ 360 (ϵ 28 434).

Example 10

SYNTHESIS OF A CYANOCOBALAMIN/LYSOSOMOTROPIC COMPOUND (STREPTOMYCIN) RECEPTOR MODULATING AGENT

This example demonstrates coupling of streptomycin to a cyanocobalamin or cobalamin derivative. Streptomycin (21) is conjugated with cyanocobalamin monocarboxylate (2, 3, 4) or a diaminoalkylsuccinate derivative (14, 15, 16) through the use of an oxime coupled linking moiety (FIGURE 7). The linking group, ((3-aminopropyl)aminoxy)acetamide (22) is prepared by reaction of the N-hydroxysuccinimidyl ester of 1,1-dimethylethoxycarbonyl-aminoxyacetic acid (23) (J. Med. Chem. 36:1255-126, 1993) with an excess of diaminopropane in anhydrous THF. The linking group is separated from other compounds in the reaction mixture by preparative chromatography. The linker (1 g) is then mixed with streptomycin (0.5g) in 10 mL of H₂O containing sodium acetate. The aqueous solution is warmed in a H₂O bath for 10 minutes to yield a crude streptomycin-linker adduct (25), that may be purified by chromatography on acid-washed alumina (J. Am. Chem. Soc. 68:1460, 1946). The aqueous solution containing the streptomycin linker adduct (0.15 mmol) is mixed with an aqueous solution of activated cyanocobalamin (2, 3, 4) (0.1 mmol) and EDC (0.5 mmol) is added. The reaction mixture is stirred at room temperature for 24 hours, then run over a reversed-phase preparative chromatography column for purification of the cyanocobalamin-streptomycin receptor modulating agent (26).

Example 11

SYNTHESIS OF A CYANOCOBALAMIN/LYSOSOMOTROPIC COMPOUND (ACRIDINE) RECEPTOR MODULATING AGENT

This example demonstrates the coupling of the vitamin B₁₂ to acridine. Chloroquine, quinacrine and acridine are lysosomotropic dyes that are relatively nontoxic and concentrated as much as several hundredfold in lysosomes. Acridine derivatives may be covalently attached to a targeting moiety (such as cyanocobalamin) by the reaction scheme illustrated in FIGURE 8, method A, or similarly as described in method B. Both reaction schemes produce a cyanocobalamin-acridine conjugate.

Method A: A diamine side chain is first synthesized in a manner analogous to the side chain of quinacrine. Specifically, mono-phthaloyl protected 1,4-diaminobutane (27) is reacted with 6,9-dichloro-2-methoxyacridine (28) in phenol (*J. Am. Chem. Soc.* 66:1921-1924, 1944). The reaction mixture is then poured into an excess of 2 N NaOH and extracted with ether. The ether extract is washed with 1 M NaHCO₃, then H₂O, and dried over MgSO₄. The crude product is recrystallized from H₂O-alcohol. The phthaloyl protecting group is removed using anhydrous hydrazine in MeOH (*Bioconjugate Chem.* 2:435-440, 1991) to yield the aminoacridine, (29). Aminoacridine (29) is then conjugated with vitamin B₁₂ monocarboxylic acid (2, 3, 4) to yield a cyanocobalamin-acridine conjugate (30).

Method B: Acridine derivative (31) (0.098 mmol, 0.045 g) was dissolved in 0.5 mL of trifluoroacetic acid. This solution was stirred at room temperature for 0.5 h. TFA was removed by aspirator vacuum. The residue was dissolved in 5 mL of acetonitrile and was neutralized by few drops of triethylamine. Acetonitrile was then removed by aspirator vacuum. The residue was dissolved in DMSO (10 mL) and cyanocobalamin carboxylic acid-diaminododecane-succinyl derivative (15, 16, 17) (0.098 mmol, 134 mg) was added followed by triethylamine (12 μ L). The reaction mixture was then stirred at room temperature for 24 h. (HPLC monitored), and evaporated to dryness. The residue was digested with 100 mL of acetone and the solvent was decanted yielding a cyanocobalamin-acridine conjugate (32). Yield: 120 mg (62%). mp 182-188 °C.

¹H NMR (MeOH-d₄, δ): 0.43 (s, 3H, C-20 CH₃); 1.17 (s, 4H, C-46 CH₃); 1.23 (d, 4H, Pr₃ CH₃); 1.29 (s, 24H); 1.36 (br s, 6H); 1.4 (s, 6H); 1.65 (m, 2H); 1.87 (s, 4H); 2.05 (m, 2H); 2.25 (s, 6H, B10 & B11 CH₃); 2.35 (m, 3H); 2.4 (d, 5H); 2.44 (d, 2H); 2.55 (d, 10H); 2.64 (s, 5H); 2.8-2.9 (m, 8H); 3.1-3.15 (m, 6H); 3.3 (s, 6H); 3.4 (m, 1H); 3.65 (m, 2H); 3.75 (d, 1H); 3.9 (d, 1H); 3.98 (s, 2H); 4.0 (m, 2H); 4.1 (d, 1H); 4.16 (m, 1H); 4.3 (m, 1H); 4.48 (m, 1H); 4.6 (m, 2H); 6.0 (s, 1H, C-10); 6.3 (d, 1H, R₁); 6.5 (s, 1H, B4); 7.1 (s, 1H, B2); 7.2 (s, 1H, B7); 7.3 (t, 1H); 7.4 (dd, 1H); 7.6 (dd, 1H); 7.7 (2dd, 2H); 7.8 (d, 1H); 7.9 (d, 1H); 8.4 (d, 1H).

Example 12

SYNTHESIS OF A CYANOCOBALAMIN/LYSOSOMOTROPIC COMPOUND (AMIKACIN) RECEPTOR MODULATING AGENT

This example demonstrates conjugation of amikacin to a cyanocobalamin molecule to form a cyanocobalamin-amikacin conjugate. A reaction scheme for the

conjugation is depicted in FIGURE 6. As noted above, chemical moieties that are retained subcellularly within lysosomes are termed lysosomotropic. Aminoglycosides are lysosomotropic compounds, and thus may be used as rerouting moieties of this invention. The primary long chain amine on the hydroxyaminobutyric acid side chain of the aminoglycoside, amikacin, is preferentially reactive. Specifically, amikacin (33) (Sigma Chemical Co., St. Louis), is reacted with a vitamin B₁₂ monocarboxylate (2, 3, 4) in the presence of EDC. A cyanocobalamin-amikacin conjugate (34) is then separated and purified by reverse-phase LC chromatography under conditions noted above.

Example 13

CYANOCOBALAMIN MONOCARBOXYLIC ACID DIAMINODODECANE CONJUGATE DIMER: ISOPHTHALOYL DICHLORIDE CROSS-LINKING

This example demonstrates the production of a cyanocobalamin dimer suitable for use as a cross-linking receptor modulating agent. Cross-linking of receptors in some receptor systems is sufficient to cause a rerouting of cell surface receptors to lysosomes for degradation, rather than their normal pathway of receptor recycling.

To a solution of cyanocobalamin monocarboxylic acid diaminododecane conjugate (8, 9, 10) (0.192 mmol, 0.300 g) in DMF (30 mL), was added triethylamine (18 μ L). Isophthaloyl dichloride (35) (0.096 mmol, 0.0195 g) was added over a period of 10-15 min. The reaction mixture was stirred at 55-60°C for 48 h (HPLC monitored) and evaporated to dryness. The solid residue was dissolved in 20 mL of methanol:H₂O (7:3) and applied to a reverse-phase C-18 column (500 mm x 25 mm, Alltech, 150 psi) that was developed with the same solvent. RAININ Rabbit-plus peristaltic pumping system was used with a DYNAMAX (model UV-1) UV visible absorbance detector; the eluant was collected with an automatic fraction collector. The fractions containing the final product (HPLC monitored) were evaporated to dryness.

b-acid dimer (36): yield 96 mg (30%), mp 217-220°C with decomposition, ¹H NMR (D₂O, δ) 0.43 (s, 6H, C-20 CH₃); 1.18 (s, 8H); 1.3 (m, 36H); 1.37 (m, 12H); 1.46 (s, 10H); 1.6 (m, 8H); 1.9 (d, 12H); 2.05 (m, 10H); 2.2 (d, 16H, B10 & B11 CH₃); 2.35 (m, 8H); 2.6 (d, 18H); 2.8-3.0 (m, 16H); 3.15 (m, 6H); 3.3 (s, 8H); 3.37 (m, 14H); 3.6 (m, 4H); 3.76 (m, 2H); 3.9 (d, 2H); 4.07 (m, 2H); 4.12 (m, 2H); 4.18 (m, 2H); 4.3 (m, 2H); 4.5 (m, 2H); 4.6 (s, 2H); 4.68 (m, 2H); 6.0 (s, 2H, 2C-10); 6.26 (d, 2H, 2R1); 6.6 (s, 2H, 2B4); 7.1 (s, 2H, 2B2); 7.25 (s, 2H, 2B7); 7.54 (t, 1H); 7.95 (d, 2H); 8.25 (s, 1H); MS (FAB⁺): m/e 3208. IR

(KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm^{-1} ; UV: λ_{360} (ϵ_{42} 380).

e-acid dimer (37): yield 121 mg (38%), mp 220-222°C with decomposition, ^1H NMR (D_2O , δ) 0.43 (s, 6H, C-20 CH_3); 1.17 (s, 8H); 1.22 (d, 13H); 1.29 (s, 45H); 1.36 (d, 22H); 1.44 (s, 10H); 1.6 (m, 8H); 1.87 (s, 8H); 2.04 (m, 10H); 2.25 (s, 12H, B10 & B11 CH_3); 2.36 (m, 8H); 2.55 (d, 20H); 2.8 (m, 8H); 3.15 (m, 8H); 3.29 (s, 10H); 3.36 (m, 14H); 3.6 (m, 4H); 3.73 (m, 2H); 3.9 (d, 2H); 4.07 (m, 2H); 4.12 (m, 2H); 4.16 (m, 2H); 4.3 (m, 2H); 4.5 (m, 2H); 4.6 (s, 2H); 4.66 (m, 2H); 6.0 (s, 2H, 2C-10); 6.26 (d, 2H, 2R1); 6.6 (s, 2H, 2B4); 7.1 (s, 2H, 2B2); 7.25 (s, 2H, 2B7); 7.54 (t, 1H); 7.93 (d, 2H); 8.25 (s, 1H); MS (FAB^+): m/e 3208. IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm^{-1} . UV (MeOH): λ_{360} (ϵ_{33} 854).

d-acid dimer (38): yield 96 mg (30%), mp 225-228°C with decomposition, ^1H NMR (D_2O , δ) 0.43 (s, 6H, C-20 CH_3); 1.16 (s, 8H); 1.29 (m, 36H); 1.35 (d, 12H); 1.44 (s, 10H); 1.53 (m, 6H); 1.6 (m, 8H); 1.85 (s, 12H); 2.03 (m, 8H); 2.25 (d, 12H, B10 B11 CH_3); 2.33 (m, 8H); 2.54 (d, 20H); 2.8 (m, 8H); 3.13 (m, 8H); 3.28 (s, 12H); 3.35 (m, 12H); 3.6 (m, 4H); 3.73 (m, 2H); 3.9 (d, 2H); 4.07 (m, 2H); 4.12 (m, 2H); 4.16 (m, 2H); 4.3 (m, 2H); 4.5 (m, 2H); 4.64 (m, 2H); 4.7 (s, 2H); 6.0 (s, 2H, 2C-10); 6.26 (d, 2H, 2R1); 6.6 (s, 2H, 2B4); 7.1 (s, 2H, 2B2); 7.25 (s, 2H, 2B7); 7.54 (t, 1H); 7.93 (d, 2H); 8.25 (s, 1H); MS (FAB^+): m/e 3208. IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm^{-1} . UV (MeOH): λ_{360} (ϵ_{31} 747).

Example 14

CYANOCOBALAMIN MONOCARBOXYLIC ACID DIAMINODODECANE

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CONJUGATE DIMER: ETAC CROSS-LINKING

This example serves to illustrate synthesis of a bivalent receptor modulating agent using a heterotrifunctional cross-linker. The reaction scheme for this synthesis is depicted in FIGURE 9. The heterotrifunctional cross-linker is formed an ETAC reagent (Bioconjugate Chem. 1:36-50, 1990; Bioconjugate Chem. 1:51-59, 1990; J. Am. Chem. Soc. 101:3097-3110, 1979). Bivalency, in addition to enhancing affinity of binding, also imparts the ability to cross-link neighboring receptors and trigger endocytosis. The bivalent "arms" of the agent may be lengthened with peptide or other linking molecules to enable simultaneous binding of both "arms". In the case of vitamin B₁₂, this may be assessed by gel filtration. If the linkers allow simultaneous interaction, there will be 2 moles of TcII for every mole of ETAC dimer present in a

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single peak of 80,000 m.w. (versus 40,000 m.w. of monomeric TcII). Simultaneous binding of 2 moles of TcII will then have the potential for bivalent binding to cell surface receptor. This can be tested by comparing the affinity of monomer and dimer binding to receptor. While the bivalent agent can be synthesized to include any rerouting moiety of this invention that enhances lysosomal targeting and retention, the compound tyramine, useful for radio-labeling is disclosed for the purpose of illustration.

Referring to FIGURE 9, carboxy-ETAC (39) is prepared by the method of Liberatore et al. (Bioconjugate Chem. 1:1990). The carboxy-ETAC is converted to its acid chloride by reaction in thionyl chloride. Addition of amine (40) gives the amine-ETAC adduct (41). Reaction of amine-ETAC (1 mmol) in CH₃CN with 1 M aqueous cysteamine (10 mmol) is conducted by stirring at room temperature for 24 h. This compound is reduced with NaCNBH₃ under acidic conditions. The crude amine-ETAC-cysteamine adduct (42) is purified by reverse-phase LC, using conditions noted above. A vitamin B₁₂ monocarboxylate (2, 3, 4) is conjugated with tyramine-ETAC-cysteamine compound by reaction with EDC in H₂O. The resultant vitamin B₁₂-ETAC-tyramine dimer (43) is purified by reverse-phase LC, using conditions described above.

Example 15

20 **CYANOCOBALAMIN MONOCARBOXYLIC ACID DIAMINODODECANE**

CONJUGATE DIMER: ISOPHTHALATE CROSS-LINKING WITH BIOTIN MOIETY

This example illustrates the synthesis of a bivalent receptor modulating agent that is additionally coupled to a biotin moiety (44). Further modification can be obtained by coupling of this molecule with an avidin or streptavidin moiety.

25 Reaction Step A: Biotin (12.3 mmol, 3 g) was dissolved in warm (bath temperature 70°C) DMF (60 mL) under argon atmosphere. It was then cooled to ambient temperature and DCC (13.5 mmol, 2.79 g) was added, followed by tetrafluorophenol (24.6 mmol, 4.08g). The reaction mixture was then cooled to 0°C and stirred for 0.5 h. It was then brought back to ambient temperature and stirred for
30 another 4-5 h. The reaction mixture was filtered and the filtrate was evaporated to dryness. The precipitate was washed with acetonitrile (50 mL) and was filtered to yield 5 g (98%) of white solid (45).

¹H NMR (DMSO, δ): 1.4 (m, 2H); 1.7 (m, 2H); 2.5 (t, 2H); 2.8 (t, 2H); 3.1 (m, 1H); 4.1 (m, 1H); 4.3 (m, 1H); 6.4 (d, 2H); 7.9 (m, 1H).

Reaction Step B: 6-aminocaproic acid (46) (7.5 mmol, 0.99g) was dissolved in H₂O (75 mL). Triethylamine (0.5 mL) was added followed by a solution of TFP ester of biotin (5 mmol, 1.96 g) in warm acetonitrile (300 mL). The reaction was stirred overnight at room temperature. It was then filtered, washed with H₂O (50 mL) and dried on high vacuum. Yield: 0.870 g (47%). The filtrate was evaporated to dryness. The residue was taken in boiling acetonitrile (75 mL) and was filtered, washed with hot acetonitrile. The solid (47) was dried on high vacuum to give 0.6 g, for a total yield of 1.47 g (79%).

¹H NMR (DMSO-d₆, δ): 1.2-1.6 (m, 8H); 2.0 (t, 2H); 2.2 (t, 2H); 2.5 (dd, 2H); 2.8 (dd, 2H); 3.1 (m, 3H); 4.1 (m, 1H); 4.3 (m, 1H); 6.4 (d, 2H); 7.7 (m, 1H).

Reaction Step C: Biotin conjugated caproic acid (47) (2.68 mmol, 1 g) was dissolved in DMSO (50 mL). Triethylamine (0.4 mL) was added followed by TFP acetate (4.02 mmol, 1.05 g). The reaction mixture was then stirred at room temperature for 15-20 min (HPLC monitored). It was then evaporated to dryness. The residue was washed with ether and dichloromethane and dried on high vacuum (48). Yield: 1.24 g (89%).

¹H NMR (DMSO-d₆, δ): 1.2 (t, 2H); 1.3-1.7 (m, 5H); 2.1 (t, 2H); 2.6 (dd, 2H); 2.8 (m, 4H); 3.1 (m, 4H); 4.2 (m, 1H); 4.4 (m, 1H); 6.4 (d, 2H); 7.8 (t, 1H); 8.0 (m, 1H).

Reaction Step D: TFP ester of biotin-caproic acid (48) (0.67 mmol, 0.35 g) was dissolved in DMF (40 mL). Triethylamine (80 μL) was added followed by aminoisophthalic acid (1.005 mmol, 0.182 g). The reaction was stirred at room temp. for 8 days (HPLC monitored) while adding triethylamine (80 μL) every after 24 h. It was then evaporated to dryness. The residue was then applied to a column of silica and was initially eluted with acetonitrile (450 mL). It was then eluted with methanol, 20 mL of fractions were collected, at the fraction 2 the solvent was changed to DMF. The fractions containing the final product (HPLC monitored) were evaporated to dryness (49) to yield 230 mg (65%).

¹H NMR (DMSO-d₆, δ): 1.3-1.7 (m, 8H); 2.1 (t, 2H); 2.3 (t, 2H); 2.6 (m, 2H); 2.8 (m, 2H); 3.1 (m, 3H); 4.1 (m, 1H); 4.3 (m, 1H); 6.4 (d, 2H); 7.8 (t, 1H); 8.1 (m, 1H); 8.46 (s, 2H).

Reaction Step E: Biotin-caproic acid-isophthalic acid (49) (0.376 mmol, 200 mg) was dissolved in DMF (30 mL) under argon atmosphere. TFP acetate (0.94 mmol, 241 mg) was added by double-ended needle, followed by triethylamine (112 μ

L). The reaction was then stirred at room temp. for 24 h (HPLC monitored). It was then evaporated to dryness. The light brownish oil was taken in ether, solid was filtered and was washed with ether (50 mL) (50) to yield 250 mg (86%).

¹H NMR (DMSO-d₆, δ): 1.3-1.7 (m, 8H); 2.1 (t, 2H); 2.3 (t, 2H); 2.6 (m, 2H); 2.8 (m, 2H); 3.1 (m, 3H); 4.2 (m, 1H); 4.4 (m, 1H); 6.4 (d, 2H); 7.8 (t, 1H); 8.1 (m, 2H); 8.57 (s, 1H); 8.9 (s, 2H).

Reaction Step F: In a solution of cyanocobalamin carboxylic acid - diaminododecane conjugate (8, 9, 10) (0.130 mmol, 0.2 g) in a mixture of DMF:H₂O (3:1) (40 mL) triethylamine (12 μL) was added. DiTFP ester of biotin-caproic acid-isophthalic acid (50) (0.065 mmol, 0.050 g) was added over a period of 5-10 min. The reaction mixture was stirred at room temperature for 3 h (HPLC monitored). It was then evaporated to dryness. The residue was digested with 100 mL of acetone and the solvent was decanted to yield 230 mg (62%) (51) mp 195-198°C with decomposition.

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Example 16

CYANOCOBALAMIN MONOCARBOXYLIC ACID DIAMINODODECANE CONJUGATE

DIMER: ISOPHTHALATE CROSS-LINKING WITH PARA-IODOBENZOYL MOIETY

This is an example of a bivalent receptor modulating agent that is also conjugated to a *para*-iodobenzoyl moiety.

Reaction Step A: A 5g (28 mmol) quantity of 5-aminoisophthalic acid (52) was dissolved in 30 mL 1N NaOH and placed in an ice/water bath. To the cold solution was added 7.5g (28 mmol) 4-iodobenzoyl chloride (52) in 60 mL of acetonitrile, dropwise. The thick white precipitate was then stirred for 10 minutes before removing the ice/water bath and allowing the mixture to stir an additional 10 minutes. The reaction mixture was adjusted to pH 4 with acetic acid and the resulting solid collected. This solid was then dissolved in 30 mL 1N NaOH and washed with ether (2 x 50 mL). The resulting aqueous solution was filtered and acidified to pH 4 with acetic acid. The white precipitate was then collected and dried on high vacuum to yield .6 g (99+%) of (54). mp >300°C; IR (Nujol, cm⁻¹) 3570(m), 3300(m), 1645, 1580(m), 1525(m), 760(m); ¹H NMR (DMSO-d₆, δ), 8.51 (2H, d, J = 0.7 Hz), 8.27 (1H, s), 7.94 (2H, d, J = 4.2 Hz), 7.84 (2H, d, J = 4.1 Hz).

Reaction Step B: A 5g (12.2 mmol) quantity of 5-[N-(*p*-iodobenzoyl)amino]-isophthalic acid (54) was suspended in 100 mL anhydrous ethyl acetate. To this was added 12.5g (73 mmol) 2,3,5,6-tetrafluorophenol (55) followed by 5g (24.2 mmol) 1,3-dicyclohexylcarbodiimide. This suspension was then stirred at room temperature

for 3 days before filtering off the solid and washing with an additional 20 mL of ethyl acetate. The filtrate was then evaporated to dryness. The resulting sticky white solid was suspended in 50 mL acetonitrile and stirred for 30 minutes. Filtering yielded 3.75g of white solid (43%) (56). mp 250-251°C; IR (Nujol, cm⁻¹) 3220(m), 3060(m), 1750, 1655, 1520, 1485, 1330, 1195, 1110, 1085, 955(m), 945(m); ¹H NMR (DMSO-d₆, δ), 9.06 (2H, d, *J* = 0.7 Hz), 8.57 (1H, t, *J* = 1.4 Hz), 8.04 (2H, m), 7.94 (2H, d, *J* = 4.2 Hz), 7.81 (2H, d, *J* = 4.3 Hz).

Reaction Step C: To a solution of cyanocobalamin carboxylic acid-diaminododecane conjugate (56) (0.192 mmol, 0.3 g) in a mixture of DMF:H₂O (3:1) (40 mL) was added triethylamine (0.018 mL). To this solution, DiTFP ester of 5-[N-(p-iodobenzoyl)amino]-isophthalic acid (57) (0.096 mmol, 0.068 g) was added over a period of 5-10 min. The reaction mixture was stirred at room temperature for 4-5 h (HPLC monitored). It was then evaporated to dryness. The solid residue was dissolved in 20 mL of methanol:H₂O (8:2) and applied to a reverse-phase C-18 column (500 mm x 25 mm, Alltech, 150 psi) that was developed with the same solvent. RAININ Rabbit-plus peristaltic pumping system was used with a DYNAMAX (model UV-1) UV visible absorbance detector; the eluant was collected with an automatic fraction collector. The fractions containing the final product (HPLC monitored) were evaporated to dryness.

b-acid dimer (58): yield: 280 mg (76%), mp 230-233°C with decomposition, ¹H NMR (D₂O, δ) 0.43 (s, 6H, C-20 CH₃); 1.19 (s, 8H); 1.3 (m, 36H); 1.37 (d, 12H); 1.46 (s, 10H); 1.63 (m, 8H); 1.87 (s, 12H); 2.05 (m, 10H); 2.27 (d, 16H, B10 & B11 CH₃); 2.35 (m, 8H); 2.6 (d, 18H); 2.8 (s, 8H); 3.0 (s, 10H); 3.15 (m, 8H); 3.3 (d, 8H); 3.37 (m, 14H); 3.6 (m, 2H); 3.68 (d, 2H); 3.76 (m, 2H); 3.9 (d, 2H); 4.07 (m, 2H); 4.12 (m, 2H); 4.18 (m, 2H); 4.3 (m, 2H); 4.5 (m, 2H); 4.64 (m, 4H); 6.0 (s, 2H, 2C-10); 6.26 (d, 2H, 2R₁); 6.6 (s, 2H, 2B4); 7.1 (s, 2H, 2B2); 7.25 (s, 2H, 2B7); 7.7 (d, 2H); 7.9 (d, 2H); 7.99 (d, 1H); 8.28 (s, 2H); MS (FAB⁺): *m/e* 3453. IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹. UV (MeOH): λ_{360.6} (ε₄₈ 871).

e-acid dimer (59): yield: 258 mg (70%), mp 285-290°C with decomposition, ¹H NMR (D₂O, δ) 0.43 (s, 6H, C-20 CH₃); 1.17 (s, 8H); 1.22 (d, 13H); 1.29 (s, 45H); 1.36 (d, 22H); 1.44 (s, 10H); 1.6 (m, 8H); 1.86 (s, 12H); 2.04 (m, 10H); 2.25 (s, 12H, B10 & B11 CH₃); 2.36 (m, 8H); 2.55 (d, 20H); 2.83 (m, 8H); 3.15 (m, 8H); 3.29 (s, 10H); 3.36 (m, 8H); 3.58 (m, 2H); 3.65 (m, 2H); 3.75 (m, 2H); 3.9 (d, 2H); 4.06 (m, 2H); 4.12 (m, 2H); 4.16 (m, 2H); 4.3 (m, 2H); 4.5 (m, 2H);

4.57 (s, 2H); 4.65 (m, 2H); 6.0 (s, 2H, 2C-10); 6.26 (d, 2H, 2R₁); 6.5 (s, 2H, 2B₄); 7.1 (s, 2H, 2B₂); 7.25 (s, 2H, 2B₇); 7.7 (d, 2H); 7.89 (d, 2H); 7.98 (s, 1H); 8.26 (s, 2H); MS (FAB⁺): m/e 3453. IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹; UV (MeOH): λ 360 (ϵ 41 481).

5 *d*-acid dimer (60): yield 265 mg (72%), mp 253-255°C with decomposition, ¹H NMR (D₂O, δ) 0.43 (s, 6H, C-20 CH₃); 1.16 (s, 8H); 1.22 (d, 12H); 1.33 (m, 36H); 1.43 (s, 10H); 1.53 (m, 6H); 1.6 (m, 8H); 1.86 (s, 12H); 2.03 (m, 8H); 2.25 (d, 12H, B₁₀ & B₁₁ CH₃); 2.33 (m, 8H); 2.54 (d, 20H); 2.8 (s, 4H); 3.0 (s, 4H); 3.28 (s, 10H); 3.35 (m, 8H); 3.58 (m, 2H); 3.65 (m, 2H); 3.73 (m, 2H);
10 3.88 (d, 2H); 4.05 (m, 2H); 4.1 (m, 2H); 4.17 (m, 2H); 4.3 (m, 2H); 4.5 (m, 2H); 4.57 (s, 2H); 4.63 (m, 2H); 6.0 (s, 2H, 2C-10); 6.26 (d, 2H, 2R₁); 6.5 (s, 2H, 2B₄); 7.1 (s, 2H, 2B₂); 7.25 (s, 2H, 2B₇); 7.7 (d, 2H); 7.89 (d, 2H); 7.98 (s, 1H); 8.26 (s, 2H); MS (FAB⁺): m/e 3453. IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹; UV (MeOH): λ 360 (ϵ 48 245).

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Example 17

CYANOCOBALAMIN MONOCARBOXYLIC ACID DIAMINODODECANE CONJUGATE DIMER: ISOPHTHAHATE CROSS-LINKING WITH PARA-(TRI-BUTYLSTANNYL)BENZOYL MOIETY

This is an example of a bivalent receptor modulating agent coupled to a *para*-tri-N-butyl stannyl moiety.

20 Reaction Step A: A 2 g (2.8 mmol) quantity of the diTFP ester of 5-[N-(*p*-iodobenzoyl)amino]-isophthalic acid (57) (as prepared above) was dissolved in 20 mL dry toluene under argon. To this was added 2.8 mL (5.5 mmol) of *bis*(tributyltin) (61) followed by 40 mg (0.04 mmol) tetrakis(triphenylphosphine)palladium (62). The
25 mixture was stirred at room temperature for 15 minutes before heating to 80°C for 2 h. Since the mixture only darkened slightly over the 2 h period, an additional 40 mg of palladium catalyst was added. Within 1 hour the mixture had turned black. After cooling to room temperature, the toluene was removed by rotary evaporation. The resulting black oil (containing solids), was then taken into 20 mL ethyl acetate and
30 dried onto 10 g silica gel (via rotoevaporation). This solid was then added to a 250 g (40 x 3.5 cm) silica gel column and eluanted initially with hexanes containing 5% acetic acid. After 600 mL, the solvent was changed to 90/10 hexanes/ethyl acetate (containing 5% acetic acid). Fractions 14-16 were combined and dried to yield 1.5 g (62%) of white solid (62). mp 120-123°C;

^1H NMR (CDCl_3 , δ), 8.87 (2H, d, $J = 0.7$ Hz), 8.76 (1H, t, $J = 1.6$ Hz), 8.38 (1H, s), 7.84 (2H, d, $J = 4.1$ Hz), 7.62 (2H, d, $J = 4.1$ Hz), 7.07 (2H, m), 1.55 (6H, m), 1.36 (15H, m), 1.11 (6H, m), 0.89 (9H, t, $J = 7.3$ Hz); MS (FAB^+) $\text{M}+\text{H}$ patterns calculated 870 (75.1%), 871 (52.9%), 872 (100%), 873 (41.0%), 874 (21.4%), found 5 870 (82.1%), 871 (55.1%), 872 (100%), 873 (42.1%), 874 (25.2%).

IR (Nujol, cm^{-1}) 1750, 1645, 1520, 1480(m), 1185, 1100, 1085.

Reaction Step B: In a solution of cyanocobalamin carboxylic acid-diaminododecane conjugate (8, 9, 10) (0.065 mmol, 0.1 g) in a mixture of DMF: H_2O (3:1) (40 mL) triethylamine (0.006 mL) was added. DiTFP ester of 5-[N-(p-tributyltin benzoyl) amino]-isophthalic acid (63) (0.0325 mmol, 0.028 g) was added 10 over a period of 5-10 min. The reaction mixture was stirred at room temperature for 12-14 h (HPLC monitored). It was then evaporated to dryness. The residue was digested with 100 mL of acetone and the solvent was decanted.

b-acid dimer (64): yield: 90 mg (70%), mp 208-212°C with decomposition, 15 ^1H NMR (D_2O , δ) 0.43 (s, 6H, C-20 CH_3); 0.88 (t, 9H); 1.15 (t, 12H); 1.19 (s, 8H); 1.3 (m, 36H); 1.37 (d, 12H); 1.46 (s, 10H); 1.6 (m, 8H); 1.9 (s, 12H); 2.05 (m, 10H); 2.28 (d, 16H, B10 & B11 CH_3); 2.35 (m, 8H); 2.6 (d, 18H); 2.8-2.9 (m, 16H); 3.15 (m, 8H); 3.3 (s, 8H); 3.37 (m, 14H); 3.6 (m, 4H); 3.76 (m, 2H); 3.9 (d, 2H); 4.07 (m, 2H); 4.12 (m, 2H); 4.18 (m, 2H); 4.3 (m, 2H); 4.5 20 (m, 2H); 4.68 (m, 2H); 6.0 (s, 2H, 2C-10); 6.26 (d, 2H, 2R₁); 6.6 (s, 2H, 2B4); 7.1 (s, 2H, 2B2); 7.25 (d, 2H, 2B7); 7.6 (d, 2H); 7.9 (d, 2H); 7.99 (br s, 1H); 8.28 (br s, 2H); IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm^{-1} .

e-acid dimer (65): yield: 93 mg (72%), mp >300°C, ^1H NMR (D_2O , δ) 0.43 (s, 6H, C-20 CH_3); 0.88 (t, 9H); 1.12 (t, 12H); 1.17 (d, 8H); 1.22 (d, 13H); 1.29 25 (s, 45H); 1.36 (d, 22H); 1.44 (s, 10H); 1.6 (m, 8H); 1.87 (d, 12H); 2.04 (m, 10H); 2.25 (s, 12H, B10 & B11 CH_3); 2.36 (m, 8H); 2.55 (d, 20H); 2.8 (m, 8H); 3.15 (m, 8H); 3.29 (s, 10H); 3.36 (m, 14H); 3.6 (m, 4H); 3.73 (m, 2H); 3.9 (d, 2H); 4.07 (m, 2H); 4.12 (m, 2H); 4.16 (m, 2H); 4.3 (m, 2H); 4.5 (m, 2H); 4.66 (m, 2H); 6.0 (s, 2H, 2C-10); 6.26 (d, 2H, 2R₁); 6.6 (s, 2H, 2B4); 7.1 (s, 2H, 2B2); 7.25 (s, 2H, 2B7); 7.6 (d, 2H); 7.9 (d, 2H); 7.98 (br s, 1H); 8.28 (br s, 2H); IR (KBr): 3400, 30 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm^{-1} .

d-acid dimer (66): yield: 100 mg (78%), mp 202-205°C with decomposition, ^1H NMR (D_2O , δ) 0.43 (s, 6H, C-20 CH_3); 0.88 (t, 9H); 1.12 (t, 12H); 1.15 (s, 8H); 1.29 (m, 36H); 1.35 (d, 12H); 1.44 (s, 10H); 1.53 (m, 6H); 1.6 (m, 8H); 35 1.86 (d, 12H); 2.03 (m, 8H); 2.25 (d, 12H, B10 & B11 CH_3); 2.33 (m, 8H); 2.54

(d, 20H); 2.8 (m, 8H); 3.13 (m, 8H); 3.28 (s, 10H); 3.35 (m, 10H); 3.6 (m, 4H); 3.73 (m, 2H); 3.9 (d, 2H); 4.05 (m, 2H); 4.1 (m, 2H); 4.17 (m, 2H); 4.3 (m, 2H); 4.5 (m, 2H); 4.6 (m, 2H); 6.0 (s, 2H, 2C-10); 6.26 (d, 2H, 2R1); 6.6 (s, 2H, 2B4); 7.1 (s, 2H, 2B2); 7.25 (s, 2H, 2B7); 7.6 (d, 2H); 7.9 (d, 2H); 7.98 (br s, 1H); 8.28 (br s, 2H); IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm^{-1} .

Example 18

EVALUATION OF THE ABILITY OF VITAMIN B₁₂ RECEPTOR MODULATING AGENTS TO BIND TO TCII

This example serves to demonstrate a competitive binding assay suitable for evaluating the ability of vitamin B₁₂ receptor modulating agents to bind TcII. Binding of the vitamin B₁₂ derivatives to recombinant transcobalamin II was conducted in picomolar concentrations and the percent bound ascertained.

In this competitive binding assay, various B₁₂ derivatives, including vitamin B₁₂ receptor modulating agents, were evaluated for their ability to bind to TcII relative to radiolabeled B₁₂. Varying concentrations of each derivative were incubated with immobilized TcII in the presence of a constant amount of radiolabeled B₁₂. After incubation for 20 minutes at 37°C, the free radiolabeled B₁₂ was separated from the TcII bound tracer by removal of the supernatant. The radioactivity of the supernatant solution was then measured to determine the amount of free radiolabeled B₁₂ present at the end of each competition. By measuring the amount of free radiolabeled B₁₂ for each competition, the ability of each derivative to inhibit radiolabeled B₁₂ binding was determined. A binding curve was then constructed for each B₁₂ derivative where the amount of radiolabeled B₁₂ bound (% radiolabel bound) was correlated with the concentration of derivative present in the original mixture. The more effective the derivative is in binding to TcII, the lower the percent bound radiolabeled vitamin B₁₂.

FIGURE 15 illustrates the binding curve of transcobalamin II to the cyanocobalamin monocarboxylic acids produced in Example 1. AD = cyanocobalamin (1); AL = cyanocobalamin *b*-monocarboxylic acid (2); AM = cyanocobalamin *e*-monocarboxylic acid (3); and AN = cyanocobalamin *d*-monocarboxylic acid (4). The *d*-carboxylate (3) appears to bind nearly as well as cyanocobalamin. Two samples of vitamin B₁₂ were used, one as a known standard and the other as an unknown.

FIGURE 16 illustrates the binding curve of transcobalamin II to the cyanocobalamin diaminododecane adducts (8, 9, 10) and succinate adduct (13) produced in Examples 3 and 4 above. AH = cyanocobalamin *b*-monocarboxylic acid

conjugate diaminododecane (7); AI = cyanocobalamin *e*-monocarboxylic acid conjugate diaminododecane (8); AJ = cyanocobalamin *d*-monocarboxylic acid conj diaminododecane (9); AK = cobalamin *e*-monocarboxylic acid conj diaminododecane, and AE = cyanocobalamin ribose-succinate (11). The *b*-conjugate (17) has the least binding, whereas the *e*-conjugate (18) has intermediate binding, and the *d*-conjugate (19) binds quite well. The biotin conjugate attached to the ribose site (13) appears to bind very well, as does its precursor amino derivative (12). The additional compound studied is of unknown structure, but may have the amine group coordinated with the cobalt atom as the mass spectrum indicates that it has the appropriate mass for (7) minus HCN. It is clear that this unknown compound is not likely to bind TcII.

FIGURE 17 illustrates the binding curve of transcobalamin II to a series of vitamin B₁₂ dimers. Dimer X = *b*-acid dimer with isophthaloyl dichloride (36); dimer Y = *e*-acid dimer with isophthaloyl dichloride (37); dimer Z = *d*-acid dimer with isophthaloyl dichloride (38); dimer A = *b*-acid dimer with *p*-iodo benzoyl isophthaloyl dichloride (58); dimer B = *e*-acid dimer with *p*-Iodo benzoyl Isophthaloyl dichloride (59); and dimer C = *d*-acid dimer with *p*-iodo benzoyl isophthaloyl dichloride (60).

FIGURE 18 illustrates the binding curve of Transcobalamin II to a series of biotinylated vitamin B₁₂ molecules. AA = cyanocobalamin *b*-monocarboxylic acid conjugate diaminododecane and Biotin (17); AB = cyanocobalamin *e*-monocarboxylic acid conjugate diaminododecane and biotin (18); AC = cyanocobalamin *d*-monocarboxylic acid conjugate diaminododecane and Biotin (19); AF = cyanocobalamin ribose-succinate conjugate diaminododecane (13); and AG = cyanocobalamin ribose-succinate conjugate diaminododecane and biotin (20).

EXAMPLE 19

25 **ASSAY FOR BIOLOGICAL ACTIVITY OF VITAMIN B₁₂** **RECEPTOR MODULATING AGENTS**

This example serves to demonstrate the use of an assay to ascertain biological activity of the receptor modulating agents of the present invention.

Receptor down-modulation involves a comparison of treatment of a target cell line such as K562; each sample is treated with vitamin B₁₂ or a vitamin B₁₂ receptor modulating agent at 4°C for 24 hours. Following this period, cells of each sample are separated from a vitamin B₁₂ or a vitamin B₁₂ receptor modulating agent by centrifugation. The cells are then washed and resuspended in phosphate buffered saline containing 2 mM EDTA for a brief period of time not to exceed 15 minutes at

4°C. Then, the cells are washed again and returned to a tissue culture medium at 4°C. The tissue culture medium contains TcII and a radiolabeled TcII/B₁₂ complex. The time course of TcII/B₁₂ binding to the cell receptor is determined by measuring the percent radiolabel bound to the cell at 0, 15, 30, 60, 120, and 240 minutes. Those samples exposed to the vitamin B₁₂ receptor modulating agents of the present invention show significantly reduced TcII/B₁₂ complex binding compared to cells cultured in vitamin B₁₂. Trypsin treated cells reveal any nonspecific binding or uptake of the labeled vitamin B₁₂ on or within the cell.

EXAMPLE 20

10 **METHOD FOR ASSESSING BIOLOGICAL ACTIVITY OF A RECEPTOR MODULATING AGENT**

This example serves to demonstrate a method suitable for assessing the biological activity of a receptor modulating agent of the present invention.

0.2x10⁶ cells/ml K562 cells were cultured in RPMI medium modified by addition of 10 µM MeTHF, 2.7 nM vitamin B₁₂ and 1% human serum. No folate was added. 10 µM *d*-diaminododecane adduct (7) was added and cultured over 9 days at 37°C. 10 µM vitamin B₁₂ cultured under identical conditions as (7) was utilized as a control. The cultures were then independently assessed for proliferation and cell death by Trypan blue exclusion. The results are described in Table 10, below, in terms of the percent cell death.

Table 7

	Control	<i>d</i> -diaminododecane adduct (7)
Proliferation	98%	9 %
Cell Death	8 %	85 %

The receptor modulating agent, in this case *d*-diaminododecane adduct (7), clearly demonstrates the marked biological activity of the receptor modulating agent.

Example 21

25 **SYNTHESIS OF AN ANTI-INFLAMMATORY RECEPTOR MODULATING AGENT**

The synthetic peptide f-met-leu-phe is equivalent to a bacterial cell wall constituent (Biochem. Soc. Trans. 19:1127-9, 1991; Agents Actions Suppl. 35:3-8, 1991; Agents Actions Suppl. 35:11-6, 1991; J Immunol. 146:975-80, 1991). This peptide is recognized by receptors on PMN that can respond by chemotaxis to sites of

local inflammation along a gradient of the peptide. During inflammation, receptor expression can be dramatically increased by mobilizing receptor from intracellular pools. Nonspecific methods used to abrogate this up-regulation also inhibit chemotaxis and presumably the antiinflammatory reaction associated with local inflammation (J. Immunol. 145:2633-8, 1990). The synthesis of a receptor modulation agent useful as an inhibitor of early inflammation is described below.

The peptide f-met-leu-phe-(gly)₃-leu-O-Me is synthesized using tea-bag methodology or solid phase peptide synthesis procedures described by Merrifield et al. (Biochemistry 21:5020-31, 1982) and Houghten (Proc. Nat'l. Acad. Sci. (USA) 82:5131-35, 1985), or using a commercially available automated synthesizer, such as the Applied Biosystems 430 A peptide synthesizer. The peptide-amide is deprotected in 45% trifluoroacetic acid-51% methylene chloride-2% ethanedithiol-2% anisole for 20 minutes, and cleaved from the 4-methylbenzhydrylamine resin using the Tam-Merrifield low-high HF procedure (J. P. Tam et al., J. Am. Chem. Soc. 105:6442-55, 1983). The peptide is then extracted from the resin using 0.1 M ammonium acetate buffer, pH 8, and is lyophilized. The crude peptide is purified using reverse-phase HPLC on a Vydac C-4 analytical column (The Separations Group, Hesperia, Calif.), and a linear gradient of 0.5-1.0%/min. from 100% acetonitrile + 0.1%v/v trifluoroacetate to 100% acetonitrile + 0.1% trifluoroacetate. The HPLC-purified peptide is analyzed by amino acid analysis (R.L. Heinriksen and S.C. Meredith, Anal. Biochem. 160:65-74, 1984) after gas phase hydrolysis (N.M. Meltzer et al., Anal. Biochem. 160:356-61, 1987). The sequence of the purified peptide may be confirmed by Edman degradation on a commercially available sequencer (R.M. Hewick et al., J. Biol. Chem. 15:7990-8005, 1981). The peptide amide is converted to an O-methyl ester (*i.e.*, f-met-leu-phe-(gly)₃-leu-O-Me) by treatment with dimethylformamide (5g/60 mL with 1.3 equivalents of NaHCO₃ in excess methyl iodide (4 equivalents). The mixture is stirred under argon gas at room temperature for 40 hours. If required, the peptide is extracted to dryness with 150 mL of ethyl acetate. The receptor for modulating agent is used to treat PMN, activated with GM-CSF (to increase expression of fMLP receptors). Loss of binding of biotinylated fMLP is compared on fMLP versus f-MLP receptor modulating agent treated cells.

Example 22

SYNTHESIS OF A FUSION PROTEIN RECEPTOR MODULATING AGENT

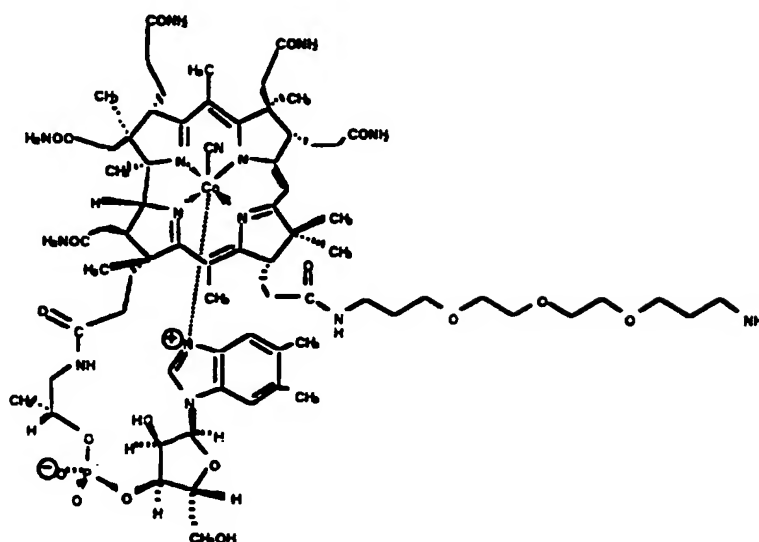
An EGF receptor modulating agent containing a genetically engineered fusion protein is hereby described. Briefly, the C-terminus of a DNA sequence encoding

EGF, or its receptor binding domain, is ligated by conventional procedures (*e.g.*, using T₄DNA ligase) to a DNA sequence corresponding to a GGG spacer. The C-terminus of the EGF-GGG DNA sequence is then fused to the N-terminus of a DNA sequence encoding the conditional, membrane binding peptide KGEAALA(EALA)₄-EALALAA. Alternately, peptide-spacer DNA sequences may be synthesized *in vitro* using standard oligonucleotide synthesis procedures (see, *e.g.*, U.S. Patent Numbers 4,500,707 and 4,668,777). The recombinant EGF peptide DNA sequence is cloned in an *E. coli* expression vector using conventional procedures. *E. coli* strain HB101 is transformed with the fused recombinant DNA sequence and cultured to produce the EGF peptide. The fusion protein is purified from the transformed *E. coli* culture by standard methods, including anti-EGF affinity chromatography. The fusion protein may be eluted from the affinity matrix using standard techniques, such as high salt, chaotropic agents, or high or low pH. Loss of EGF receptor is measured by flow cytometry and mouse monoclonal antibody to EGF receptor.

Example 23

SYNTHESIS OF A VITAMIN B₁₂ DERIVATIVE HAVING A WATER-SOLUBILIZING LINKER

The preparation of a vitamin B₁₂ derivative having a water-solubilizing linker is described. Briefly, the example describes a procedure for the reaction of a cyanocobalamin monocarboxylic acid with 4,7,10-trioxa-1,13-tridecane diamine. The results for the *b*- and *e*-monoacids of cyanocobalamin are described. The reaction product for the *e*-isomer is shown below.



Cyanocobalamin monocarboxylic acid (1.472 mmol, 2 g) and N-hydroxysuccinimide (680 mg) were dissolved in water (100 mL) and 1.456 g of sodium cyanide was added. 4,7,10-Trioxa-1,13-tridecanediamine (36 mmol, 16 g) was then added and the pH was adjusted to 6 with 1 N HCl. N-ethyl-N'-dimethylamino-propyl-carbodiimide-hydrochloride (EDC) (1.136 g) was added and the pH of the solution was readjusted to 5.5. The reaction mixture was then stirred overnight in the dark at room temperature. In 5 intervals of 6 to 14 h 680 mg of N-hydroxysuccinimide and 1.136 g of EDC were added to the solution, readjusting the pH value to 5.5 each time. After a total reaction time of 4 days (HPLC monitored) the solution was evaporated to dryness, the residue was digested with 100 mL of acetone, and the solvent was decanted. The solid residue was dissolved in 50 mL of H₂O and applied to an Amberlite XAD-2 (200 g; 4 cm x 60 cm) column. The column was eluted with 1 L water, then the desired product was eluted with 500 mL methanol. The methanol fractions were evaporated to dryness, and the residue was dissolved in 25 mL of water and was applied to a Dowex Cl⁻ column (100 g, 2.5 cm x 60 cm, acetate form, 200-400 mesh). The final product was eluted using 250 mL water, thereby leaving nonconverted acid bound to the column, that was later eluted with 0.04 mol/L sodium acetate buffer pH 4.7. The fractions containing the final product were evaporated to dryness, then digested with acetone and filtered. The solid obtained was recrystallized from aqueous acetone.

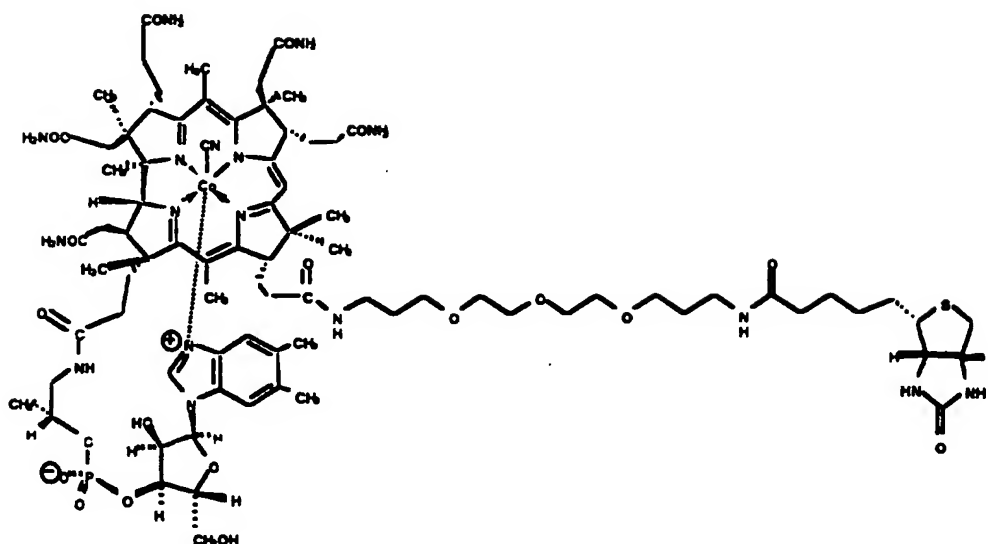
b-isomer: yield: 2 g (87%), mp: 213-217°C with decomposition. ¹H NMR (MeOH-*d*₄, δ): 0.44 (s, 3H); 1.17 (d, 5H); 1.25 (d, 4H); 1.36 (d, 7H); 1.45 (s, 4H); 1.74 (m, 10H); 1.88 (s, 11H); 2.27 (d, 8H); 2.34 (m, 11H); 2.56 (m, 11H); 3.17 (t, 3H); 3.2 (m, 9H); 3.3 (m, 6H); 3.4 (m, 4H); 3.5 (s, 7H); 3.58 (s, 8H); 3.6 (m, 11H); 3.7 (m, 1H); 3.88 (m, 1H); 4.07 (m, 1H); 4.1 (m, 1H); 4.17 (m, 1H); 4.3 (m, 1H); 4.5 (m, 1H); 4.6 (m, 1H); 6.04 (d, 1H); 6.27 (s, 1H); 6.52 (s, 1H); 7.13 (d, 1H); 7.25 (s, 1H). MS (FAB⁺): m/e 1558 (M⁺). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹. UV (H₂O): λ₃₆₁ (ε 17470).

e-isomer: yield: 1.5 g (65%), mp: 112-116°C with decomposition. ¹H NMR (MeOH-*d*₄, δ): 0.44 (s, 3H); 1.18 (s, 3H); 1.25 (d, 5H); 1.37 (d, 8H); 1.45 (s, 4H); 1.74 (m, 10H); 1.88 (s, 11H); 2.28 (d, 7H); 2.3 (m, 15H); 2.56 (d, 11H); 3.17 (t, 3H); 3.2 (t, 4H); 3.3 (m, 11H); 3.4 (m, 4H); 3.5 (s, 7H); 3.58 (d, 3H); 3.6 (m, 5H); 3.7 (m, 1H); 4.0 (m, 1H); 4.1 (d, 1H); 4.19 (m, 1H); 4.3 (m, 1H); 4.5 (d, 1H); 4.6 (m, 1H); 6.05 (d, 1H); 6.27 (s, 1H); 6.57 (s, 1H); 7.1 (d, 1H); 7.25 (s, 1H). MS (FAB⁺): m/e 1558 (M⁺). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹. UV (H₂O): λ₃₆₁ (ε 12818).

Example 24

SYNTHESIS OF A VITAMIN B₁₂/BIOTIN CONJUGATE HAVING A WATER-SOLUBILIZING LINKER

The preparation of a vitamin B₁₂/biotin conjugate having a water-soluble linker is described. Briefly, the vitamin B₁₂ derivative having a water-solubilizing linker prepared as described in Example 23 is treated with an NHS-ester of biotin. The results for the *b*- and *e*-isomers of cobalamin are described. The reaction product for the *e*-isomer is shown below.



In a solution of cyanocobalamin monocarboxylic acid trioxadiazine conjugate (0.193 mmol, 300 mg) in DMF (10 mL), triethylamine (0.193 mmol, 0.027 mL) was added. N-hydroxysuccinimidobiotin (0.232 mmol, 79 mg) was then added and the reaction was stirred overnight. It was then evaporated to dryness. The residue was dissolved in 10 mL of methanol-water (1:1 v/v) and the solution was applied to a reverse-phase column (500mm X 25mm, Alltech, 150 psi) (octadecyl) that was developed with the same solvent. RAININ Rabbit-plus peristaltic pumping system was used with a DYNAMAX (model UV-1) UV visible absorbance detector; the eluant was collected with an automatic fraction collector. The fractions containing the final product (HPLC monitored) were evaporated to dryness.

b-isomer (9): yield: 160 (53%) mp: 195-197°C with decomposition. ¹H NMR (MeOH-*d*₄, δ): 0.44 (s, 3H); 1.17 (s, 3H); 1.23 (s, 4H); 1.35 (s, 6H); 1.43 (s, 4H); 1.6 (m, 4H); 1.73 (s, 8H); 1.86 (s, 4H); 2.05 (m, 4H); 2.17 (s, 2H); 2.26 (s, 7H); 2.35 (s, 3H); 2.55 (m, 10H); 2.88 (m, 4H); 3.2 (m, 6H); 3.5 (s, 6H); 3.57 (s, 3H); 3.6 (s, 5H); 3.75 (d, 1H); 3.87 (d, 1H); 4.0 (s, 1H); 4.1 (d, 1H); 4.16 (s, 1H); 4.28 (s, 2H); 4.49 (m, 2H); 4.66 (m, 1H); 6.03 (s, 1H); 6.26 (s, 1H); 6.55 (s, 1H); 7.11 (s, 1H); 7.24 (s, 1H). MS (FAB⁺): *m/e* 1784 (M⁺). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹. UV (H₂O): λ₃₆₁ (ε 22770).

e-isomer (10): yield: 80 mg (32%), mp: 224-227°C with decomposition. ¹H NMR (MeOH-*d*₄, δ): 0.44 (s, 3H); 1.17 (s, 3H); 1.23 (d, 4H); 1.35 (d, 6H); 1.44 (s, 4H); 1.74 (m, 5H); 1.87 (s, 4H); 2.06 (m, 4H); 2.18 (m, 3H); 2.26 (d, 7H);

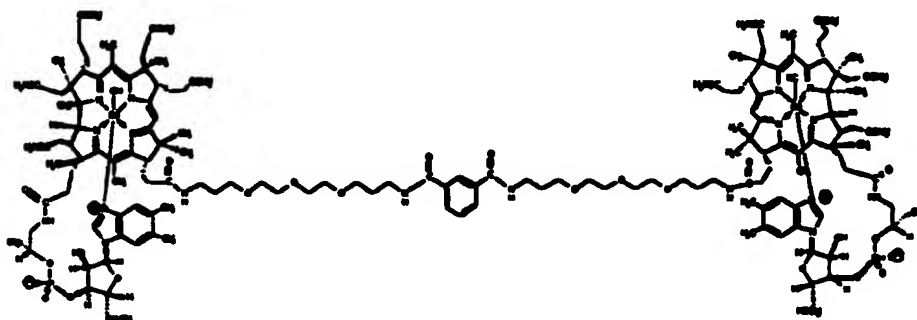
2.35 (s, 3H); 2.55 (d, 9H); 2.9 (m, 4H); 3.24-3.3 (m, 4H); 3.5 (s, 6H); 3.57 (s, 3H); 3.6 (s, 5H); 3.75 (dd, 1H); 3.87 (dd, 1H); 4.0 (m, 1H); 4.1 (d, 1H); 4.17 (t, 1H); 4.28 (m, 2H); 4.47 (m, 1H); 4.5 (m, 1H); 4.66 (m, 1H); 6.04 (s, 1H); 6.27 (d, 1H); 6.56 (s, 1H); 7.11 (s, 1H); 7.24 (s, 1H). MS (FAB⁺): m/e 1784 (M⁺). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹. UV (H₂O): λ 361 (e 22021).

Example 25

SYNTHESIS OF A VITAMIN B₁₂ DIMER HAVING A WATER-SOLUBILIZING LINKER

10 The preparation of two vitamin B₁₂ dimers having water-solubilizing linkers is described. Briefly, the dimers are prepared by coupling the vitamin B₁₂ derivative of Example 23 with either a bifunctional crosslinker or a trifunctional crosslinker.

 A. Isophthaloyl crosslinked dimer. The preparation and results for crosslinking using isophthaloyl dichloride and the *b*-isomer of cyanocobalamin monocarboxylic acid trioxadiazine conjugate are presented. The reaction product for the *e*-isomer is shown below.

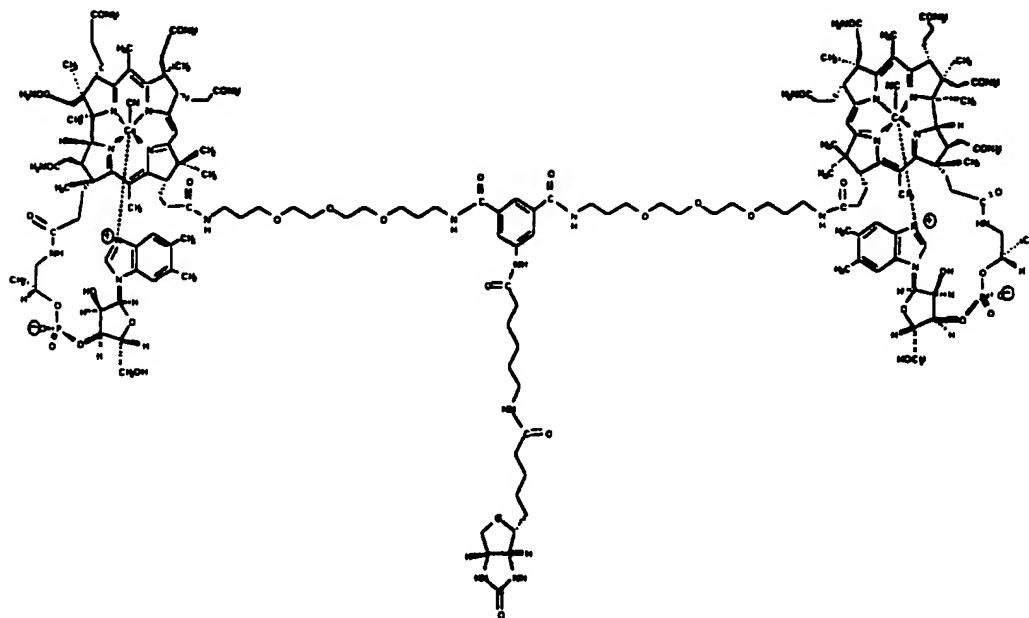


20 In a solution of cyanocobalamin monocarboxylic acid trioxadiazine conjugate (0.193 mmol, 0.300 g) in DMF (20 mL), triethylamine (0.030 mL) was added. Isophthaloyl dichloride (0.096 mmol, 0.0195 g) was added over a period of 10-15 min. The reaction mixture was stirred at room temperature for 4-5 days (HPLC monitored), and triethylamine (0.030 mL) was added every after 24 hours. After
25 evaporating to dryness, the solid was dissolved in 20 mL of methanol:H₂O (1:1) and applied to a reverse-phase column (500 mm X 25 mm, Alltech, 150 psi) (octadecyl) that was developed with the same solvent. A RAININ Rabbit-plus peristaltic pumping system was used with a DYNAMAX (model UV-1) UV visible absorbance

detector, the eluant was collected with an automatic fraction collector. The fractions containing the final product (HPLC monitored) were evaporated to dryness.

b-isomer (11): yield: 100 mg, mp: 195-198°C with decomposition. ¹H NMR (MeOH, δ): 0.44 (s, 6H); 1.18 (s, 6H); 1.25 (d, 7H); 1.31 (t, 20H); 1.36 (s, 14H); 1.45 (s, 8H); 1.74 (m, 20H); 1.88 (d, 15H); 2.27 (s, 11H); 2.37 (m, 22H); 2.56 (d, 20H); 2.85 (s, 5H); 2.99 (s, 2H); 3.2 (m, 18H); 3.3 (m, 12H); 3.4 (m, 10H); 3.5 (s, 14H); 3.58 (s, 18H); 3.6 (s, 30H); 3.9 (d, 4H); 4.0 (d, 2H); 4.1 (d, 2H); 4.18 (d, 2H); 4.3 (m, 2H); 4.5 (m, 2H); 4.7 (m, 2H); 6.0 (s, 2H); 6.28 (s, 2H); 6.56 (s, 2H); 7.1 (s, 2H); 7.25 (s, 2H); 7.56 (m, 1H); 7.8 (d, 2H); 8.3 (s, 1H). IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹. UV (H₂O): λ₃₆₁ (ε 33865).

B. Trifunctional crosslinked dimer. The preparation and results for crosslinking using a trifunctional crosslink and the *b*- and *e*-isomers of the cobalamin derivative are presented. The reaction for the *e*-isomer is shown below.



In a solution of cyanocobalamin carboxylic acid - trioxadamine conjugate (0.193 mmol, 0.3 g) in DMF (15 mL), triethylamine (0.030 mL) was added. DiTFP ester of Biotin-caproic acid-isophthalic acid (0.0965 mmol, 0.079 g) was added over a period of 5-10 min. The reaction mixture was stirred at room temperature for 3-4 days (HPLC monitored), adding triethylamine (0.030 mL) after every 24 hours. It

was then evaporated to dryness. The solid was dissolved in 20 mL of methanol:H₂O (1:1) and applied to a reverse-phase column (500 mm X 25 mm, Alltech, 150 psi) (octadecyl) that was developed with the same solvent. A RAININ Rabbit-plus peristaltic pumping system was used with a DYNAMAX (model UV-1) UV visible absorbance detector; the eluant was collected with an automatic fraction collector. The fractions containing the final product (HPLC monitored) were evaporated to dryness.

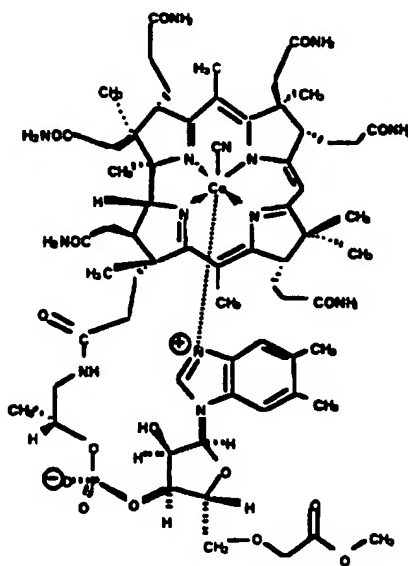
b-isomer (12): yield: mp: 192-195°C with decomposition. IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹.

e-isomer (13): IR (KBr): 3400, 3200, 2950, 2060, 1660, 1570, 1490, 1060 cm⁻¹.

Example 26

SYNTHESIS OF A VITAMIN B₁₂ 5'-O-RIBOSE ETHER DERIVATIVE

The preparation of a cyanocobalamine methyl acetate derivative having a 5-O-ribose ether linkage is described. Briefly, alkylation of cyanocobalamin with methyl bromoacetate provides the ether having the structure shown below.



Preparation of Cyanocobalamine - methylacetate derivative :

Cyanocobalamine (0.15 mmol, 200 mg) was dissolved in 40 mL of DMSO containing 10 g (65 mmol) of methyl bromoacetate and 6.4 mL of pyridine. After 14-16 h at 50-55°C, 500 mL of water was added and the pH of the reaction mixture was adjusted to 6 with 10% KOH. KCN was then added at a final concentration of 0.01 M and the

pH of the solution was readjusted to 6 with 3 N HCl. After 1 h the cyanocobalamin components were desalted by phenol extraction and applied to a 100 g of Dowex Cl⁻ (2.5 cm x 60 cm) column (acetate form 200-400 mesh). Cyanocobalamin was eluanted with water. Methylacetate derivative was eluanted with NaOAc (0.04 M, pH 4.67).

While the preferred embodiment of the invention has been illustrated and described, it will be appreciated that various changes can be made therein without departing from the spirit and scope of the invention.

The embodiment of the invention in that an exclusive property or privilege is claimed are defined as follows:

1. A vitamin B₁₂ receptor modulating agent having a vitamin B₁₂ moiety coupled to a rerouting moiety by a linker, wherein said linker is a water-solubilizing linker or said linker couples to a ribose coupling site on the vitamin B₁₂ moiety.
2. The receptor modulation agent of Claim 1 wherein the linker is a water-solubilizing linker.
3. The receptor modulating agent of Claim 2 wherein the water-solubilizing linker is selected from the group consisting of a polyhydroxy linker, a polyamino linker, and a polyether linker.
4. The receptor modulating agent of Claim 3 wherein the polyhydroxy linker is selected from the group consisting of a glycol, a glycerol, and a polysaccharide linker.
5. The receptor modulating agent of Claim 3 wherein the polyether linker is a polyethylene glycol linker.
6. The receptor modulating agent of Claim 3 wherein the polyether linker is selected from the group consisting of a 4,7,10-trioxa-1,13-tridecanediamine linker and a 3,6-dioxa-1,8-octadecanediamine linker.
7. The receptor modulating agent of Claim 1 wherein the linker is selected from the group consisting of an aminoacid linker and a diamino linker.
8. The receptor modulating agent of Claim 7 wherein the diamino linker is selected from the group consisting of diamino alkyl, diamino alkylaryl, diamino heteroalkyl, and diamino heteroalkylaryl linkers.
9. The receptor modulating agent of Claim 7 wherein the diamino alkyl linker is -NH-(CH₂)_x-NH- wherein x = 2-20.
10. The receptor modulating agent of Claim 7 wherein the aminoacid linker is -NH-(CH₂)_y-CO- wherein y = 3-12.

11. The receptor modulating agent of Claim 1, wherein the linker is covalently coupled to a vitamin B₁₂ coupling site selected from b-, d-, and e- coupling sites.

12. The receptor modulating agent of Claim 1 wherein the linker is covalently coupled to a ribose coupling site on the vitamin B₁₂ molecule.

13. The receptor modulating agent of Claim 12 wherein the ribose coupling site is a 5'-OH coupling site.

14. The receptor modulating agent of Claim 12 wherein the linker is coupled to the vitamin B₁₂ through an ether linkage.

15. The receptor modulating agent of Claim 14 wherein the ether linkage is selected from the group consisting of alkyl ether, aryl ether, benzyl ether, and silyl ether linkages.

16. A vitamin B₁₂ receptor modulating agent having a vitamin B₁₂ moiety coupled to a rerouting moiety by a polyether linker.

17. The receptor modulating agent of Claim 16 wherein the polyether linker is a polyethylene glycol linker.

18. The receptor modulating agent of Claim 16 wherein the polyether linker is covalently coupled to the vitamin B₁₂ ribose 5'-OH coupling site.

19. The receptor modulating agent of Claim 1 wherein said rerouting moiety is selected from the group consisting of lysosomotropic moieties, intracellular polymerizing moieties, peptide sorting sequences, conditional membrane binding peptides, and bi- or multivalent receptor cross-linking moieties, and membrane anchors.

20. A vitamin B₁₂ dimer comprising a first vitamin B₁₂ molecule covalently coupled to a second vitamin B₁₂ molecule by a linker.

21. The receptor modulating agent of Claim 20 wherein the linker is a polyether linker.

22. The receptor modulating agent of Claim 20 wherein the linker is covalently coupled to the vitamin B₁₂ ribose 5'-OH coupling site.

23. A vitamin B₁₂ derivative comprising a first vitamin B₁₂ molecule covalently coupled to a biotin molecule by a linker.

24. The receptor modulating agent of Claim 23 wherein the linker is a polyether linker.

25. The receptor modulating agent of Claim 23 wherein the linker is covalently coupled to the vitamin B₁₂ ribose 5'-OH coupling site.

26. A method for modulating a vitamin B₁₂ receptor, comprising administering an effective amount of a receptor modulating agent according to Claim 1 to a warm-blooded animal such that a vitamin B₁₂ receptor is modulated.

27. A complex comprising a vitamin B₁₂ derivative according to Claim 20 bound to a transcobalamin II.

28. A pharmaceutical composition, comprising a vitamin B₁₂ derivative according to Claim 20 and a suitable pharmaceutical carrier or diluent.

29. A kit for determining the presence or amount of transcobalamin in a sample using a vitamin B₁₂ derivative according to Claim 20.

30. A vitamin B₁₂/avidin conjugate wherein the conjugate comprises four vitamin B₁₂ molecules.

31. A vitamin B₁₂/avidin conjugate wherein the conjugate comprises four vitamin B₁₂ molecules.

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Mechanism of Action

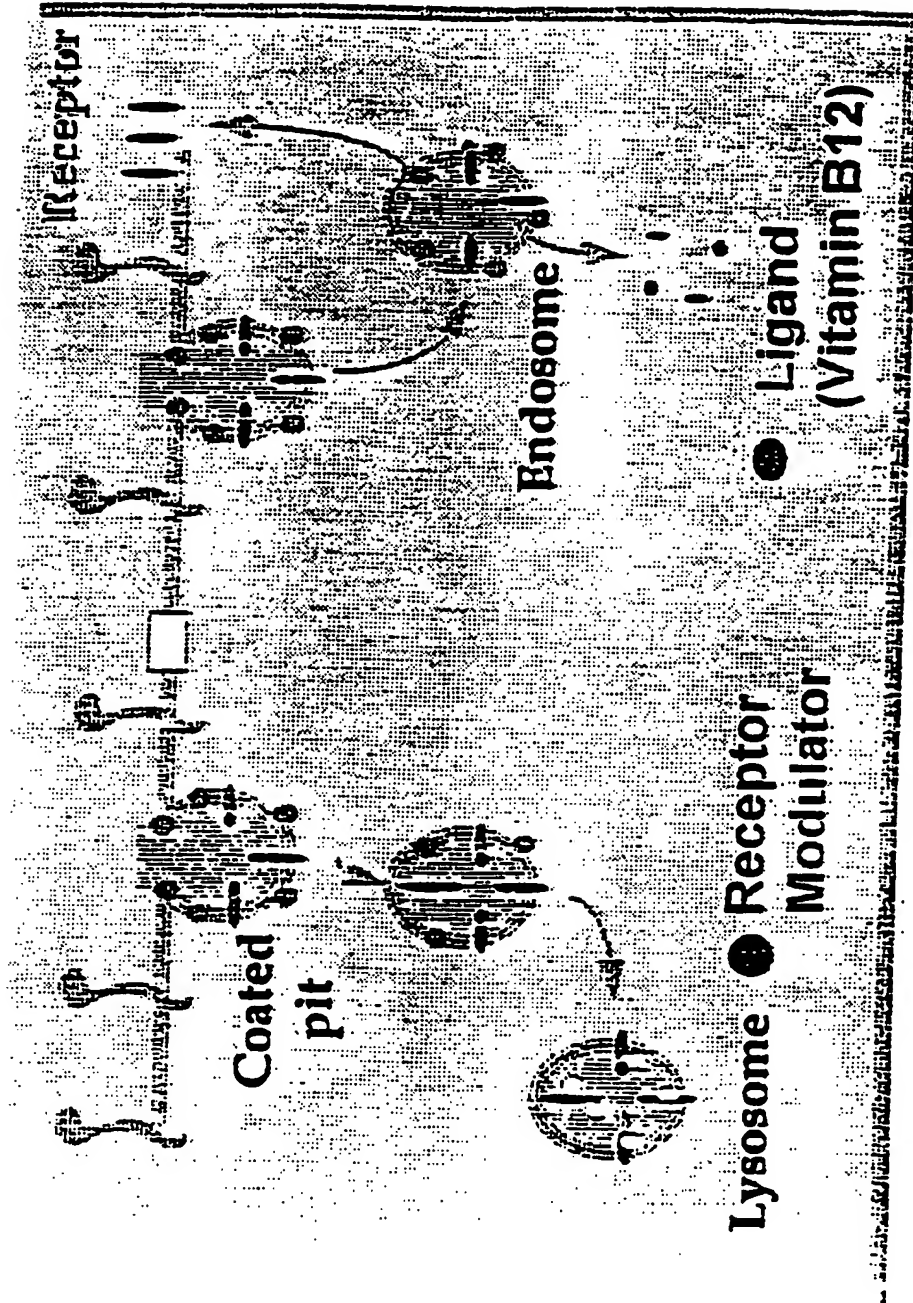
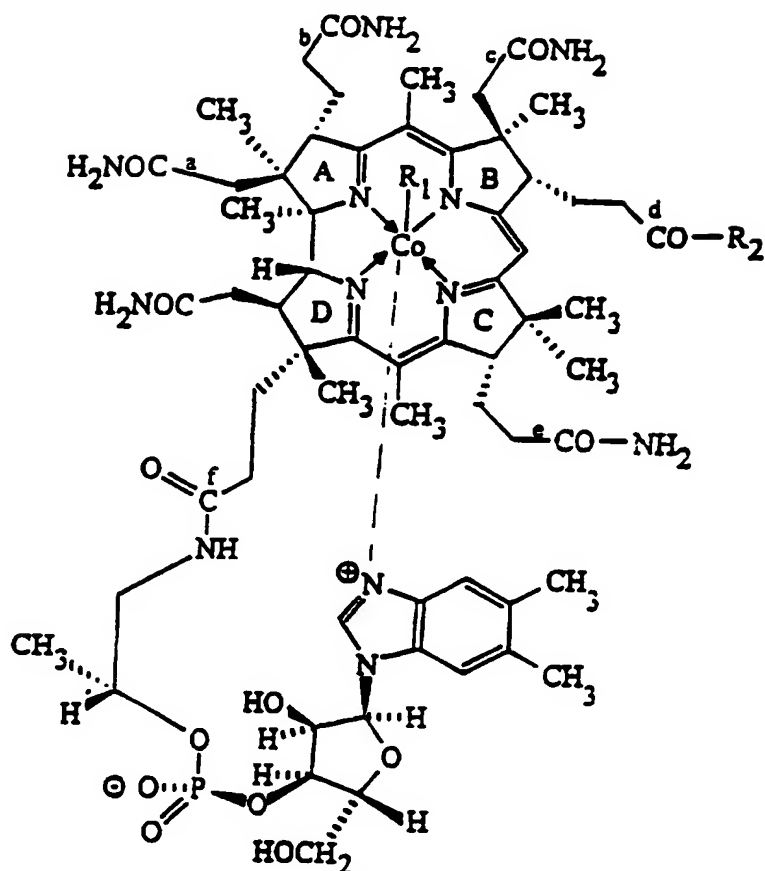


FIGURE 1

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- $R_1 = \text{CN} ; R_2 = \text{NH}_2$ (Cyanocobalamin)
 $R_1 = \text{CN} ; R_2 = \text{OH}$ (Cyanocobalamin -(3)-free acid)
 $R_1 = \text{CN} ; R_2 = \text{HN-CH}_2\text{-CH}_2\text{-CH}_2\text{-CO}_2\text{H}$ (GABA adduct)
 $R_1 = \text{CN} ; R_2 = \text{GABA - Peptide (where GABA = linker)}$
 $R_1 = \text{CN} ; R_2 = \text{Peptide}$
 $R_1 = \text{CN} ; R_2 = \text{HN-(linker)-tyramine-125I}$
 $R_1 = \text{CN} ; R_2 = \text{HN-(linker)-lysosomotropic agent}$
 $R_1 = \text{CN} ; R_2 = \text{HN-(linker)-X-linking agent}$
 $R_1 = \text{CN} ; R_2 = \text{HN-(linker)-biotin}$
 $R_1 = \text{CN} ; R_2 = \text{NH-(CH}_2\text{)}_{12}\text{NH}_2$

Fig. 2

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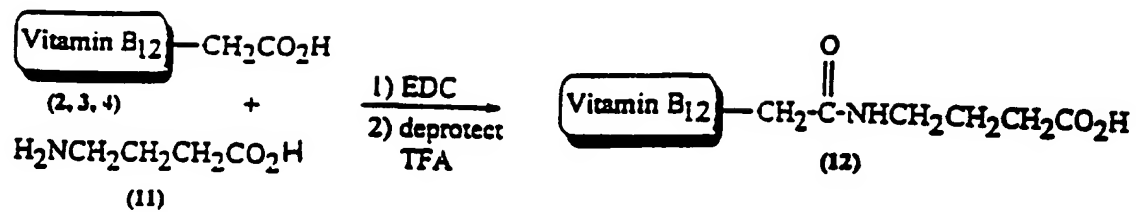


Fig. 3

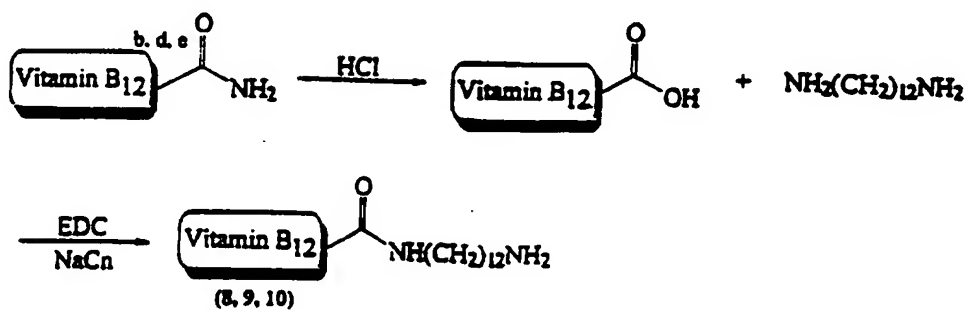


Fig. 4a



Fig. 4b

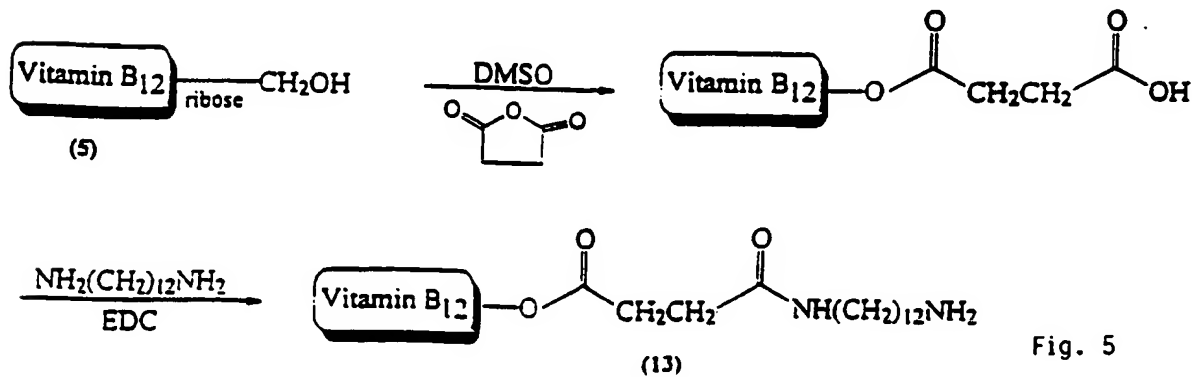
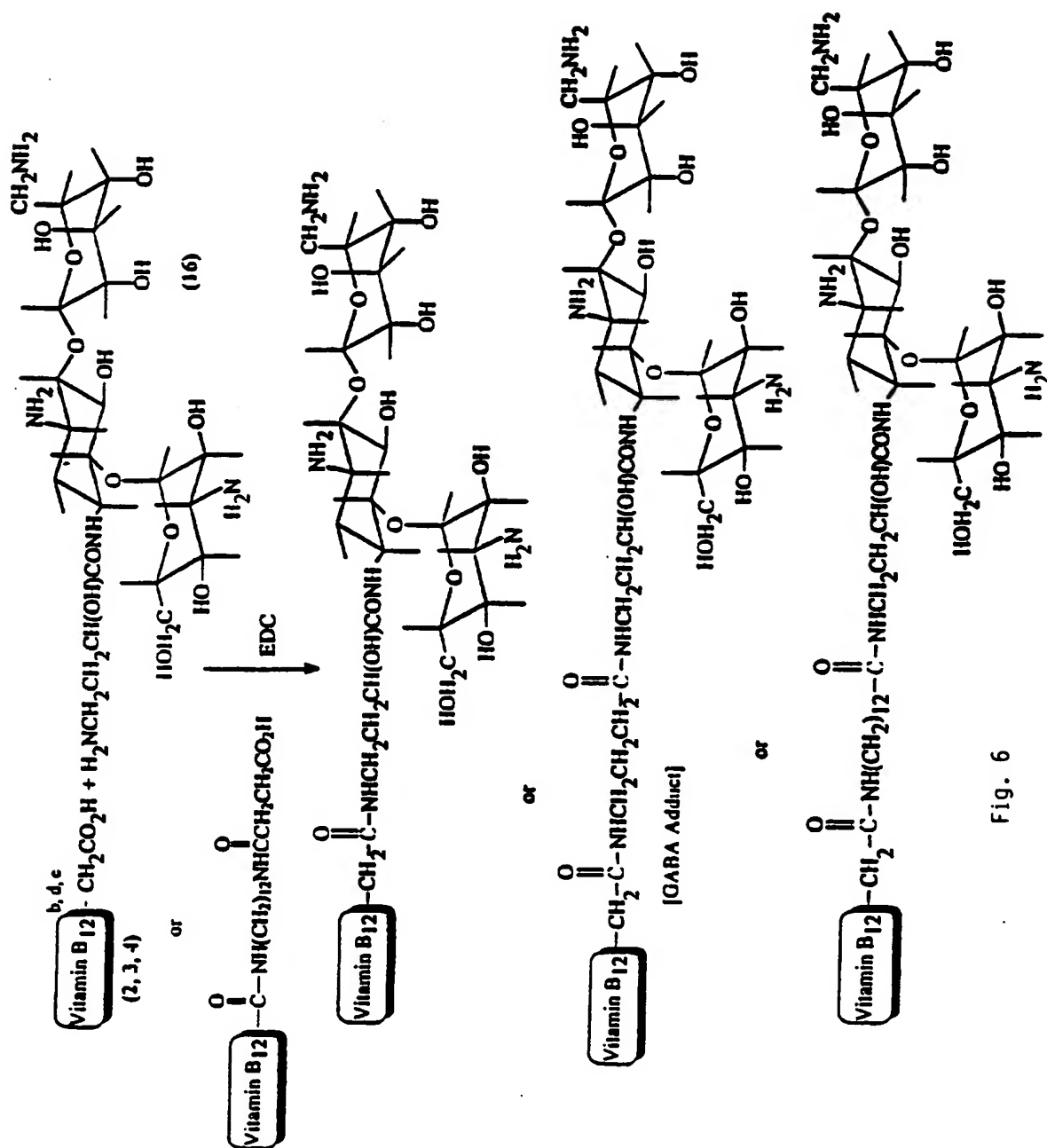


Fig. 5

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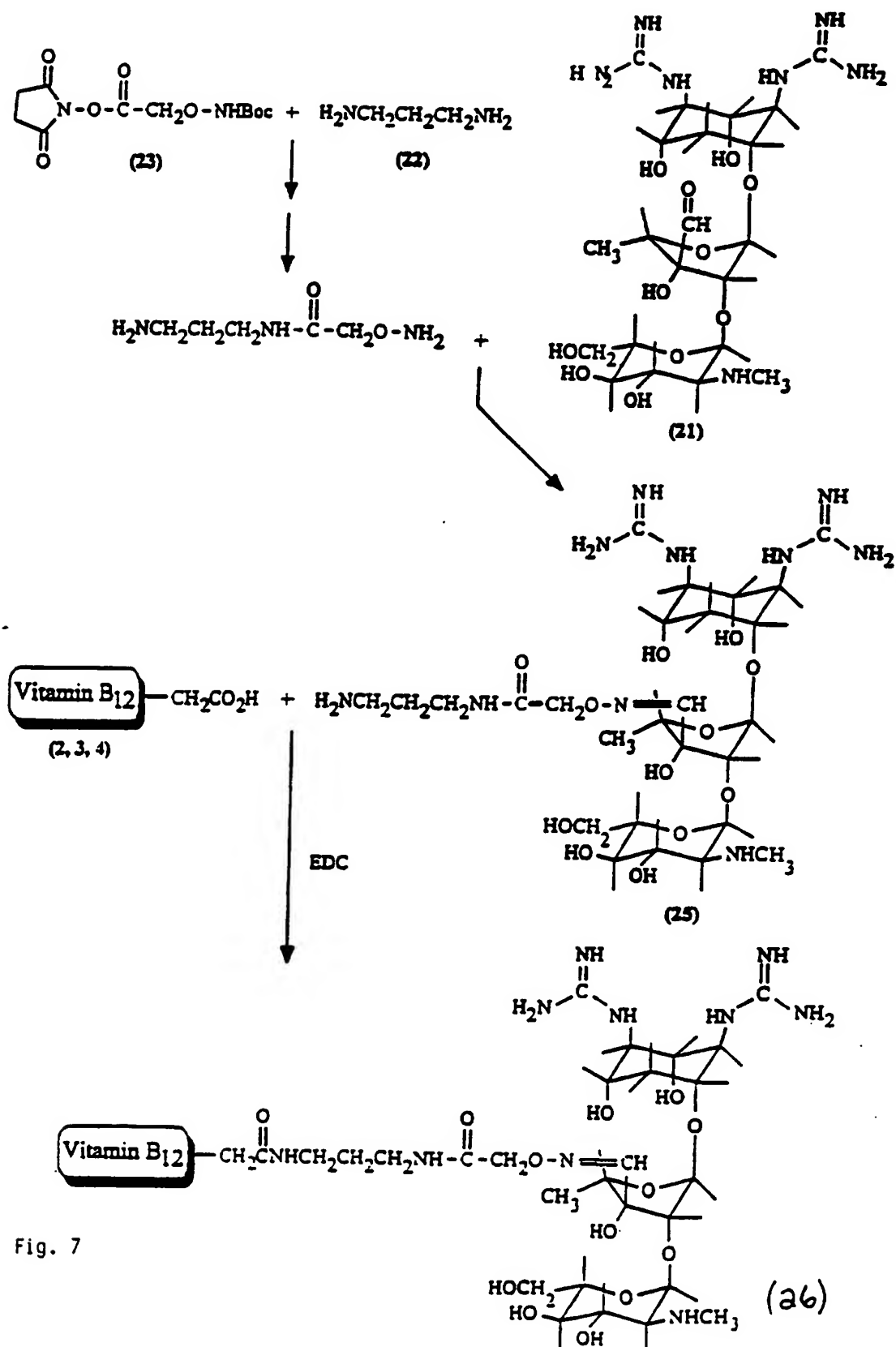


Fig. 7

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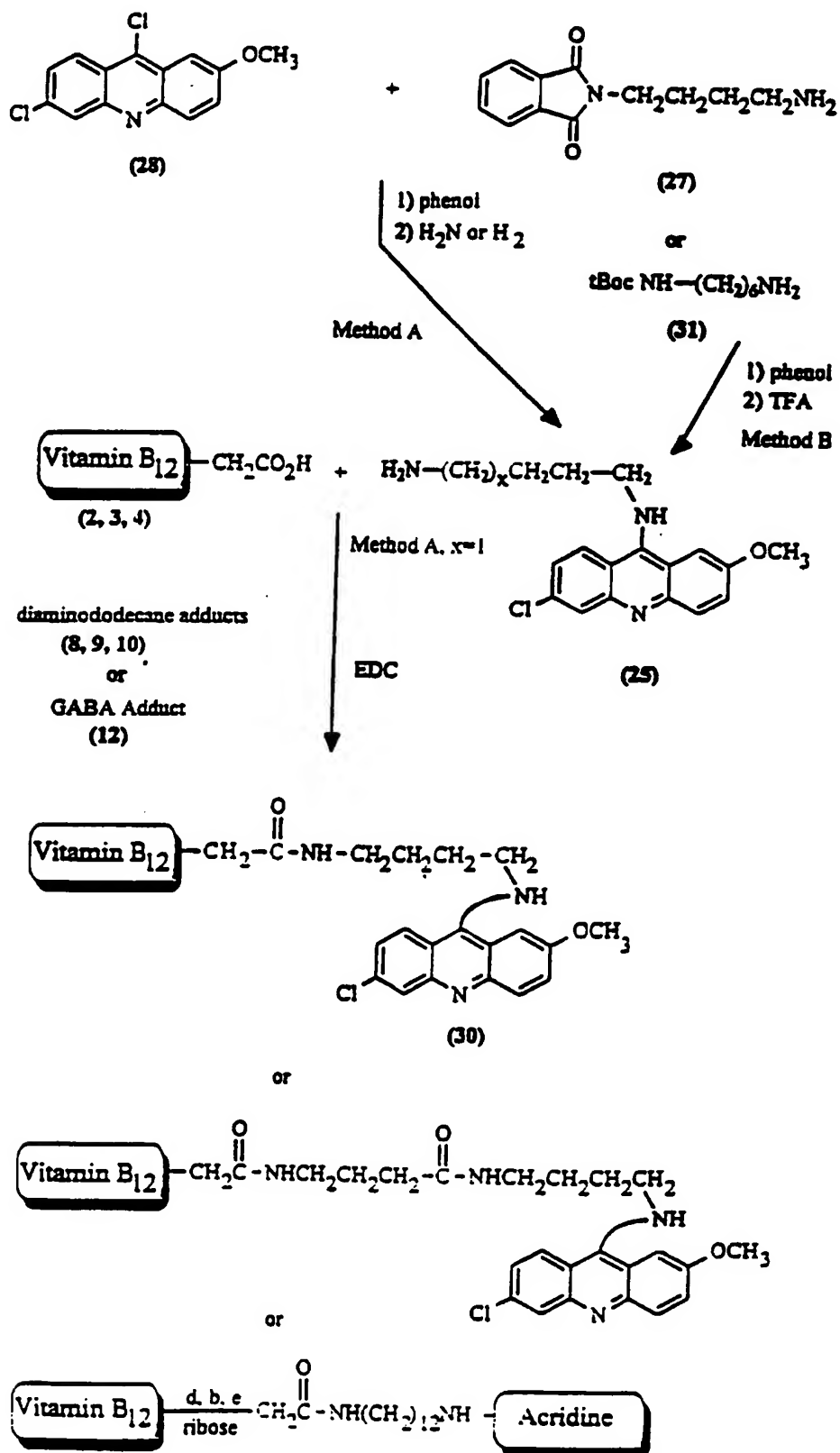


Fig. 8

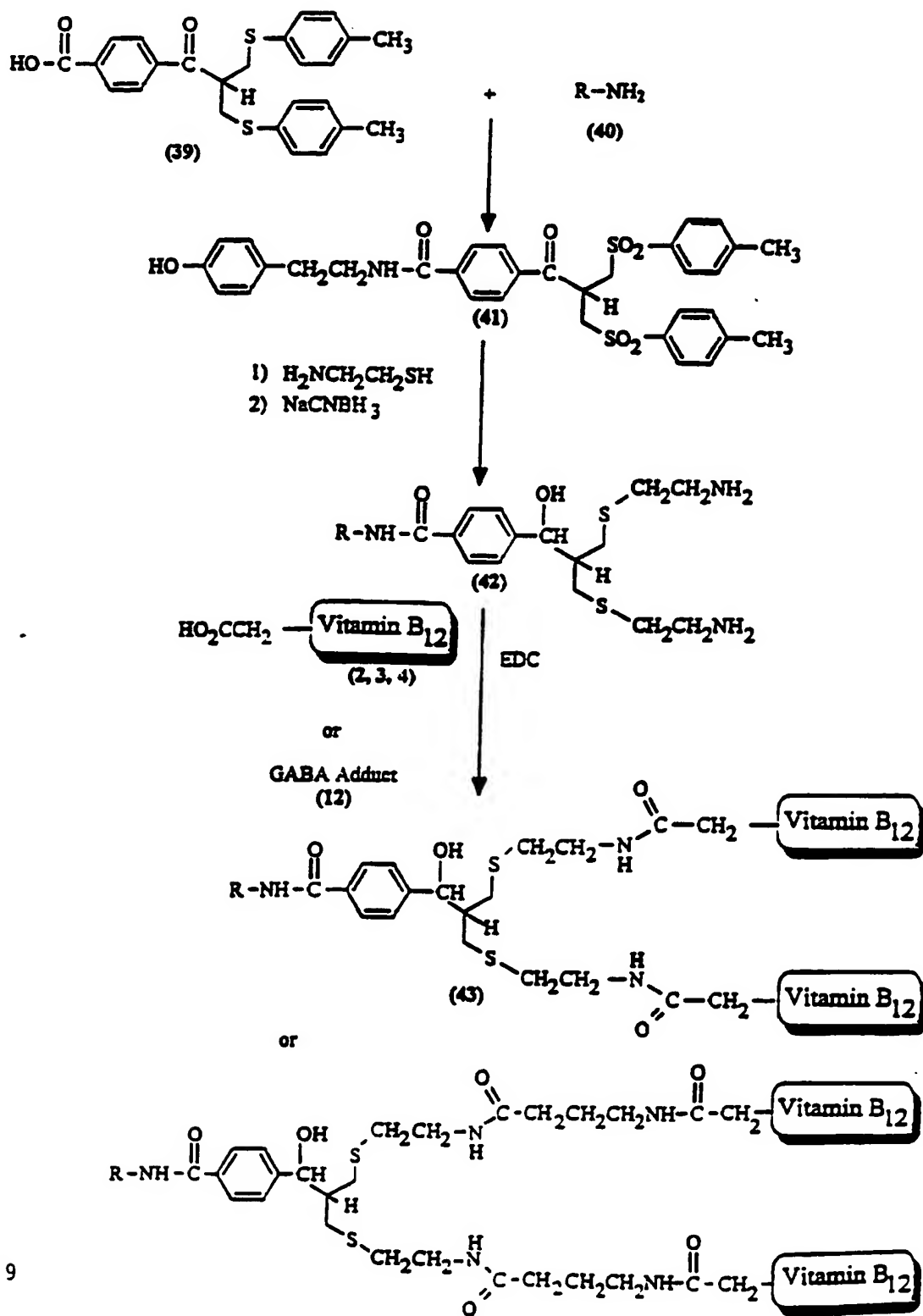


Fig. 9

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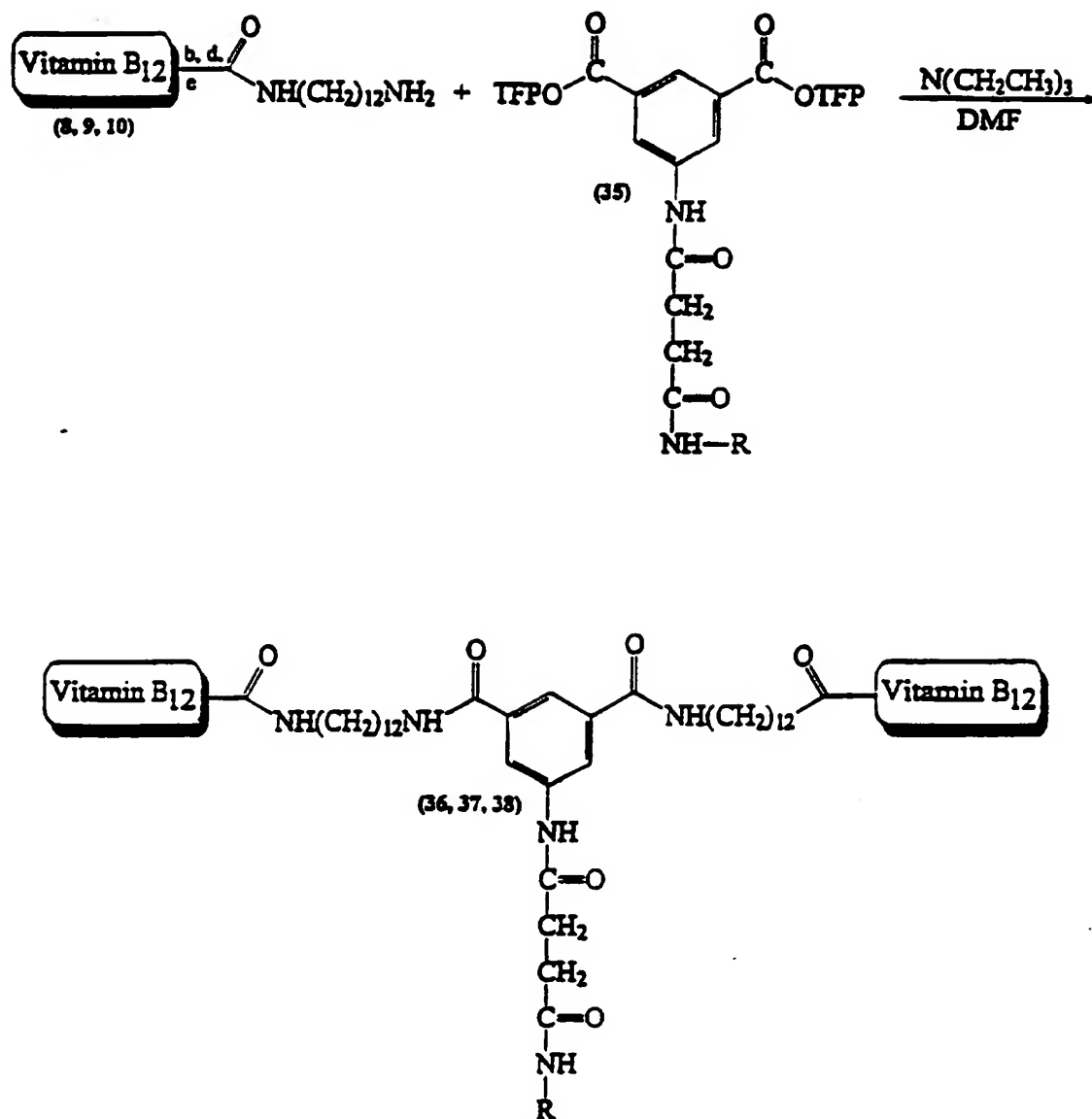
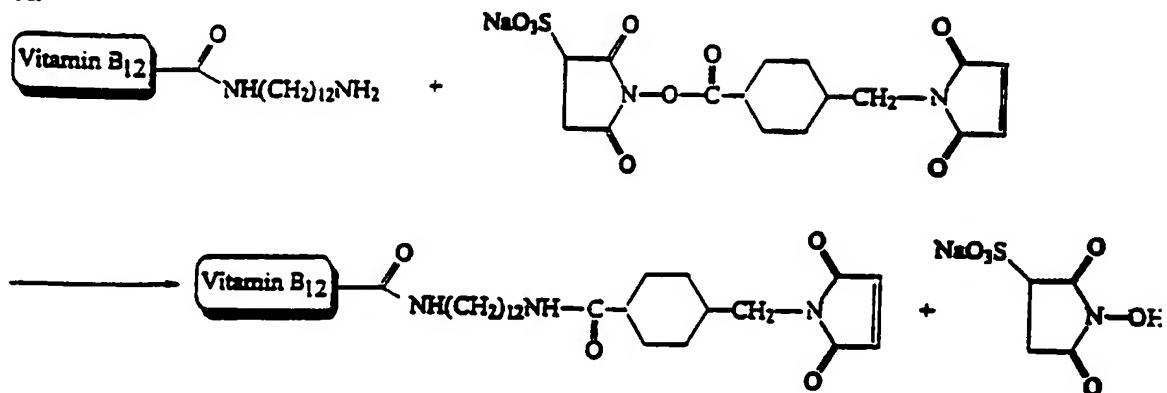


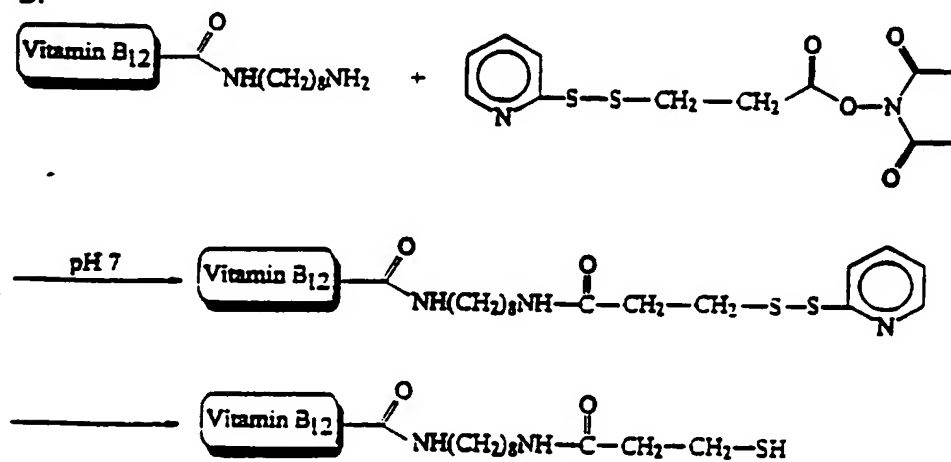
Fig. 10

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A.



B.



C.

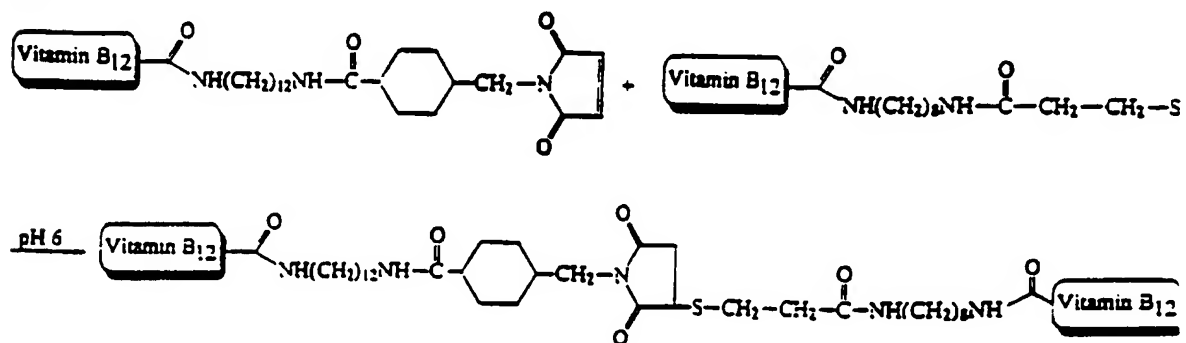


Fig. 11

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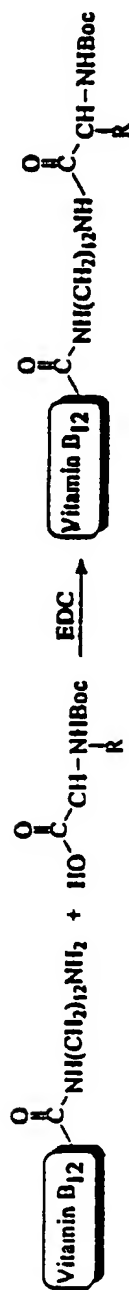


Fig. 12



Fig. 13

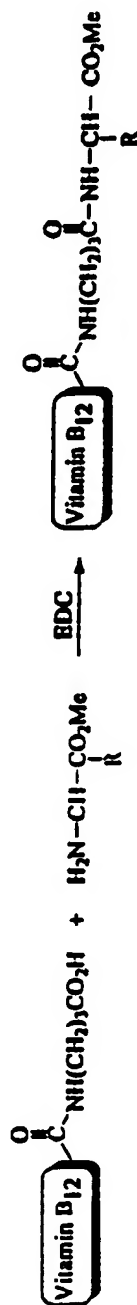


Fig. 14

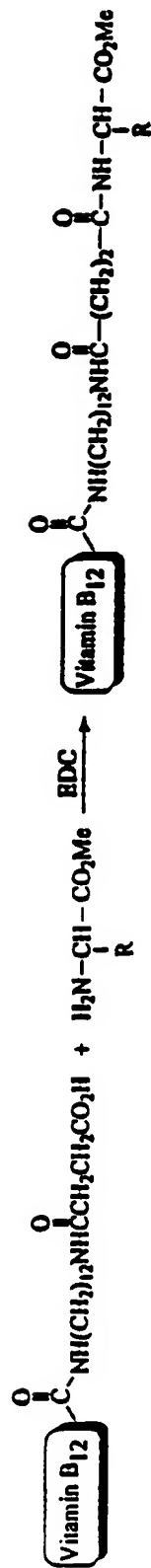


Fig. 15

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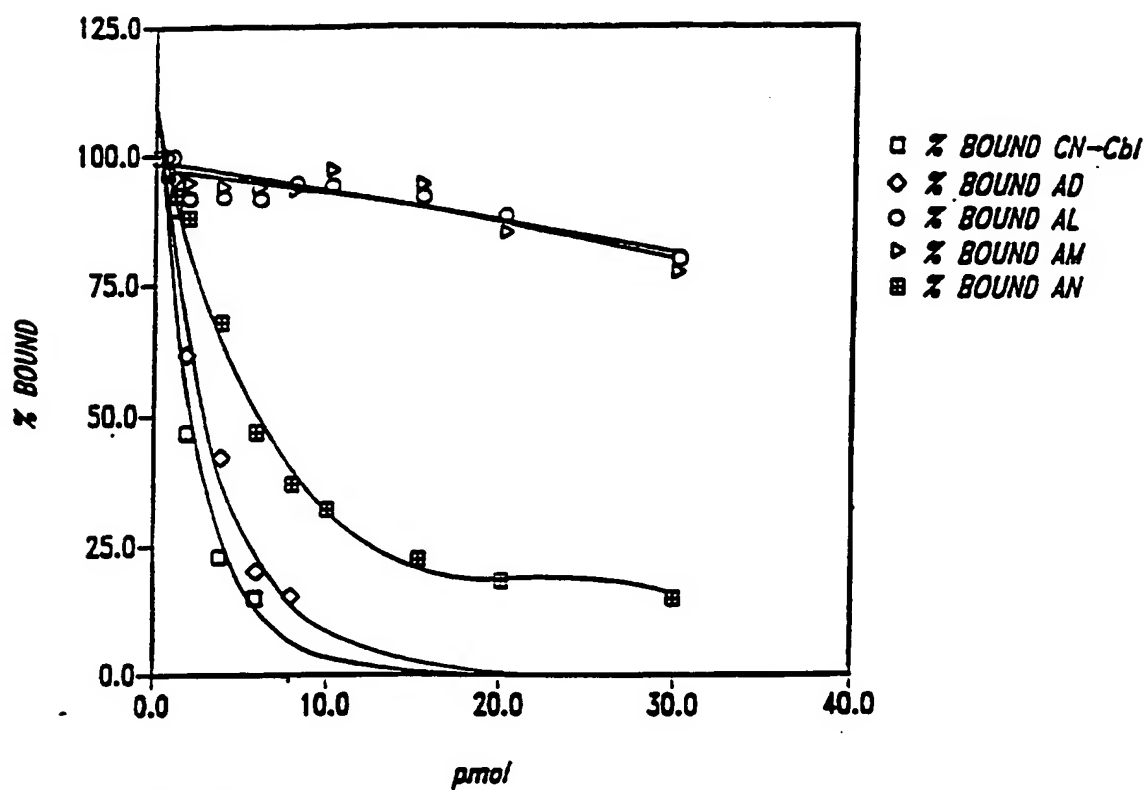


Fig. 16

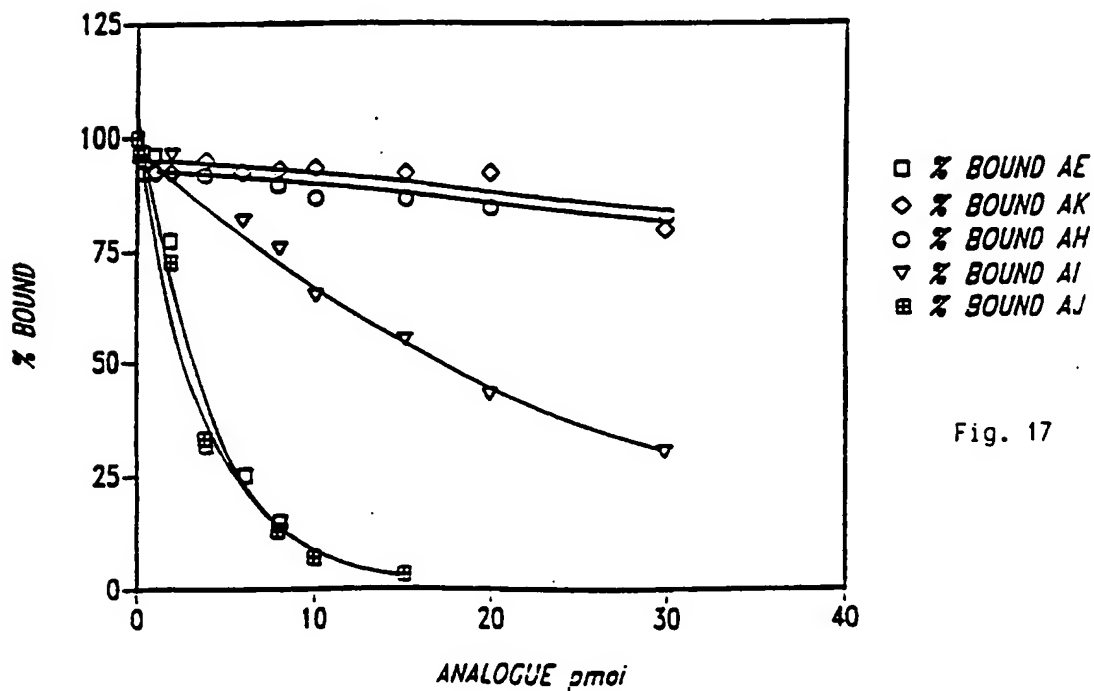


Fig. 17

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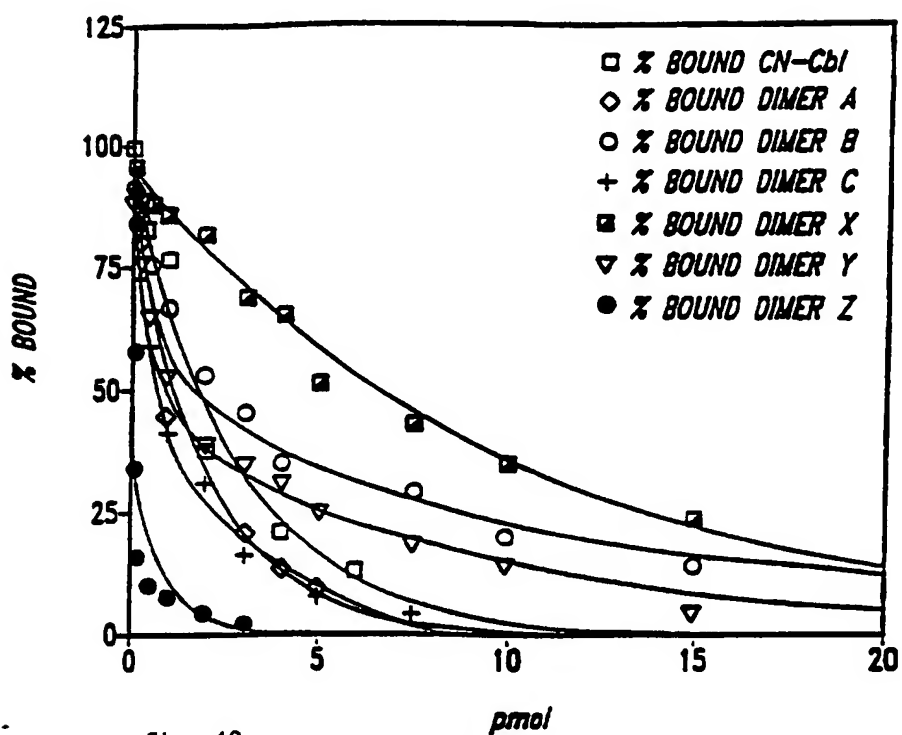


Fig. 18

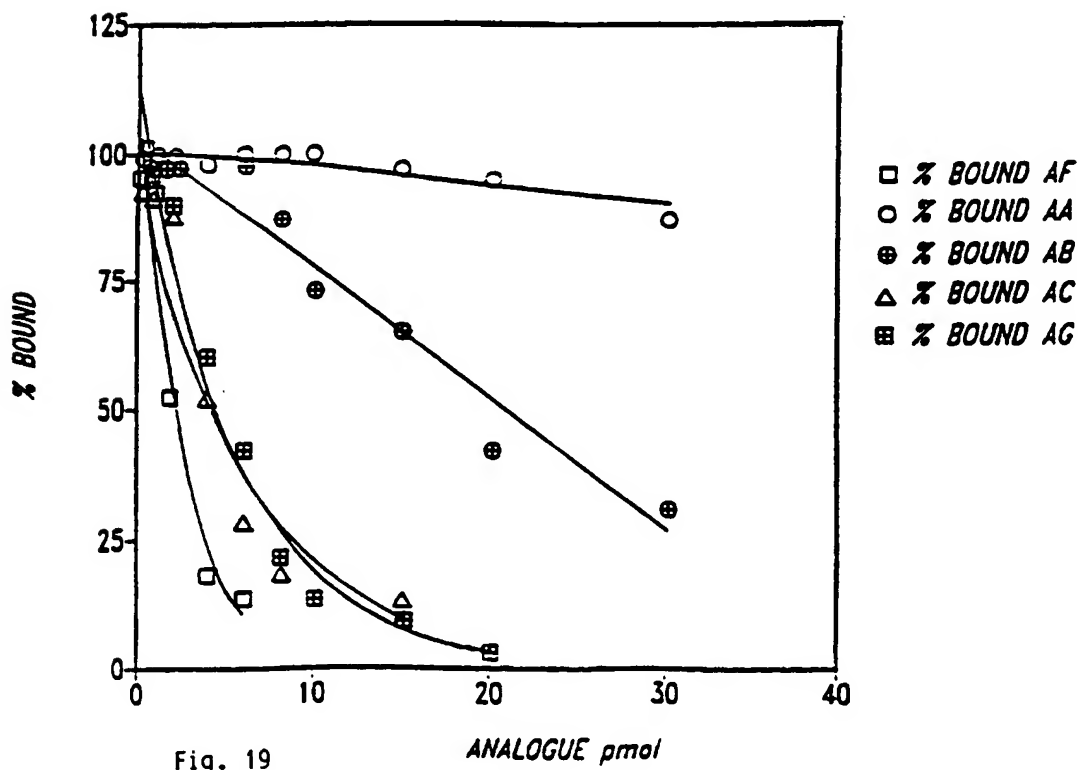


Fig. 19

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 96/16672

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C07H23/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C07H

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

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X	EP 0 599 325 A (BOEHRINGER MANNHEIM GMBH) 1 June 1994 see page 21, compound '15' (example 14) ---	23,24
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A	WO 93 23557 A (VITAMED INC) 25 November 1993 ---	1-31
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☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

14 February 1997

Date of mailing of the international search report

05 -03- 1997

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INTERNATIONAL SEARCH REPORT

Int'l. Application No
PCT/US 96/16672

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

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A	EP 0 378 203 A (BOEHRINGER MANNHEIM GMBH) 18 July 1990 ---	1-31
A	US 5 428 023 A (RUSSELL-JONES GREGORY J ET AL) 27 June 1995 ---	1-31
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P,X	BIOCONJUG. CHEM., vol. 7, 1996, pages 217-32, XP002025283 P. PATHARE ET AL.: "Synthesis of Cobalamin-Biotin Conjugates That Vary in the Position of Cobalamin Coupling" see the whole document -----	1-31

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INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 96/16672

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